

# Effects of Cholecalciferol and Phytase on Phytate Phosphorus Utilization in Laying Hens

A. MUSAPUOR, J. POURREZA†, A. SAMIE† AND H. MORADI SHAHRBABA‡<sup>1</sup>

*Department of Animal Science, Faculty of Agriculture, Shahid Bahonar Kerman University, Kerman, Iran*

*†Department of Animal Sciences, Faculty of Agriculture, Isfahan Technology University, Isfahan, Iran*

*‡Department of Animal Science, Faculty of Agriculture, University of Tehran, Karaj, Iran*

<sup>1</sup>Corresponding author's e-mail: [hmoradis@ut.ac.ir](mailto:hmoradis@ut.ac.ir)

## ABSTRACT

This experiment was conducted to study the effects of different levels of phytase, vitamin D<sub>3</sub>, calcium and available phosphorus on phytate phosphorus utilization in laying hens. Dietary phytase caused a significant ( $P<0.05$ ) increase in feed consumption, feed conversion ratio, tibia ash weight, tibia ash percentage, tibia phosphorus plasma phosphorus and phosphorus digestibility. However, dietary phytase caused a significant ( $P<0.05$ ) decrease in plasma alkaline phosphatase activity and excreta phosphorus percentage. Also phytase had no beneficial effect on egg shell quality traits. Increasing dietary vitamin D<sub>3</sub> caused a significant ( $P<0.05$ ) increase in feed consumption but a significant ( $P<0.05$ ) decrease in plasma alkaline phosphatase activity and no effect on the other traits. Available phosphorus levels had significant ( $P<0.05$ ) effect on tibia ash weight and tibia ash percentage. Reduction in dietary available phosphorus caused a significant ( $P<0.05$ ) decrease in feed consumption. Effect of dietary calcium was significant ( $P<0.05$ ) on tibia ash weight, feed consumption and plasma phosphorus. Interaction between phytase and calcium on tibia phosphorus, plasma calcium and excreta phosphorus were significant ( $P<0.05$ ). Interaction between phytase and available phosphorus on tibia phosphorus was significant ( $P<0.05$ ). Overall, it could be concluded that in low phosphorus diet which food consumption is low, phytase would increase food consumption as well as retention of phosphorus in bones. Also, the lower excreta of phosphorus by using phytase could decrease pollution.

**Key Words:** Phytase enzyme; Vitamin D<sub>3</sub>; Calcium; Phosphorus

## INTRODUCTION

A number of studies have indicated that supplementing laying diets with microbial phytase results in improved performance (Van der Klis *et al.*, 1996), particularly when dietary levels of non phytate P (NPP) are low (Gordon & Roland, 1997). The results of a number of research studies with laying hens have shown that a diet with 0.1-0.13% available phosphorus (AP) in the presence of 100 to 300 units phytase can result in comparable performance to the control group which were fed a normal level of 0.4-0.45% available phosphorus. It is well established that a high dietary level of calcium (Ca) reduces the activity of phytase (Ravindran *et al.*, 2000). However, a portion of the phytase benefit observed in poultry fed NPP deficient diets can be attributed not only directly to P, but also to the influence of P on improved Ca utilization. The results of experiments with laying hens have shown that the vitamin D<sub>3</sub> per se is effective in releasing some part of the phytate-phosphorus and make it available to the birds. Also, it has been shown that some additivity or synergistic effects exist between phytase and vitamin D<sub>3</sub> for increasing the availability of phytate phosphorus to the birds (Edwards, 1993). Reports of the effect of phytase enzyme in layer diets are fewer and have not fully investigated the interactions among AP, phytase, Ca and vitamin D<sub>3</sub>.

This paper reports on a laying trial in which hens were fed two levels of P, three levels of phytase enzyme, two levels of Ca and two levels of vitamin D<sub>3</sub> in a factorial arrangement designed to investigate interactions among these factors on production parameters of layers.

## MATERIALS AND METHODS

Twenty four diets were fed to Hy-line W-36 hens from 30 to 42 wks of age. The treatments consisted of a 3×2×2 factorial arrangement with three levels of Natuphos® phytase (0, 500, and 1000 FTU/kg diet), two levels of Ca (2.27 and 3.25%), two levels of NPP (0.175 and 0.25%) and two levels of vitamin D<sub>3</sub> (0 and 400 IU/kg diet). Each treatment was randomly assigned to four replicate cages for a total of 96 cages. Each cage was an experimental unit and contained four hens. The experimental diets were formulated to meet National Research Council (1994) nutrient requirement of laying hens (Table I). Records of daily egg production and weekly feed consumption were kept during the experiment. All the eggs produced during the last three days of every 28 day periods were saved for determination of egg weight and two eggs per cage were used to determine egg shells breaking strength, shell thickness, shell ash, and shell phosphorus. At 42 wks of age, four birds from each dietary treatment were killed and their

left tibia was removed. They were solvent-extracted to remove fat and then dried and ashed. At 42 wks of age, one bird was randomly selected from each pen, and blood samples were obtained for subsequent determination of minerals (Ca & P) and alkaline phosphatase (ALP) in serum. At 32 wks of age, chromic oxide was added to all diets as an analytical marker for P digestibility and fed for 10 days. Representative fecal samples were collected from each cage on the last day of Cr<sub>2</sub>O<sub>3</sub> feeding to determine digestibility of P at 42 wks. Diets and excreta were analyzed for P (AOAC, 1995) and chromium (Fenton & Fenton, 1979). Analysis of variance was performed on the data using the General Linear Models of SAS® software (SAS Institute, 1995).

## RESULTS AND DISCUSSION

Egg shell quality measurements were not consistently affected by the dietary treatments. Other researchers have reported mixed results of phytase supplementation on eggshell quality measurements (Gordon & Roland, 1998). Addition of 1000 FTU/kg phytase to the diet significantly increased feed intake and feed conversion of laying hens (Table II). There were no significant effects of enzyme supplementation on egg weight and egg production. The inclusion of phytase to the diets possibly increased feed intake by liberating the phytate phosphorus. These findings are in agreement with those of Keshavarz (2000). Supplementing diets with 1000 FTU/kg phytase resulted in an increased bone ash weight and percentage and bone mineral content (P). At the 0.175% NPP level, hens with phytase had higher bone phosphorus percent than when diets were not supplemented with phytase (Table II). The improvement in bone ash and bone phosphorus content associated with phytase supplementation could be attributed to phytate P liberation. There was a highly significant positive influence of phytase supplementation on P digestibility at 42 wks of age as expected (Table III). Other study (Um & Paik, 1999) found that supplementation phytase increased P retention by increasing the liberation of bound phytate P. As expected, phytase supplementation to diets increased ( $p < 0.05$ ) serum P level, and decreased serum ALP activity (Table III). The increase in serum P and decrease in ALP activity associated with the diets supplemented with phytase might be reflected the downregulation of this enzyme resulting from the increased availability of phosphorus. Vitamin D<sub>3</sub> supplementation (400 IU/kg) to basal diet was effective in improving the feed intake of hens (Table II). The results of this experiment were consistent with previous report that no beneficial effect on laying hen performance can be obtained by increasing the level of vitamin D<sub>3</sub> above the level that normally is used commercial practice (Keshavarz, 2000). Laying hen performance, bone ash measurements, serum Ca and ALP level and phosphorus digestibility were not affected by dietary Ca level (Table II and III). Hens fed low 2.27% Ca had significantly lower feed intake, serum P level and

**Table I. Ingredients and nutrient composition of experimental diets**

Ingredient (%)	0.25%AP, 3.25% Ca	0.25%AP, 2.27%Ca	0.175%AP, 3.25%Ca	0.175%AP, 2.27%CA
Corn	65.86	72.27	66.30	72.48
Soybean meal	21.37	20.13	21.28	20.09
Fat	3.1	0.50	2.92	0.50
Oyster shell	7.95	5.39	8.18	5.62
Dicalciumphosphate	0.70	0.71	0.30	0.31
Vitamin premix <sup>1</sup>	0.30	0.30	0.30	0.30
Mineral premix <sup>2</sup>	0.30	0.30	0.30	0.30
Salt	0.35	0.34	0.35	0.34
DL-Methionine	0.07	0.06	0.07	0.06
Nutrient Composition				
ME, kcal/kg	2900	2905	2900	2911
Protein (%)	15	15	15	15
Calcium (%)	3.25	2.27	3.25	2.275
Nonphytate P (%)	0.25	0.25	0.175	0.175
Sodium (%)	0.15	0.15	0.15	0.15
Arg. (%)	0.921	0.906	0.92	0.906
Lys. (%)	0.746	0.729	0.745	0.728
TSAAs (%)	0.58	0.58	0.58	0.58
Try. (%)	0.197	0.192	0.197	0.192

<sup>1</sup>Vitamin mix supplied the following per kilogram of diet: vitamin A, 10000 IU; vitamin D<sub>3</sub>, 500 IU; vitamin E, 10 IU; B1, 2.2 mg; B2, 4 mg; B3, 8 mg; B6, 2 mg; B9, 0.56 mg; B12, 15 mg; H2, 0.15 mg; <sup>2</sup>Mineral mix supplied the following per kilogram of diet: Mn, 800 mg; Zn, 60 mg; Fe, 50 mg; Cu, 5 mg; Co, 0.1 mg; I, 1 mg; Se, 0.1 mg; Choline chloride, 200 mg

**Table II. Influence of phytase, vitamin D<sub>3</sub>, P and Ca levels on production traits and bone ash measurements**

Diet	Egg weight (g)	Egg prod. (%)	Feed intake (g/h/d)	FCR (g/g)	Bone ash (%)	Bone phos. (%)
Phytase(FTU/kg)						
0	57.02	83.40	95.28c	2.01b	61.55b	7.87b
500	57.32	83.72	96.61b	2.02ab	62.35ab	8.82a
1000	57.37	83.51	99.46a	2.08a	62.54a	8.60a
Vit. D <sub>3</sub> (IU/kg)						
0	57.02	84.25	96.74b	2.02	61.99	8.23
400	57.45	82.84	97.50a	2.05	62.30	8.63
Calcium (%)						
2.27	57.20	82.61	94.88b	2.10	62.19	8.45
3.25	57.27	82.48	99.35a	2.11	62.10	8.41
Phosphorus (%)						
0.175	57.09	83.40	96.32b	2.03	61.75b	8.45
0.25	57.39	83.69	97.91a	2.05	62.54a	8.41
Pooled SEM	4.32	16.18	2.08	0.015	3.23	1.71

FCR= Feed conversion ratio; <sup>abc</sup>means with in columns with no common superscript differ ( $p < 0.05$ )

**Table III. Influence of phytase, vitamin D<sub>3</sub>, P and Ca levels on production traits and bone ash measurements**

Diet	Plasma Ca (mg/100ml)	Plasma P (mg/100ml)	AP (U/L)	Excreta Ash (%)	Excreta P (%)	PD (%)
Phytase(FTU/kg)						
0	17.34	5.55b	632.19a	37.63	1.13a	34.59a
500	18.16	6.88a	506.81b	39.09	0.97b	42.88b
1000	17.30	6.63a	544.50b	37.69	0.94b	48.12c
Vit. D <sub>3</sub> (IU/kg)						
0	17.72	6.26	605.21a	39.16	1.01	41.98
400	17.48	6.37	517.13b	37.11	1.01	41.76
Calcium (%)						
2.27	17.67	6.06b	544.58	37.58	0.87b	42.24
3.25	17.53	6.57a	577.75	38.68	1.15a	41.49
Phosphorus (%)						
0.175	17.76	6.35	600.75a	37.14	1.05	40.36
0.25	17.44	6.29	521.58b	39.13	0.97	43.04
Pooled SEM	2.89	1.02	-	-	0.043	-

AP= Alkaline phosphatase; PD= Phosphorus digestibility; <sup>abc</sup>means with in columns with no common superscript differ ( $p < 0.05$ )

excreta phosphorus percent than hen fed diets with high level of Ca (Table II and III). Although the extent of Ca withdrawal practiced in this study is not at this point recommended commercially, but hens consuming the 2.27% Ca diet performed as well as hen fed diets containing higher level of Ca. In our current study, the use of a NPP regimen of 0.175% which was used for the age periods of 30-42 wks of age was adequate to support all the production traits, in spite of negative effect on feed intake and bone ash measurements at 0.175% NPP. Apparently, under condition of 0.175% AP diets, the P could be utilized for production and was not deposited in the bones. From our results, it is concluded that supplemental phytase has beneficial effects on the performance of laying hen. It is recommended that laying hen diets be formulated to provide 0.175% NPP, 2.75% Ca with supplemental phytase to hen early in the production cycle. Microbial phytase supplementation with low-Ca, low-P diet can decrease the level of phytate P excretion in the manure and limit soil and water contamination.

## REFERENCES

- AOAC, 1995. Animal Feeds. *Official Methods of Analysis. Association of Official Analytical Chemists*, p. 27.
- Edwards, H.M. Jr., 1993. Dietary 1, 25-dihydroxycholecalciferol supplementation increases natural phytate phosphorus utilization in chickens. *J. Nut.*, 123: 567-73
- Fenton, T.W. and M. Fenton, 1979. Determination of chromic oxide in feed and feces. *Canadian J. Anim. Sci.*, 58: 631-4
- Gordon, R.W. and D.A. Roland, 1997. Performance of commercial laying hens fed various phosphorus levels, with and without supplemental phytase. *Poultry Sci.*, 76: 1172-7
- Gordon, R.W. and D.A. Roland, 1998. Influence of supplemental phytase on calcium and phosphorus utilization in laying hens. *Poultry Sci.*, 77: 290-4
- Keshavarz, K., 2000. Non phytate phosphorous requirement of laying hens with and without phytase on a phase feeding program. *Poultry Sci.*, 78: 699-706
- National Research Council, 1994 *Nutrient Requirements of Poultry*, 9<sup>th</sup> Revised ed. National Academy of Sciences, Washington, DC
- Ravindran, V., S. Cabahug, P.H. Selle and W.L. Bryden, 2000. Response of broiler chickens to microbial phytase supplementation as influenced by dietary phytic acid and non-phytate phosphorus levels. II. Effects on apparent metabolisable energy, nutrient digestibility and nutrient retention. *British Poultry Sci.*, 41: 193-200
- Sas Institute, 1995. *SAS/STAT® User's Guide: Statistics*. 1996 ed. SAS Institute, Inc., Cary, NC
- Um, J.S. and I.K. Paik, 1999. Effects of microbial phytase supplementation on egg production, eggshell quality, and mineral retention of laying hens fed different levels of phosphorus. *Poultry Sci.*, 78: 75-9
- Van der Klis, J.D., H.A.J. Versteegh and P.C.M. Simons, 1996. Natuphos in laying hen nutrition. In: *BASF Technical Symposium Phosphorus and Calcium Management in layers*, pp. 71-82. Atlanta, GA, January 23, 1996.

(Received 26 February 2005; Accepted 12 June 2005)