In Vitro Evaluation of Ovicidal and Larvicidal Effects of Culture Filtrate of Verticillium chlamydosporium Against Meloidogyne javanica

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ABSTRACT

Effects of culture filtrate of *Verticillium chlamydosporium* were evaluated on mortality of larvae and hatching of eggs of *Meloidogyne javanica*. Mortality of *M. javanica* larvae was found to be directly proportional to concentrations of culture filtrate of *V. chlamydosporium* and duration of exposure. Maximum mortality of larvae (65.5%) was observed in 100% concentration of culture filtrate after 72 h followed by 80% concentration which caused 53.25% mortality. Similarly different dilutions of culture filtrate significantly inhibited hatching *of M. javanica* eggs. Larval emergence was, however, inversely proportional to filtrate concentrations. Significant lower hatching was observed in 100% concentration.

Key Words: Culture filtrate; Verticillium chlamydosporium; Mortality; Hatching; Meloidogyne javanica

INTRODUCTION

Controlling any soil inhabitant pathogen in a crop is difficult and root-knot disease caused by *Meloidogyne* spp. is no exception. Nematicides may play a major role in crop protection and production threatened by diseases when other practical and economical measures are lacking. In recent years the ban on certain nematicides due to health hazards and environmental concerns has prompted the search for alternatives to manage nematode damage.

Of various approaches that could be adopted for the control of nematodes, biological control is receiving increased interest among nematologists. Biological control includes the use of predaceous or parasitic organisms such as fungi, bacteria, protozoa, viruses, nematodes and mites etc. Nematode destroying fungi are widespread in soil and consistently associated with the bio-control of nematodes in nature. These fungi kill and destroy nematodes by prey catching devices such as sticky branches, networks, knobs, constricting and non-constricting rings; by producing toxic substances and adhesive spores and by colonizing nematode reproductive structures.

Verticillium chlamydosporium is a wide spread fungus that proliferates in the rhizosphere and parasitizes females and eggs of cyst and root-knot nematodes. A lot of literature is available on *V.chlamydosporium* which suggests that the fungus has potential as a biocontrol agent for cyst and root-knot nematodes (Kerry et al., 1984; De Leij & Kerry, 1991; Mousa et al., 1995). Since fungi and nematodes occur concomitantly in the rhizosphere, the toxic metabolites naturally produced by these fungi may be responsible for keeping low levels of nematode population. Adverse effects of such substances present in the culture filtrates of several

soil borne fungi on hatching and mortality of root-knot nematodes have been studied by many workers (Desai *et al.*, 1972; Cayrol *et al.*, 1989; Saifullah, 1996). The present study was, however, conducted to evaluate the ovicidal and larvicidal effects of culture filtrate of *V. chlamydosporium* against *M. javanica*.

MATERIALS AND METHODS

Culture filtrates were obtained by growing *V. chlamydosporium* on Potato Dextrose broth. The medium (100 mL in 250 mL flasks) was autoclaved at 15 psi for 15 minutes. Each flask was then inoculated with four scoops of 5 mm diameter of the fungus from an actively growing culture of the fungus on corn meal agar under sterilized conditions and incubated at 25°C for 15 days. Some flasks were inoculated with the scoops of the medium only. At the end of incubation period, the cultures were passed through Whatman filter paper No. 1 to remove the mycelial mats. Filtrates thus obtained were designated as 100% concentration. Further concentrations (20, 40, 60, and 80%) were made by adding requisite amount of sterilized distilled water. The filtrates were stored at 4°C until use.

The population of *Meloidogyne javanica* used in the experiment was already maintained on the roots of tomato plants in the glasshouse of the Department of Plant Pathology, University of Agriculture, Faisalabad and mass cultured on tomato cv. Money maker. Eggs were isolated and nematodes were extracted (Hussey & Barker, 1973).

To investigate the effect of different concentrations of the culture filtrate of *V. chlamydosporium* on larval hatching of *M. javanica*, 5 mL of culture filtrate from each concentration was pipetted into 5-cm-dia. Petri dishes.

Three uniformed-sized egg masses of *M. javanica* were hand picked and placed into each dish. Egg masses placed in distilled water served as control. Each treatment was replicated five times. All dishes were maintained at 25°C. Data on egg hatching were recorded after 24, 48, and 72 h. Fresh fungal filtrate was replaced after each data. The same procedure was repeated using potato dextrose broth in order to compare the effect of culture filtrate with that of the medium.

Similarly, to study the effect of different concentrations of the culture filtrate on larval mortality, 5 ml of the culture filtrate from each concentration was poured into each Petri dish and about 80 freshly hatched second-stage juveniles of *M.javanica* in 0.2 mL distilled water were added to each Petri dish. Juveniles added in distilled water served as control. Each treatment was replicated five times. Number of dead (unmoved) larvae in each Petri dish were counted after 24, 48 and 72 h and their percentages were calculated. In order to compare the effect of culture filtrate with that of the medium, the same procedure was followed using the medium. All the data collected were analyzed statistically by using Analysis of Variance and Regression Analysis (Muhammad, 1991).

RESULTS

Effect of culture filtrates on egg hatching of M. javanica.

The analysis of variance regarding effect of culture filtrates on hatching of M.javanica juveniles showed significant interaction (P < 0.01) between filtrates and concentrations (F X C), filtrates and time interval (F X T), concentrations and time(C X T) and among filtrates, their concentrations and time (F X C X T).

Data presented in Table I showed that mean number of juveniles hatched in medium filtrate was significantly higher than those hatched in the culture filtrate. As mean hatching observed in different concentrations of the medium filtrate was found to be statistically similar to that observed in distilled water (0% concentration), it was concluded that medium itself did not show ovistatic or ovicidal property.

Larval emergence was found to be inversely proportional to filtrate concentration and time interval. Different dilutions of the culture filtrate significantly inhibited hatching of *M. javanica* eggs. Maximum inhibition in hatching was observed in 100% and 80% concentrations of the culture filtrate followed by 60%, which, however, was statistically not different from the highest concentrations. Significantly lesser juveniles emerged after 72 h as compared to 48 and 24 h interval.

With an increase in the concentration of culture filtrate there was a significant decrease in hatching. These relationships are shown by regression equations given below.

Effect of culture filtrates on larval mortality of *M. javanica.* The analysis of variance regarding effect of culture filtrate on percent mortality of *M. javanica* juveniles showed significant interaction (P< 0.01) between filtrates and concentrations (F X C), filtrates and time interval (F X T), concentrations and time (C X T) and among filtrates, their concentrations and time (F X C X T).

It is apparent from the perusal of the data presented in Table-II that mean percent mortality in culture filtrate of the fungus differed significantly from that in the medium filtrate and it is concluded that medium itself had no nematostatic or nematicidal effect as the mean mortality observed in different concentrations of the medium filtrate was statistically similar to the mortality observed in distilled water (0% concentration).

Percent juvenile mortality in the culture filtrates of the fungus was found to be directly proportional to the concentrations of the culture filtrate and the duration of exposure. Maximum mortality (50.92%) was observed in 100% concentration of the fungal filtrate whereas it was minimum in 20% concentration which was statistically similar to that in 0% concentration (distilled water).

The mean percent mortality in fungal filtrate after 24 hours was 13.25% which increased significantly to 20.42% after 48 hours and finally reached to 28.42% after 72 h.

Mean individual mortality (65.5%) was observed in 100% concentration of culture filtrate after 72 h followed by 80% concentration which caused 53.25% mortality. Mean percent mortalities in medium filtrate and culture filtrate at different concentrations and durations are given in Table II.

A progressive increase in the concentrations of the culture filtrate resulted in increase in the percent mortality of juveniles. The relationships between the concentrations and percent mortality at different time intervals are given by regression equations

DISCUSSION

Adverse effects of culture filtrates of several fungi on hatching and mortality of root-knot nematodes have been reported by many workers (Cayrol *et al.*, 1989; Saifullah, 1996 & Zaki, 1999).

Nematicidal action of culture filtrates of *V.chlamydosporium* against *M.javanica* may be attributed to the production of certain enzymes (Webb *et al.*, 1972; Segers *et al.*, 1994) and toxins like Verticillin A, B and C (Minato *et al.*, 1973) which help in weakening and dissolving the barriers of its hosts. Species of *Aspergillus, Penicillium, Talaromyces, Curvularia* and *Aternaria* are known to produce toxins and antibiotics like aflatoxin, penicillin, vermiculin, vermicillin, talaron, vermistatin, viridin, fusaric acid, rhizopin, lilacinin, leucinostatin, P-168 and phytoalternarin (Nafe-Roth, 1972; Arai *et al.*, 1973; Das

Table I. Effect of culture filtrate of V. chlamydosporium on hatching of M. javanica eggs

Filtrates	Concentration (%)	Hatching after			Mean
		24 hours	48 hours	72 hours	_
	0	218.8 a	60.6 e	43.6 fgh	107.7 A
Medium	20	209.4 ab	57.0 e	37.0 h	101.1 A
Filtrate	40	205.0 b	58.8 e	40.8 gh	101.5 A
(No fungus)	60	212.2 ab	57.4 e	40.8 gh	103.5 A
	80	207.2 b	56.4 e	39.4 h	101.0 A
	100	213.6 ab	55.0 e	37.6 h	102.1 A
		211.03 A	57.53 C	39.87 D	102.81 A
	0	218.8 a	60.6 e	43.6 fgh	107.70 A
	20	99.8 c	20.8 i	6.4 jk	42.33 B
Culture	40	77.4 d	15.2 ij	5.0 jk	32.53 C
Filtrate	60	61.4 e	9.6 ijk	5.2 jk	25.40 CD
	80	54.8 ef	4.6 jk	2.6 k	20.67 D
	100	51.8 efg	8.6 jk	1.8 k	20.73 D
		94.00 B	19.90 E	10.77 F	41.56 B

LSD values at 0.05 for Filtrates (F) = 13.35; Concentrations (C) = 5.45; Time (T) = 3.29; FX C = 7.71 FX T = 4.65; CX T = 8.04; FX CX T = 11.39

Table II. Effect of culture filtrate of V. chlamydosporium on mortality of M. javanica

Filtrates	Concentration (%)	% Mortality			Mean
		24 hours	48 hours	72 hours	_
	0	0.00 n	0.25 mn	2.25 ijkl	0.83 E
Medium	20	0.00 n	0.50 lmn	3.50 hi	1.33 E
Filtrate	40	0.00 n	0.00 n	1.75 ijklmn	0.58 E
(No fungus)	60	0.50 lmn	0.75 lmn	1.25 jklmn	0.83 E
	80	0.50 lmn	1.00 klmn	1.50 jklmn	1.00 E
	100	0.50 lmn	1.00 klmn	2.00 ijklmn	1.17 E
		0.25 E	0.58 E	2.04 D	0.96 B
	0	0.00 n	0.25 mn	2.25 ijkl	0.83 E
	20	1.50 jklmn	2.75 hijk	4.25 h	2.83 E
Culture	40	3.00 hij	4.25 h	15.00 g	7.42 D
Filtrate	60	13.75 g	24.50 f	30.25 e	22.83 C
	80	25.50 f	39.25 c	53.25 b	39.33 B
	100	35.75 d	51.50 b	65.50 a	50.92 A
		13.25 C	20.42 B	28.42 A	20.69 A

LSD values at 0.05 for Filtrates (F) = 4.58; Concentrations (C) = 2.24; Time (T) = 0.55; F X C = 3.17; F X T = 0.78; C X T = 1.36; F X C X T = 1.92

& Pal, 1974; Wheeler, 1975; Fuska *et al.*, 1979; Isogai *et al.*, 1981; Agarwal & Bisen, 1984; Ghewande *et at.*, 1984; Mikamy *et al.*, 1989). There are some fungi which produce appreciable amount of oxalic acid in the culture medium (Franke & de Boer, 1961) which killed root-knot nematode larvae (Desai *et al.*, 1972).

These toxic chemicals may affect the phytonematodes in different ways. Cayrol *et al.* (1989) suggested that the toxic metabolites have a neurotropic action upon the nervous receptors of the nematodes. Saifullah (1996) found that culture filtrate of *V. chlamydosporium* caused 100% mortality of potato cyst nematode males and *Panagrellus redivivus* and a strong contraction of the nematode muscles causing the spicules to be pushed out. Nematodes ruptured in the middle probably because of the digestive enzymes, the fungus released into the medium. The enzymes of the fungus were not able to make the cuticles of nematodes permeable. The toxic substances might, therefore, have entered through the natural pores.

A number of nematophagous fungi are known to have proteolytic and chitinolytic activities which cause alteration in eggs' cuticular structure, changes in egg shell permeability or cause perforations in the cuticle which allows seepage of toxic metabolites into the eggs and cause physiological disorders (Webb et al., 1972; Jatala et al., 1985; Lopez-Llorca, 1990). These factors may have important role in the inhibition of egg hatch. V. chlamydosporium produces a subtilisin, which breaks down the outer membrane of the egg shell and exposes the chitin layer of root-knot nematode eggs (Segers et al., 1994). Lopez-Llorca and Robertson (1992) reported the secretion of a serine protease designated P-32, produced by V. chlamydosporium during infection of nematode eggs. Similar enzyme was also produced by nematode trapping fungus, Arthrobotrys oligospora (Tunlid et al., 1994). It has also been reported that serine protease from P. lilacinus liquid culture plays an important role in the penetration of the fungus through egg shell of nematodes as incubation of purified protease with nematode eggs significantly influenced their development (Bonants et al., 1995).

It is suggested that the study should be extended to characterize the substances produced in the culture filtrate of *V. chlamydosporium*.

REFERENCES

- Agarwal, G.R. and P.S. Bisen, 1984. Toxin production by *Curvularia lunata var. aeria* causing leaf blight in French bean (*Phaseolus vulgaris* L.). *J. Ind. Bot. Soc.*, 63: 292–7
- Arai, T., Y. Mikamy, K. Fukushima, T. Utsumi and K. Kazawa, 1973. A new antibiotic, leucostatin, derived from *Pencillium lilacinum*. J. Anitib. Tokyo, 26: 157–61
- Bonants, P.J.M., P.F.L. Fitters, H. Thijs, E. Beldar, C. Waalwijk and J.W.D.M. Henfling, 1995. A basic serine protease from Paecilomyces lilacinus with biological activity against Meloidogyne hapla eggs. Microbiol., 141: 775–84
- Cayrol, J.C., C. Djian and L. Pijarowski, 1989. Study of the nematocidal properties of the culture filtrate of the nematophagous fungus Paecilomyces lilacinus. Rev. de Nematol., 12: 331–6
- Das, C.R. and A. Pal, 1974. Rhizopin: An antibiotic produced by *Rhizopus nigricans*. *Indian Phytopathol.*, 27: 33–6
- De Leij, F.A. and B.R. Kerry, 1991. The nematophagous fungus Verticillium chlamydosporium as a potential biocontrol agent for Meloidogyne arenaria. Rev. de Nematol., 14: 157–64
- Desai, M.V., H.M. Shah and S.N. Pillai, 1972. Effect of Aspergillus niger on root-knot nematode, Meloidogyne incognita. Ind. J. Nematol., 2: 210–4
- Franke, W. and W. De Boer, 1961. Oxidative enzymes from molds and yeasts. II. The enzymic degradation of glycolic acids by Aspergillus niger. Z. Physiol. Chem., 31: 70–89
- Fuska, J., A. Fuskova and P. Nemec, 1979. Vermistatin, an antibiotic with cytotoxic effects produced from *Penicillium vermiculatum*. *Biologia* (*Bratislava*), 34: 735–40
- Ghewande, M.P., R.N. Pandey, A.K. Shukla and D.P. Mishra, 1984. Toxicity of culture filtrates of Aspergillus flavus on germination of seeds and seedlings growth of groundnut. Ind. Bot. Rep., 3: 107–11
- Hussey, R.S. and K.R. Barker, 1973. A comparison of methods of collecting inoculum of *Meloidogyne* spp. including a new technique. *Pl. Dis. Rep.*, 57: 1025–8
- Isogai, A., A. Suzuki, S. Higashikawa, S. Kuyama and S. Tamura, 1981.
 Isolation and biological activity of a peptidal antibiotic P-168. Agri. Biol. Chem., 45: 1023–4
- Jatala, P., J. Franco, A. Gonzales and C.M. O'Hara, 1985. Hatching stimulation and inhibition of *Globodera pallida* eggs by enzymatic and exopathic toxic compounds of some biocontrol fungi. *J. Nematol.*, 17: 501

- Kerry, B.R., A. Simon and A.D. Rovira, 1984. Observations on the introduction of *Verticillium chlamydosoprium* and other parasitic fungi into soil for control of the cereal cyst-nematode, *Heterodera* avenae. Ann. Appl. Biol., 105: 509–16
- Lopez-Llorca, L.V., 1990. Purification and properties of extracellular proteases produced by the nematophagous fungus Verticillium suchlasporium. Canadian J. Microbiol., 36: 530–7
- Lopez-Llorca, L.V. and W.M. Robertson, 1992. Immuno-cytochemical localization of a 32-k Da protease from the nematophagous fungus Verticillium suchlasporium in infected nematode eggs. Exp. Mycol., 16: 261–7
- Mikamy, Y., K. Yazawa, K. Fukushima, T. Arai, S. Udagawa and R.A. Samson, 1989. Paecilotoxin production in clinical or terrestrial isolates of *Paecilomyces lilacinus* strains. *Mycopath.*, 108: 195–9
- Minato, H., M. Matsumoto and T. Katayama, 1973. Studies on the metabolites of *Verticillium* sp. structure of verticillins A, B and C. *Chem. Comm.*,: 1819–25
- Mousa, E.M., A.M. Basiony and M.E. Mahdy, 1995.Control of Meloidogyne javanica by Verticillium chlamydosporium on nematodes. Afro-Asian J. Nematol., 5: 113–5
- Muhammad, F., 1991. Statistical Methods and Data Analysis. Chenab Publications, Faisalabad, Pakistan
- Nafe-Roth, S., 1972. Production and bioassay of phytotoxins. In: Phytotoxins in Plant Diseases. Wood, R.S.K., A. Ballio and A. Granite (eds.). Academic Press, New York.
- Saifullah, 1996. Nematicidal and nematostatic effect of cell-free culture filtrates of *Verticillium chlamydosporium* Goddard *in vitro*. *Afro-Asian J. Nematol.*, 6: 32–5
- Segers, R., T.M. Butt, B.R. Kerry and J.F. Peberdy, 1994. The nematophagous fungus Verticillium chlamydosporium produces a chymoelastase-like protease that hydrolyses host nematode proteins in situ. Microbiol., 140: 2715–23
- Tunlid, A., S.E.K.B. Rosen and L. Rask, 1994. Purification and characterization of an extracellular serine protease from the nematode-trapping fungus Arthrobotrys oligospora. Microbiol., 14: 1687–95
- Webb, H.M., A. Ghafoor and J.B. Heale, 1972. Protein and enzyme patterns in strains of *Verticillium. Trans. Brit. Mycol.*, Soc. 59: 393–402
- Wheeler, H., 1975. *Plant Pathogens. P. 106.* Springer-Verlag Berlin. Heidleberg, New York.
- Zaki, M.J., 1999. Effect of fungal culture filtrates on mortality and hatching of Meloidogyne javanica. Pakistan J. Biol. Sci., 2: 161–3

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