



Review Article

Beneficial Effects of Mycorrhizal Association for Crop Production in the Tropics - A Review

Umme Aminun Naher¹, Radziah Othman^{1,2*} and Qurban Ali Panhwar²

¹Institute of Tropical Agriculture, Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia

²Department of Land Management, Universiti Putra Malaysia, 43400 Serdang, Selangor, Malaysia

*For correspondence: radziah@putra.edu.my

Abstract

Mycorrhiza plays a significant role in sustainable agriculture and has mutualistic symbiotic association with plant roots. There are several species of mycorrhiza and among the species studied *Glomus mosseae* is well known to colonize several vegetables, fruits, cereals and industrial crops. This paper highlighted the symbioses and beneficial effects of arbuscular mycorrhiza fungi (AMF) with tomato (*Solanum lycopersicum*), brinjal (*S. melongena*), potato (*S. tuberosum*), lady's finger (*Abelmoschus esculentus*), cucumber (*Cucumis sativus*), bean (*Phaseolus vulgaris*), pepper (*Capsicum annuum*), wheat (*Triticum aestivum*), aerobic rice (*Oryza sativa*), corn (*Zea mays*), durian (*Durio zibethinus*), rambutan (*Nephelium lappaceum*), pineapple (*Ananas comosus*), citrus (*Citrus sinensis*), banana (*Musa acuminata*), oil palm (*Elaeis guineensis*) and kenaf (*Hibiscus cannabinus*). Application of AMF increased nutrient uptake, water relations and perform as bio-protectants against pathogens and toxic stresses. In order to further improve their benefits, it is necessary to ensure the management practices comprising low tillage, abridged use of chemical fertilizers, especially the phosphatic fertilizers. © 2013 Friends Science Publishers

Keywords: Mycorrhizae association; Growth; Infection; Inoculation; Symbiotic

Abbreviations: Arbuscular mycorrhizal fungi (AMF); beach ridges interspersed with swales (BRIS); ground magnesium limestone (GML); plant growth promoting rhizobacteria (PGPR); vesicular-arbuscular mycorrhiza (VAM)

Introduction

Most of the plants form symbiotic relationship with a group of fungi called mycorrhiza, which function as a bridge for the flow of energy and matter between plants and soils (Cardon and Whitbeck, 2007). The symbiotic association involves most plant species and certain fungal species which has great relevance to soil ecosystem functions, especially nutrient dynamics, microbial processes, plant ecology and agriculture. The mycorrhiza performs beneficial functions for crops, like other microorganisms such as; phosphate-solubilizing bacteria (Panhwar *et al.*, 2009) and N₂ fixing bacteria (Naher *et al.*, 2009). The fungus colonizes the host plant's roots inside the cortical tissues. The association may be either intracellular like AMF, or extracellularly as in ectomycorrhizal fungi. The root infection by the mycorrhiza increases active absorptive surface area and stimulate nutrient and water up take even in water stress condition.

Mycorrhizal colonization increases disease suppression capability of the host plant. AMF occur naturally and are the important component of tropical soil system (Cardoso and Kuyper, 2006). The biomass of AMF accounted for 54-900 kg ha⁻¹ (Zhu and Miller, 2003). It forms natural association with several crops including annuals, vegetables, fruit trees

and also industrial crops like oil palm. Mycorrhizal colonization on various crops and their beneficial effects are well documented in many countries all over the world. Currently in Malaysia, much work has been done on selection of effective indigenous mycorrhizae and exploiting their beneficial effects for enhancing crop production. This paper briefly discusses the scope and benefits of mycorrhizal association on several vegetables, field and industrial crops.

Effect of Mycorrhizal Inoculation on Growth and Yield of Vegetables

Mycorrhizal colonization is common in tomato plants and well documented as a mycotrophic plant (Kubota *et al.*, 2005). Many studies were conducted to assess the response on seedlings of tomato with the inoculation of AMF. The inoculated seedlings showed better performance due to its higher shoot fresh weight (11.28 g plant⁻¹), high shoot/root ratio (0.236), higher root biomass (2.17 g plant⁻¹) higher relative growth rate (7.34 mg g⁻¹day⁻¹) and unitleaf rate 1.28 mg cm⁻¹ day⁻¹ (Oseni *et al.*, 2010).

Several species of AMF species had positive effect on growth of tomato plant. Studies were conducted by Tahat *et al.* (2008a) using three AMF species *Glomus mosseae*,

Scutellospora sp. and *Gigaspora margarita* on tomato plants grown on alluvial soil in Malaysia. Higher root colonization (80%), shoot (2.8 g plant⁻¹), root growth (18.56 cm³) and spores (455/100 g soil) were obtained in *G. mosseae* inoculated plants. The plant growth improvement of tomato inoculated with mycorrhizal fungi might be associated with higher nutrient uptake, which was also reported earlier by Perner et al. (2007) and Aboul-Nassar (1996).

Mycorrhizal inoculation was also beneficial for lady's finger (*Abelmoschus esculentus*). Several greenhouse and field studies showed growth and yield improvement of lady's finger with mycorrhizal inoculation. The study showed that *G. mosseae* was able to colonize lady's finger and showed a synergistic effect with plant growth promoting rhizobacterial (PGPR) isolate UPMB10 (Radziah et al., 2007). A significant increase in plant biomass, yield and zinc uptake of lady's finger was observed with microbial inoculation (Table 1).

AMF have been observed to improve growth of brinjal (*Solanum melongena*) (Matsubara et al., 1995). Studies using multistrain biofertilizer (consisting of *G. mosseae*, P-solubilizing and N-fixing bacteria) along with reduced doses of N (75%) and P(67%) fertilizers of the recommended rate showed comparable growth and fruit yield as that of full fertilized treatment (Table 2). Similar findings were reported by Sen (2008) that a application *G. intraradices* on brinjal seedling, significantly affected the shoot length, shoot diameter, number of leaves, shoot fresh weight, shoot dry weight, root fresh weight and root dry weight. The inoculation of *Gigaspora margarita*- and *G. intraradices* on cucumber seedlings showed higher Fe concentrations in the shoots. However, both the mycorrhizae had significant effect on seedling stem diameter, leaf number, shoot height and root length and micronutrients in the leaf tissue (Tufenkci et al., 2012; Rouphael et al., 2010).

Gaurav et al. (2010) conducted a study in India, to find the effect of mycorrhiza on water stress tolerance in 34 varieties of potatoes. They found that leaf area index was increased by 60-88% and yield 32-39% with mycorrhizae application under water stress condition. Tuber deformity was decreased from 33 to 100% . In another study, Douds et al. (2007) found a 35-45% increase in the fresh potato tuber yield. There was a positive correlation between mycorrhizal colonization and nodule trehalose content in bean (Ballesteros-Almanza et al., 2010). Inoculation of pepper (*Capsicum annum*) with two AMF (*G. mosseae* and *G. intraradices*) species enhanced the relative water content, P, chlorophyll and carotenoid contents of (Çekic et al., 2012). Aguilera-Gomez et al. (1999) also reported profound effect of mycorrhizal inoculation in pepper.

Effect of Mycorrhizal Inoculation on Growth and Yield of Field Crops

The beneficial effect of mycorrhizal colonization on many field crops has been proven. In wheat crop the mycorrhizal

Table 1: Effect of AMF and PGPR inoculation on dry weights of shoot, root, fruits and Zn uptake of lady's finger

Treatment	Shoot dry weight (g plant ⁻¹)	Root dry weight (g plant ⁻¹)	Fruit (g plant ⁻¹)	Zn uptake (Fruit plant ⁻¹)
Control	47.55 b*	8.28 b	1.88 b	1.51 b
AMF	53.72 ab	12.66 a	4.27 ab	3.20 ab
UPMB10	53.33 ab	9.02 b	2.76 b	1.94 b
AMF + UPMB10	61.86 a	13.08 a	6.75 a	4.96 a

*Means in column followed by different letter(s) are significantly different at (p≤ 0.05) Source: Adopted from Azizul Hafiz et al. (2007)

Table 2: Effect of multistrain biofertilizer on the growth of brinjal in Malaysia

Treatments	Plant height (cm)	Fruit weight (g plant ⁻¹)
T ₁) Organic (2t ha ⁻¹) + (NPK @ 200-75-75) kg ha ⁻¹	61.0	109.8
T ₂) Biofertilizer + (NPK @ 150 -75-75) kg ha ⁻¹	64.3	122.0
T ₃) Biofertilizer + (NPK @ 150 -50-75) kg ha ⁻¹	65.0	142.2
T ₄) Organic (2t ha ⁻¹) + (NPK @ 150 -50-75) kg ha ⁻¹	61.0	134.3
T ₅) Fertilizer (NPK @ 200-75-75 kg ha ⁻¹)	61.3	128.3

colonization was higher in well watered plants while lower in water-stressed plants. The AMF had a positive effect on many field crops. Mycorrhizal colonization was higher in well drained plants compared with stressed plants. Higher wheat plant biomass, grain yields with higher shoot P and Fe concentrations were recorded with mycorrhizal inoculation (Al-Karaki, 1998; 2004). The application of insoluble P was applied to rice plants inoculated with either *G. mosseae* or *G. intraradices*, showed increased P uptake. The root colonization by AMF inoculation increased (117%) in flooded condition. However, in the absence of P, shoot and root dry weight increased by 86-206%. Mycorrhizal colonization had also a major contribution in the uptake of P and K in the plants (Hajiboland et al., 2009).

Aerobic rice genotypes inoculated with mycorrhizal inoculums showed high root colonization (28-57%) (Gao et al., 2007). The colonization of mycorrhizal inocula with upland rice seedlings proved beneficial to achieve higher yields at optimum fertilizer dosage. Hence, it decreased the costs and environmental pollution (Rajeshkannan et al., 2009). Sweet corn inoculated with AMF (*G. mosseae*) showed an increased plant biomass and plant tissue N (24.2%), P (8.4%) and K (18.2%), content (Fig. 1).

AM fungus (*G. mosseae*) grows well in acid sulfate soil. Corn plants grown on acid sulfate soil applied with different levels of ground magnesium limestone (GML) at 2, 4, 6 and 8 ton ha⁻¹ with AM enhanced growth and uptake of nutrients (Fig. 2). Application of 8 ton ha⁻¹ GML with mycorrhiza inoculation produced higher plant height, biomass, and N and P uptake compared to treatment without mycorrhiza. The ability of AMF in regulating the low P availability together with the high Al activity of acid sulfate soil demonstrated crucial role of AMF symbiosis in nutrient cycling especially the P.

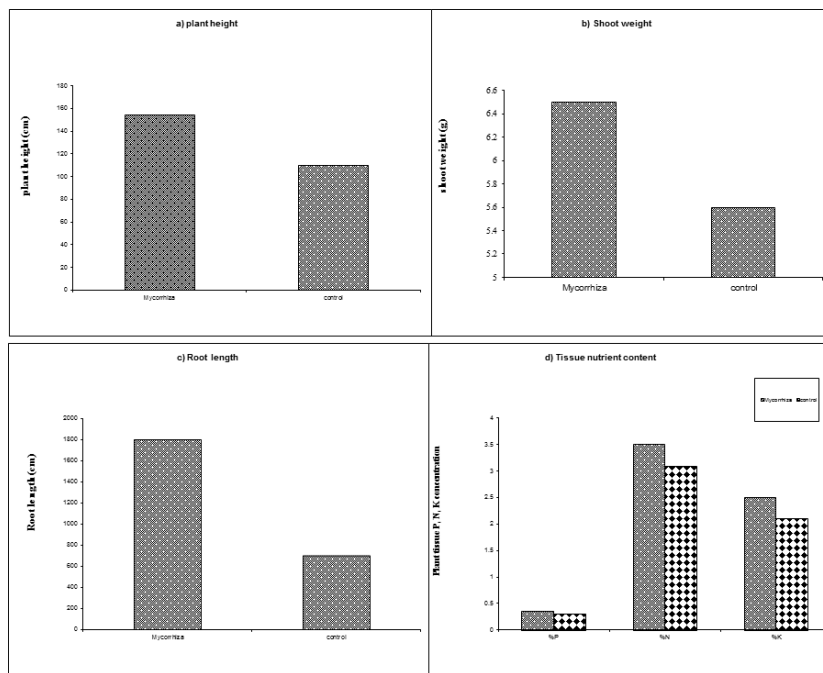


Fig. 1: Effects of *Glomus mosseae* colonization on sweet corn: (a) plant height, (b) shoot weight, (c) root length, (d) tissue nutrient concentrations. Sweet corn was grown in alluvial soil for 10 weeks

Source: Modified from Mustafa *et al.* (2010)

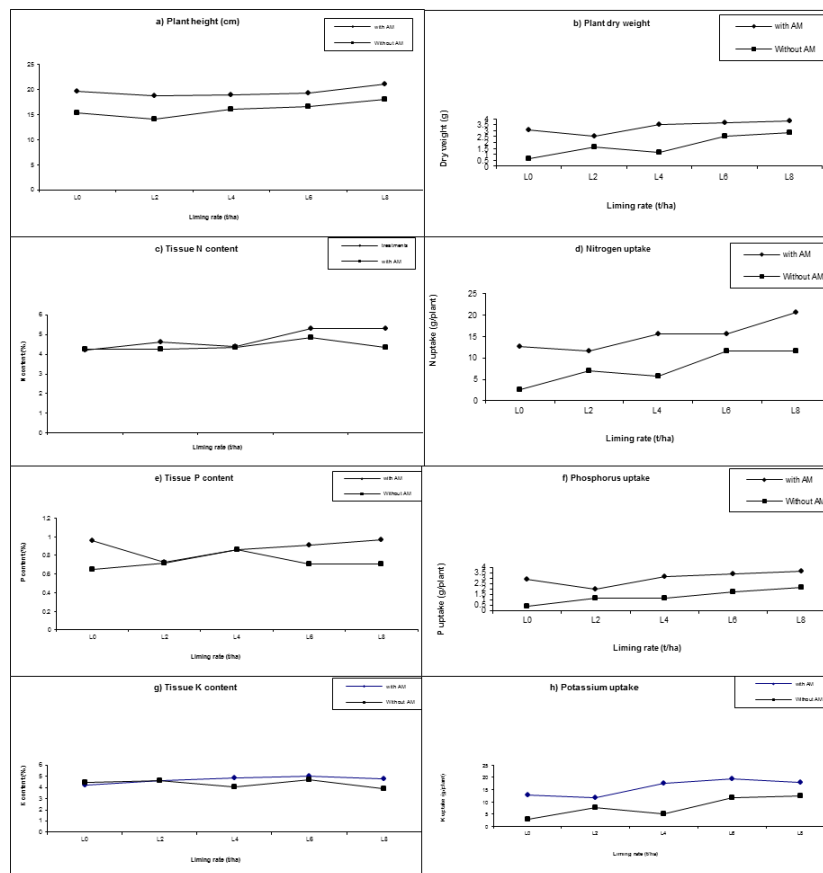


Fig. 2: Effect of different rate of liming and mycorrhiza inoculation on growth and nutrient uptake of corn: (a) plant height; (b) shoot weight, (c) tissue N, (d) N uptake, (e) tissue P, (f) P uptake, (g) tissue K, (h) K uptake

Mycorrhizal inoculation is generally accepted as substitute for the chemical fertilizers (especially phosphorus) to continuously enhance the productivity of maize crop. Mycorrhizal fungi trials perform better in terms of plant height (Shrestha *et al.*, 2009).

Furthermore, high impact of mycorrhizal inoculation on 100 seed weight and seed number per ear and grain yield of maize proved the successful symbiosis (Mobasser *et al.*, 2012). Khalil *et al.* (1992) observed marked differences in root infection of soybean by VAM fungi with a spore count of 66 to 998/100g soil. Generally, root colonization ranges from 60-100% from most soils except with very high soil P levels. The effect of AMF on plant mostly was positively related to the inoculation percentage (Sheng *et al.*, 2004). On the other hand, an increase was found in *S. viarum* medicinal plants inoculated with mycorrhiza (Hemashenpagam and Selvaraj, 2011).

Many other crops have been proven for positive growth response to AMF, however there have been genetic variation observed within plant species. The response of AMF also varied among the different cultivars of crops including *Zea mays* (Kaepler *et al.*, 2000), *Avena sativa* (Koide *et al.*, 1988), *Hordeum vulgare* (Zhu *et al.*, 2003), *Triticum aestivum* (Zhu *et al.*, 2001; Li *et al.*, 2006), tomato (Bryla and Koide, 1990a, b), soya (Bethlenfalvay *et al.*, 1989) etc.

Effect of Mycorrhizal Inoculation on Growth and Yield of Industrial Crops

Oil palm is the main industrial crop in Malaysia. Application of AMF inoculum either as single species (*Glomus* sp.) or mixed species (*Acaulospora* sp., *Gigaspora* sp., *Glomus* sp., *Scutellospora* sp.) showed better growth performance compared to that of chemical fertilizers (Fig. 3). Single AM species produced longer frond length while, significantly high pinnae number was obtained from mixed mycorrhizal inoculums. Both AM inoculums showed almost similar overall vegetative growth performance.

Even though AMF are of significance to the crops, comparatively few published reports on the interaction of AMF and oil palms were reported. Blal *et al.* (1990) showed that the coefficient of fertilizer use in micropropagated oil palms was increased 4–5-folds after inoculation with mycorrhiza, mainly when using rock phosphate. Similar findings were reported by Motta and Munévar (2005) that the inoculation of oil palm seedlings in the nursery resulted in a 3-fold growth enhance over non- inoculated plants after 570 days in natural soil substrate with no fertilizer addition.

In South East Asia, a survey on oil palm soil first of all explained a beneficial effect of mycorrhizal inoculation on oil palm (Nadarajah, 1980). The inoculation of oil palm seedlings with AMF increased plant growth and nutrient uptake of oil palm particularly P uptake enhanced by 37–44% (Widiastuti and Tahardi, 1993).

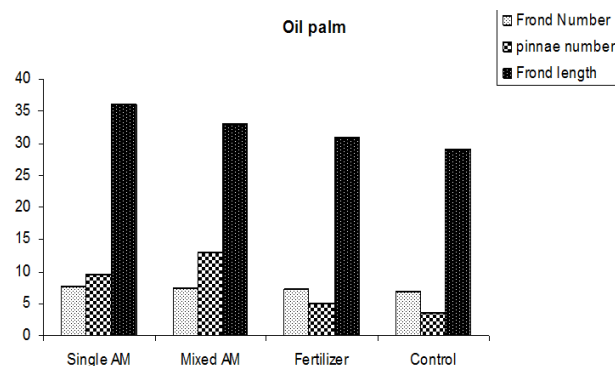


Fig. 3: Effects of AM inoculums on growth of oil palm

Source: Modified from Mohd Nazif Saifuddin *et al.* (2010)

Table 3: Effect of AM and PGPR inoculation on growth of pineapple

Treatments	Dry weight (g plant ⁻¹)	Number of leaves plant ⁻¹	Leaf length (cm)	No of spores 10 g soil ⁻¹
Control	2.08	19	16.15	0
AM	4.12	22	19.05	10
PGPR	4.95	20	22.0	0
AM + PGPR	5	20	17.02	9

AM= arbuscular mycorrhiza, PGPR = Plant growth

Kenaf is usually grown in beach ridges interspersed with swales (BRIS) marine deposit soil. This type of soil is considered a poor soil in terms of nutrient and has very low water holding capacity. Nutrient leaching is common as it is sandy. AMF play a vital role in nutrient and water deficiency management of plants grown in this soil. The AMF colonization significantly improved plant height (76.7%) and root length (40.9%) of kenaf (Fig. 4).

Effect of Mycorrhizal Inoculation on Fruit Crops

Beneficial effect of mycorrhizal inoculation found in fruit crops. The occurrence of AM fungi studied in Malaysia in two perennial fruits namely, durian (*Durio zibethinus*) and rambutan (*Nephelium lappaceum*). Higher spores were found in rambutan orchard. It is well known that AM inoculation at early stages of plant development performed better. The micropropagated banana plant inoculated at the beginning of the weaning phase showed significant growth response (Declerck *et al.*, 1994; Grant *et al.*, 2005). A glasshouse experiment was conducted to determine the effect of AMF and plant growth promoting rhizobacteria on growth and nutrient uptake of tissue cultured pineapple seedlings (Table 3). There was no significant difference for N uptake, however, significant difference for P uptake. A high P uptake was found in mycorrhizal inoculated treatments. Various glasshouse and field experiments proved that inoculation with AMF enhanced the growth and ion uptake in citrus plants, and improved tolerance to drought and salt stress and also the quality of fruit (Wu and Zou, 2009; Wu *et al.*, 2010a, b). Moreover, it is reported

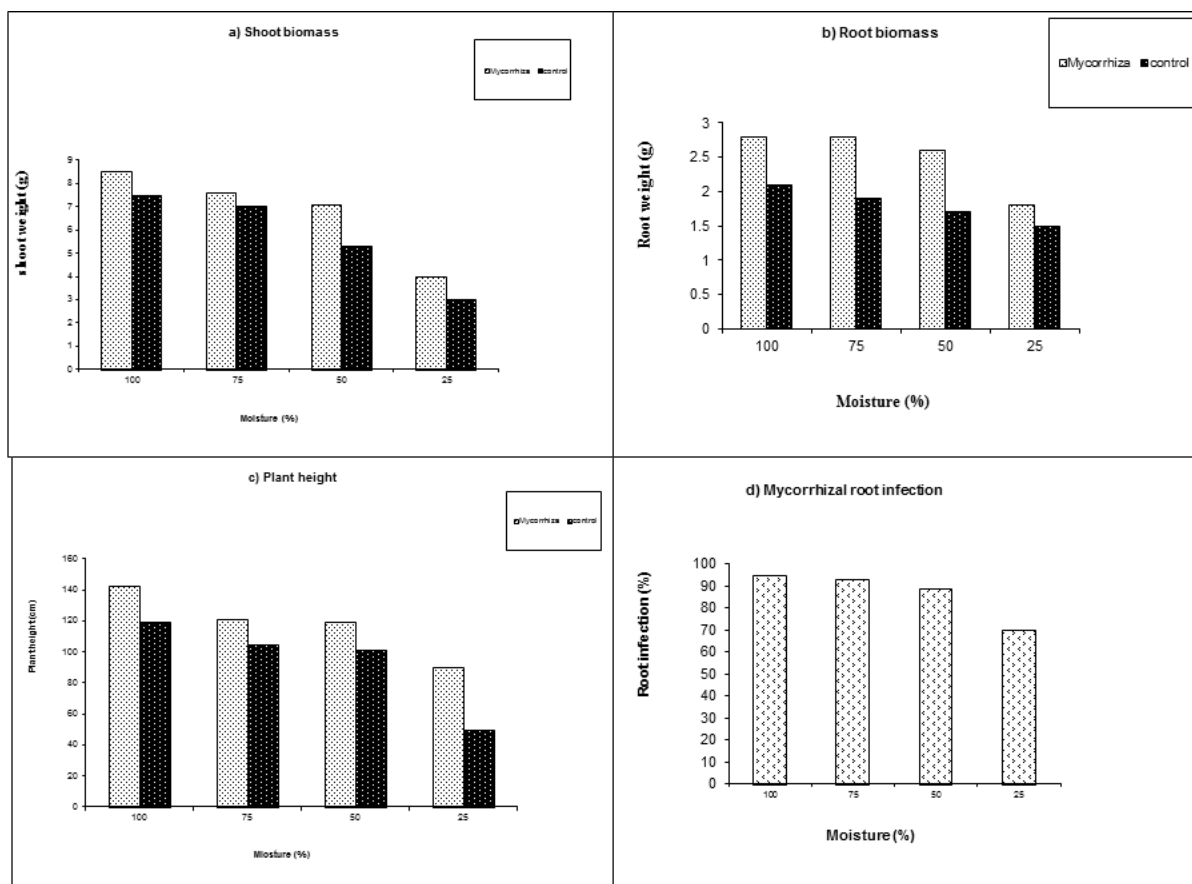


Fig. 4: Effect of mycorrhizal colonization in kenaf plant with different moisture levels grown on BRIS soil: (a) shoot biomass, (b) root biomass, (c) plant height, (d) mycorrhizal root infection

that the symbiosis of AMF in trifoliolate orange enhanced the soluble sugar and leaf chlorophyll content. However, one-layer mycorrhizal inoculation was the best for mycorrhization of trifoliolate orange (Wu and Zou, 2012).

Potential of Mycorrhiza as Biocontrol Agent

The biocontrol system in plants can be described as the management of frequent components to protect plants against pathogens. A number of findings showed that mycorrhizal inoculation directly or indirectly increased plant defense against pathogen. AMF inoculation improved plant nutrition; compensated the root damage and changed mycorrhizosphere microbial populations and augmented activation of plant defense mechanisms (Hooker *et al.*, 1994; Linderman, 1994). AM inoculated plant generated various compounds such as phytoalexins, enzymes of the phenylpropanoid pathway, chitinases, β -1,3-glucanases, peroxidases, pathogenesis-related (PR) proteins, callose, hydroxyproline-rich glycoproteins (HRGP) and phenolics (Bowles, 1990; Gianinazzi-Pearson *et al.*, 1994). These compounds are related to plant defense mechanisms. AMF initiated host defense responses which were subsequently suppressed disease infestations. A report by Gianinazzi and

Gianinazzi-Pearson (1992) showed that there was a minor change in the accumulation of defense-related transcriptions in *G. intraradices* colonized roots compared to un-inoculated controls. However, peroxidase activity concerned with epidermal and hypodermal cells was enhanced in mycorrhizal roots, which could be a mechanism for higher resistance to root pathogens. The enhanced lignification of root endodermal cells by AMF colonization has been recorded (Dehne, 1979). Nevertheless, AMF symbiosis-specific genes control the expression of the genes concerned to plant defense during the establishment of AMF (Gianinazzi-Pearson *et al.*, 1994; 1995; 1996).

AMF are the main components of many plant rhizosphere and play an important role in decreasing plant disease (Akhtar and Siddiqui, 2008). The application of AMF as a biocontrol agent performed a vital role in plant resistance and display greater potential to protect bean plants against the infection with *F. solani* (Al-Askar and Rashad, 2010). *Ralstonia solanaceum* is a causal agent of bacterial wilt disease in many vegetable crops including tomato (Vaillau *et al.*, 2007). Tomato plants inoculated with *G. mosseae* reduced infestation of *R. Solanaceum*. Moreover, the positive results of *G. etnicatumium* and *G.*

margarita spores were found against the *Verticillium* disease in brinjal (Matsubara *et al.*, 1995).

AMF are able to grow with *Trichoderma harzianum*, which is known as a bio-enhancer and have potential to suppress many diseases of oil palm. Results showed that, although, *T. harzianum* infused-compost (TC) gave highest yield, these mixed inoculum (TC and AMF) produced higher yield than AMF alone (Table 4).

Table 4: Effect of mycorrhiza and *Trichoderma harzianum* inoculation on oil palm growth

Treatments	Girth (cm)	Fronde number	Fronde length (cm)	Pinnae number
Control	28.4d	12.5b	154.2	58d
<i>T. harzianum</i>	80.2a	20.3a	193.5	112.3a
<i>Glomus</i> sp.)	60 c	15.99c	148.2	83.9c
<i>T. harzianum</i> + <i>Glomus</i> sp.)	72.3b	19b	173.2	98b

Means in column followed by different letter(s) are significantly different at ($p \leq 0.05$) Source: Adopted from Mohd Nazif Saifuddin *et al.* (2010)

Effect of P Nutrition on Mycorrhizal Infection

The possible factor that can affect AMF root colonization with plant is the P status. Increased P concentration in soil solution decreased mycorrhizal association, spore production and formation of secondary external hyphae (Menge *et al.*, 1978; Bruce *et al.*, 1994; De Miranda and Harris, 1994; Lu *et al.*, 1994; Valentine *et al.*, 2001). There are many evidences that mycorrhizal plants can absorb more P if lower P concentration exists in the soil solution. The AMF hyphae diameter may be a possible mechanism in this process. Barber (1984) explained that the radius of hyphae was much smaller than that of roots hairs and capable to absorb in low soil solution P.

High phosphorus status in the plant root tissue also reduced secretion of signal molecules that are responsible for hyphal branching and mycorrhizal association (Nagahashi and Douds, 2000). Moreover, cell phospholipids influencing membrane permeability and release of carbon compound that is essential for mycorrhizal colonization (Graham *et al.*, 1981; Schwab *et al.*, 1991). Muthukumar and Udaiyan (2000) reported that soluble carbon in cowpea root was increased with decreasing tissue P levels. However, increased P concentration reduced extra-radical mycelium less than colonization (Olsson *et al.*, 1999). Miller *et al.* (1995) found that application of P fertilizer does not always reduce mycorrhizal colonization. Factors reduced soil solution P influenced mycorrhizal association and noticeably high mycorrhizal association was found in soils with high P fixing capacity (Fardeau, 1998).

It is well understood that application of higher P levels hampers mycorrhizal formation (Bolan *et al.*, 1984) and in some plants the mycorrhizal benefits can be annulled. Wood (1992) recorded some limitations of P and suggested 5 ppm Olsen P as a common lowest amount generally promoted

mycorrhizal colonization. However, mycorrhizal colonization of roots declined quickly as P levels rose from 5 to 10 ppm. Though, Stribley *et al.* (1980) proposed the mycorrhizal effect in the highly mycorrhizal dependent leek plant was not negated at 200 ppm Olsen P in the soil solution. In general, higher rates of the P fertilizer application to the crops showed reverse effect of mycorrhiza (Phosri *et al.*, 2010); e.g., barley at 83 kg Clarke and Mosse (1981), maize at 50 kg (McGonigle *et al.*, 1990) and soyabean 176 kg (Ross, 1971) of P ha⁻¹, respectively.

AMF Inoculums Production

There are still several problems faced in the production of AMF inoculum on the commercial scale. Selection of suitable host is important in ensuring high propagule production in a short period of time. *G. mosseae* is the most common mycorrhiza adapted in tropical soil and it can form mutual association with different crops. A number of plant species were selected for the multiplication of *G. mosseae*. At 75 days of growth the highest number of AMF spores was found in corn as the host plant (Tahat *et al.*, 2008b). Nadarajah and Nawawi (1990) observed a significant correlation between root colonization and spore density. A high spore density of 400-4000 100 g⁻¹ in dried rubber soil was found in Srilanka (Jayaratne and Waidyanatha, 1982).

Conclusion

AMF colonized with several cereal, vegetables, fruits and industrial crops. Among the species studied, *G. mosseae* showed better colonization and improved plant growth, yield, nutrient uptake, water use efficiency and disease resistance. However, mycorrhizae exhibited better performance in low P added soil.

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References

- Aboul-Nassar, A., 1996. Effect of vesicular-arbuscular mycorrhiza on *Tagetes erecta* and *Zinnia elegans*. *Mycorrhiza*, 6: 61–64
- Aguilera-Gomez, L., Jr. F.T. Davis, S.A. Duray, L. Phavaphutanon and V. Olalde-Portugal, 1999. Influence of phosphorus and endomycorrhiza (*Glomus intradices*) on gas exchange and plant growth of Chile ancho pepper (*Capsicum annuum* cv. San Luis). *Photosynthetica*, 36: 441–449
- Al-Karaki, G.N. 1998. Benefit, cost and water-use efficiency of arbuscular mycorrhizal durum wheat grown under drought stress. *Mycorrhiza*, 8:41–45
- Al-Karaki, G., B. McMichael and J. Zak, 2004. Field response of wheat to arbuscular mycorrhizal fungi and drought stress. *Mycorrhiza*, 14: 263–269
- Al-Askar, A.A. and Y.M. Rashad, 2010. Arbuscular mycorrhizal fungi: A biocontrol agent against common bean fusarium root rot disease. *Plant Pathol. J.*, 9: 31–38

- Akhtar, M.S. and Z.A. Siddiqui, 2008. Arbuscular mycorrhizal fungi as potential biopesticides against plant pathogens. In: *Mycorrhizae: Sustainable Agriculture and Forestry*. Siddiqui, Z.A., M.S. Akhtar and K. Futai (eds.), Springer Netherlands, Dordrecht, The Netherlands
- Ballesteros-Almanza, L., J. Altamirano-Hernandez, J.J. Peña-Cabriales, G. Santoyo, J.M. Sanchez-Yañez, E. Valencia-Cantero, L. Macias-Rodriguez, J. Lopez-Bucio, R. Cardenas-Navarro and R. Farias-Rodriguez, 2010. Effect of Co-inoculation with mycorrhiza and rhizobia on the nodule trehalose content of different bean genotypes. *The Open Microbiol. J.*, 4: 83–92
- Barber, S.A., 1984. *Soil Nutrient Bioavailability: A Mechanistic Approach*, p: 398. John Wiley and Sons Inc., New York, USA
- Bethlenfalvai, G.J., R.L. Franson, M.S. Brown and K.L. Mihara, 1989. The *Glycine-Glomus-Bradyrhizobium* symbiosis. Nutritional, morphological and physiological-responses of nodulated soybean to geographic isolates of the mycorrhizal fungus *Glomus mosseae*. *Physiol. Plant.*, 76: 226–232
- Blal, B., C. Morel, V. Gianinazzi-Pearson, J.C. Fardeau and S. Gianinazzi, 1990. Influence of vesicular-arbuscular mycorrhizae on phosphate fertilizer efficiency in two tropical acid soils planted with micropropagated oil palm (*Elaeis guineensis* Jacq.). *Biol. Fert. Soils*, 9: 43–48
- Bolan, N.S., A.D. Robson and N.J. Barrow, 1984. Increasing phosphorus supply can increase the infection of plant roots by vesicular-arbuscular mycorrhizal fungi. *Soil Biol. Biochem.*, 16: 419–420
- Bowles, D.J., 1990. Defense-related proteins in higher plants. *Ann. Rev. Biochem.*, 59: 873–907
- Bruce, A., W.E. Smith and M. Tester, 1994. The development of mycorrhizal infection in cucumbers: effects of P supply on root growth, formation of entry points and growth of infection units. *New Phytol.*, 127: 507–514
- Bryla, D.R. and R.T. Koide, 1990a. Regulation of reproduction in wild and cultivated *Lycopersicon esculentum* Mill by vesicular-arbuscular mycorrhizal infection. *Oecologia*, 84: 74–81
- Bryla, D.R. and R.T. Koide, 1990b. Role of mycorrhizal infection in the growth and reproduction of wild vs cultivated plants. 28 wild accessions and 2 cultivars of *Lycopersicon esculentum* Mill. *Oecologia*, 84: 82–92
- Cardon, Z.G. and J.L. Whitbeck, 2007. *The Rhizosphere*, p: 235. Elsevier Academic Press
- Cardoso, I.M. and T.W. Kuyper, 2006. Mycorrhizas and tropical soil fertility. *Agric. Ecosyst. Environ.*, 116: 72–84
- Çekic, F.O., S. Unyayar and I. Ortas, 2012. Effects of arbuscular mycorrhizal inoculation on biochemical parameters in *Capsicum annuum* grown under long term salt stress. *Turk J. Bot.*, 36: 63–72
- Clarke, C. and B. Mosse, 1981. Plant growth responses to VAM. XII. Field inoculation responses of barley at two soil P levels. *New Phytol.*, 87: 695–703
- Declercq, S., B. Devos, B. Delvaux and C. Plenchette, 1994. Growth response of micropropagated banana plants to VAM inoculation. *Fruits*, 49: 103–109
- De Miranda, J.C.C. and P.J. Harris, 1994. Effects of soil phosphorus on spore germination and hyphal growth of arbuscular mycorrhizal fungi. *New Phytol.*, 128: 103–108
- Dehne, H.W. and F. Schonbeck, 1979. Influence of endotrophic mycorrhiza on plant diseases. II. Phenol metabolism and lignification. *J. Phytopathol.*, 95: 210–216
- Douds, D.D. Jr., G. Nagahashi, C. Reider and P.R. Hepperly, 2007. Mycorrhizal Fungi Increases the yield of potatoes in a high P soil. *Biol. Agric. Floricult.*, 25: 67–78
- Fardeau, J.C., C. Morel and R. Boniface, 1988. Phosphore assimilable des sols. Quelle méthode choisir en analyse de routine. *Agronomie*, 8: 577–584
- Graham, J.H., R.T. Leonard and J.A. Menge, 1981. Membrane-mediated decrease in root exudation responsible for phosphorus inhibition of vesicular-arbuscular mycorrhiza formation. *Plant Physiol.*, 68: 548–552
- Gao, X., T.W. Kuyper, C. Zou, F. Zhang and E. Hoffland, 2007. Mycorrhizal responsiveness of aerobic rice genotypes is negatively correlated with their zinc uptake when nonmycorrhizal. *Plant Soil*, 290: 283–291
- Gaurav, S.S., S.P.S. Sirohi, B. Singh and S. Pradeep, 2010. Effect of Mycorrhiza on Growth, yield and tuber deformity in potato (*Solanum tuberosum* L.) grown under water stress conditions. *Prog. Agric.*, 10: 35–41
- Gianinazzi, S. and V. Gianinazzi-Pearson, 1992. Cytology, histochemistry and immunocytochemistry as tools for studying structure and function in endomycorrhiza. *Methods Microbiol.*, 24: 109–139
- Gianinazzi-Pearson V., A. Gollotte, C. Cordier and S. Gianinazzi, 1996. Root defense responses in relation to cell and tissue invasion by symbiotic microorganisms: cytological investigations. In: *Histology, Ultrastructure and Molecular Cytology of Plant-Microorganism Interactions*, pp: 177–191. Nicole M. and V. Gianinazzi-Pearson (eds.). Kluwer Academic Publishers, Dordrecht, The Netherlands
- Gianinazzi-Pearson V., M.C. Lemoine, C. Arnould, A. Gollotte and J.B. Morton, 1994. Localization of $\beta(1\rightarrow3)$ glucans in spore and hyphal walls of fungi in the Glomales. *Mycologia*, 86: 478–485
- Gianinazzi-Pearson, V., A. Gollotte, J. Lherminier, B. Tisserant, P. Franken, E. Dumas-Gaudot, M.C. Lemoine, D. van Tuinen and S. Gianinazzi, 1995. Cellular and molecular approaches in the characterization of symbiotic events in functional arbuscular mycorrhizal associations. *Can. J. Bot.*, 73: 526–532
- Grant, C., S. Bittman, M. Montreal, C. Plenchette and C. Morel, 2005. Soil and fertilizer phosphorus: Effects on plant P supply and mycorrhizal development. *Can. J. Plant Sci.*, 85: 3–14
- Hajiboland, R., N. Aliasgharad and R. Barzeghar, 2009. Phosphorus mobilization and uptake in mycorrhizal rice (*Oryza sativa* L.) plants under flooded and non-flooded conditions. *Acta Agric. Slovenica*, 93: 153–161
- Hemashenpagam, N. and T. Selvaraj, 2011. Effect of arbuscular mycorrhizal (AM) fungus and plant growth promoting rhizomicroorganisms (PGPR's) on medicinal plant *Solanum invarium* seedlings. *J. Environ. Biol.*, 32: 579–83
- Hooker, J.E., M. Jaizme-Vega and D. Atkinson, 1994. Biocontrol of plant pathogens using arbuscular mycorrhizal fungi. In: *Impact of Arbuscular Mycorrhizas on Sustainable Agriculture and Natural Ecosystems*, pp: 191–200. Gianinazzi, S. and H. Schliepp, (eds.). Birkhäuser Verlag, Basel, Switzerland
- Jayaratne, A.H.R. and U.P. de Waidyanatha, 1982. *Endomycorrhizas of Rubber Growing Soils of Sri Lanka and their Effect on Water Uptake*, pp: 328–359. Training course on Mycorrhiza Research Techniques, Provisional Report No. 12. International Foundation for Science, Sweden
- Kaeppeler, S.M., J.L. Parke, S.M. Mueller, L. Senior, C. Stuber and W.F. Tracy, 2000. Variation among maize inbred lines and detection of quantitative trait loci for growth at low phosphorus and responsiveness to arbuscular mycorrhizal fungi. *Crop Sci.*, 40: 358–364
- Khalil, S., T.E. Loynachan and H.S. McNabb, 1992. Colonization of soybean by mycorrhizal fungi and spore populations in Oowa soils. *Agron. J.*, 84: 832–836
- Koide, R., M. Li, J. Lewis and C. Irby, 1988. Role of mycorrhizal infection in the growth and reproduction of wild Vs cultivated plants. 1. Wild Vs cultivated oats. *Oecologia*, 77: 537–543
- Kubota, M., T.P. McGonigle and M. Hyakumachi, 2005. Co-occurrence of Arumund Paris-type morphologies of arbuscular mycorrhizae in cucumber and tomato. *Mycorrhiza*, 15: 73–77
- Li, H.Y., S.E. Smith, R.E. Holloway, Y.G. Zhu and F.A. Smith, 2006. Arbuscular mycorrhizal fungi contribute to phosphorus uptake by wheat grown in a phosphorus fixing soil even in the absence of positive growth responses. *New Phytol.*, 172: 536–543
- Linderman, R.G., 1994. Role of VAM fungi in biocontrol. In: *Mycorrhizae and Plant Health*, pp: 1–125. Pflieger, F.L. and R.G. Linderman, (eds.). American Phytopathological Society, St. Paul, Minnesota, USA
- Lu, S., P.G. Braunberger and M.H. Miller, 1994. Response of vesicular-arbuscular mycorrhizae of maize to various rates of P addition to different rooting zones. *Plant Soil.*, 158: 119–128
- Matsubara, Y., T. Harada and T. Yakuwa, 1995. Effect of inoculum density of Vesicular Arbuscular Mycorrhizal Fungal spores and addition of carbonized material to bed soil on growth of Welsh onion seedlings. *J. Jpn. Soc. Hortic. Sci.*, 64: 549–554
- McGonigle, T.P., D.G. Evans and M.H. Miller, 1990. Effect of degree of soil disturbance on mycorrhizal colonization and phosphorus absorption by maize in growth chamber and field experiments. *New Phytol.*, 116: 629–636
- Menge, J.A., D. Steirle, D.J. Bagyaraj, E.L.V. Johnson and R.T. Leonard, 1978. Phosphorus concentration in plants responsible for inhibition of mycorrhizal infection. *New Phytol.*, 80: 575–578
- Miller, M.H., T.P. McGonigle and H.D. Addy, 1995. Functional ecology of vesicular arbuscular mycorrhizas as influenced by phosphate fertilization and tillage in an agricultural ecosystem. *Crit. Rev. Biotechnol.*, 15: 241–255

- Mobasser, H.R. and A.Moradgholi, 2012. Mycorrhizal bio-fertilizer applications on yield seed corn varieties in Iran. *Ann. Biol. Res.*, 3: 1109–1116
- Motta, D.V. and F.M. Munévar, 2005. Response of oil palm seedlings to mycorrhization. *Palmas J.*, 26: 11
- Muthukumar, T. and K. Udaiyan, 2000. Influence of organic manures on arbuscular mycorrhizal fungi associated with *Vigna unguiculata* (L.) Walp. in relation to tissue nutrients and soluble carbohydrate in roots under field conditions. *Biol. Fert. Soils.*, 31: 114–120
- Nadarajah, P. and A. Nawawi, 1990. Zygomycetous endomycorrhizal fungi in some Malaysian soils under rubber (*Hevea brasiliensis*). In: *Research on Multipurpose Tree Species in Asia*. David, A.T. and K.G. MacDicken (eds.). Winrock International Institute for Agricultural Development
- Nadarajah, P., 1980. Species of Endogonaceae and mycorrhizal association of *Elaeis guineensis* and *Theobroma cacao*. In: *Tropical Mycorrhiza Research*, pp: 232–237. Mikola, P. (ed.). Clarendon Press, Oxford, UK
- Nagahashi, G. and D.D. Jr. Douds, 2000. Partial separation of root exudates components and their effects upon the growth of germinated spores of AM fungi. *Mycol. Res.*, 104: 1453–1464
- Naher, U.A., O. Radziah, Z.H. Shamsuddin, M.S. Halimi and I. Mohd Razi, 2009. Growth Enhancement and Root Colonization of Rice Seedlings by *Rhizobium* and *Corynebacterium* spp. *Int. J. Agric. Biol.*, 11: 586–590
- Olsson, P.A., I. Thingstrup, I. Jakobsen and E. Baath, 1999. Estimation of the biomass of arbuscular mycorrhizal fungi in a linseed field. *Soil Biol. Biochem.*, 31: 1879–1887
- Oseni, T.O., N.S. Shongwe and M.T. Masarirambi, 2010. Effect of arbuscular mycorrhiza (am) inoculation on the performance of tomato nursery seedlings in vermiculite. *Int. J. Agric. Biol.*, 12: 789–792
- Panhwar, Q.A., O. Radziah, M. Sariah and I. Mohd Razi, 2009. Solubilization of phosphate forms by phosphate solubilizing bacteria isolated from aerobic rice. *Int. J. Agric. Biol.* 11: 667–673
- Perner, H., D. Schwarz, C. Bruns, P. Mader and G. Eckhard, 2007. Effect of arbuscular mycorrhizal colonization and two levels of compost supply on nutrient uptake and flowering of pelargonium plants. *Mycorrhiza*, 17: 469–474
- Phosri, C., A. Alia Rodriguez, I.R. Sanders and P. Jeffries, 2010. The role of mycorrhizas in more sustainable oil palm cultivation. *Agric. Ecosys. Environ.*, 135: 187–193
- Rajeshkannan, V., C.S. Sumathi and S. Manian, 2009. Arbuscular mycorrhizal fungi colonization in upland rice as influenced by agrochemical application. *Rice Sci.*, 16: 307–313
- Radziah O., I. Che Fauziah, and A. Azizul Hafiz., 2007. Effect of Arbuscular Mycorrhizal Fungi and PGPR on Heavy Metal Uptake by Lady's Finger Grown on Sewage Sludge Amended Soil. In: *Biogeochemistry of trace elements: environmental protection, remediation and human health*. pp: 548–549. Zhu, YongGuan., Lepp, Nicholas (eds). Beijing : Tsinghua university press, China.
- Ross, J.P., 1971. Effect of phosphate fertilization on yield of mycorrhizal and nonmycorrhizal soybeans. *Phytopathology*, 61: 1400–1403
- Rouphael, Y., M. Cardarelli, E. Di Mattia, M. Tullio, E. Rea and G. Colla, 2010. Enhancement of alkalinity tolerance in two cucumber genotypes inoculated with an arbuscular mycorrhizal biofertilizer containing *Glomus intraradices*. *Biol. Fert. Soil*, 46: 499–509
- Sen, O., 2008. Tuz stresi altinda yetistirilen patlican fidelinin gelismesi ve besin elementi dcerikleri uzerine arbuscular mikorizal fungus (*Glomus intraradices*) uygulamalarinin etkisi. *Master Thesis*, University of Selcuk, Faculty of Agriculture, Department of Horticulture, Konya, Turkey
- Sheng, Z.W., L.Y. Fen, Z.S. Wu and X.X. Bao, 2004. Effects of arbuscular mycorrhizal fungi on plant growth and osmotic adjustment matter content of trifoliolate orange seedling under water stress. *J. Plant Physiol. Mol. Biol.*, 30: 583–588
- Shrestha, G., G.S. Vaidya, P. Binayak and B.P. Rajbhandari, 2009. Effects of arbuscular mycorrhiza in the productivity of maize and finger millet relay cropping system. *Nepal J. Sci. Technol.*, 10: 51–55
- Stribley, D.P., P.B. Tinker and R.C. Snellgrove, 1980. Effect of vesicular arbuscular mycorrhizal fungi on the relations of plant growth, internal phosphorus concentration and soil phosphate analyses. *J. Soil Sci.*, 31: 655–672
- Schwab, S.M., J. Menge and P.B. Tinker, 1991. Regulation of nutrient transfer between host and fungus in vesicular-arbuscular mycorrhiza. *New Phytol.*, 117: 387–398
- Tahat, M.M., S. Kamaruzaman, O. Radziah, J. Kadir and N.H. Masdek, 2008a. Response of *Lycopersicon esculentum* Mill. To different arbuscular mycorrhizal fungi species. *Asian J. Plant Sci.*, 7: 479–484
- Tahat, M.M., S. Kamaruzaman, O. Radziah, J. Kadir and N.H. Masdek, 2008b. Plant host selectivity for multiplication of *Glomus mosseae* spore. *Int. J. Bot.*, 4: 466–470
- Tufenkci, S., S. Demir, S. Şensoy, S. Ünsal, E. Demirel, C. Erdinc, S. Bicer, A. Ekincial and A. Kincialp, 2012. The effects of arbuscular mycorrhizal fungi on the seedling growth of four hybrid cucumber (*Cucumis sativus* L.) cultivars. *Turk J. Agric. For.*, 36: 317–327
- Vailleau, F., E. Sartorel, M.F. Jardinaud, F. Chardon, S. Genin, T. Huguet, L. Gentzbittel and M. Petitprez, 2007. Characterization of the interaction between the bacterial wilt pathogen *Ralstonia solanacearum* and the model legume plant *Medicago truncatula*. *Mol. Plant Microbe Interact.*, 20: 159–167
- Valentine, A.J., B.A. Osborne and D.T. Mitchell, 2001. Interactions between phosphorus supply and total nutrient availability on mycorrhizal colonization, growth and photosynthesis of cucumber. *Sci. Hort.*, 88: 177–189
- Widiastuti, H. and J.S. Tahardi, 1993. Effect of vesicular-arbuscular mycorrhizal inoculation on the growth and nutrient uptake of micropropagated oil palm. *Menara Perkebunan*, 61: 56–60
- Wood, T., 1992. VA mycorrhizal fungi: challenges for commercialization. In: *Handbook of Applied Mycology, Volume 4: Fungal Biotechnology*, pp: 823–847. Arora, D.K., R.P. Elander and K.G. Mukerji, (eds.). Marcel Dekker Inc., New York, USA
- Wu, Q.S. and Y.N. Zou, 2012. Evaluating Effectiveness of Four Inoculation Methods with Arbuscular Mycorrhizal Fungi on Trifoliolate Orange Seedlings. *Int. J. Agric. Biol.*, 14: 266–270
- Wu, Q.S., Y.N. Zou and X.H. He, 2010a. Contributions of arbuscular mycorrhizal fungi to growth, photosynthesis, root morphology and ionic balance of citrus seedlings under salt stress. *Acta Physiol. Plant*, 32: 297–304
- Wu, Q.S., Y.N. Zou and X.H. He, 2010b. Exogenous putrescine, not spermine or spermidine, enhances root mycorrhizal development and plant growth of trifoliolate orange (*Poncirus trifoliata*) seedlings. *Int. J. Agric. Biol.*, 12: 576–580
- Wu, Q.S. and Y.N. Zou, 2009. Mycorrhiza has a direct effect on reactive oxygen metabolism of drought-stressed citrus. *Plant Soil Environ.*, 55: 436–442
- Zhu, Y.G. and R.M. Miller, 2003. Carbon cycling by arbuscular mycorrhizal fungi in soil-plant systems. *Trends Plant Sci.*, 8: 407–409
- Zhu, Y.G., F.A. Smith and S.E. Smith, 2003. Phosphorus efficiencies and responses of barley (*Hordeum vulgare* L.) to arbuscular mycorrhizal fungi grown in highly calcareous soil. *Mycorrhiza*, 13: 93–100
- Zhu, Y.G., S.E. Smith, A.R. Barritt and F.A. Smith, 2001. Phosphorus (P) efficiencies and mycorrhizal responsiveness of old and modern wheat cultivars. *Plant Soil*, 237: 249–255

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