INTERNATIONAL JOURNAL OF AGRICULTURE & BIOLOGY ISSN Print: 1560–8530; ISSN Online: 1814–9596 16–539/2018/20–1–15–24 DOI: 10.17957/IJAB/15.0321 http://www.fspublishers.org



Full Length Article

Identification of QTLs for Yield-Related Traits using Two Sets of Introgression Lines with a Common Donor Parent in Rice

Jian Zhang^{1†}, Xiaoxue Ou^{1†}, Hui Hu^{1†}, Bin Du¹, Wenkai Lv¹, Longwei Yang¹, Danying Xing¹, Jianlong Xu^{2,3*}, Xianjin Qiu^{1,4*}, Tianqing Zheng³, Junying Xu¹, Zhixin Li¹, Yunbo Zhang¹ and Xiaoyan Wang¹

¹Hubei Collaborative Innovation Center for Crop Industry/Engineering Research Center of Ecology and Agricultural Use of

Wetland, Ministry of Education/College of Agriculture, Yangtze University, Jingzhou 434025, China

²Shenzhen Institute of Breeding & Innovation, Chinese Academy of Agricultural Sciences, Shenzhen 518120, China

³Institute of Crop Science, Chinese Academy of Agricultural Sciences, Beijing 100081, China

⁴Hubei Key Laboratory of Food Crop Germplasm and Genetic Improvement, Wuhan 430070, China

*For correspondence: xujlcaas@126.com; xjqiu216@yangtzeu.edu.cn

[†]These authors contributed equally to this work

Abstract

Rice yield is a complex trait that is the major target in rice breeding program. In the present study, two sets of introgression line populations derived from three parents were used to dissect the genetic basis of yield-related traits, explore the genetic background effect on QTL detection and mine pleiotropic QTL for yield-related traits. Among the three parents, IR75862 was the common donor parent, and Ce258 and Zhongguangxiang1 were two recipient parents. The two introgression line populations were evaluated for yield-related traits in the summer of 2012 at Jingzhou, China. Combined with genotype of the introgression lines, 35 QTLs were detected in all 12 chromosomes except chromosome 5 for eight yield-related traits, including plant height, panicle number per plant, panicle length, filled grain number per panicle, spikelet number per panicle, seed setting rate, thousand grain weight and grain yield per plant, which explained 6.36-36.37% of phenotypic variances. No common QTL was detected in the two backgrounds, and seven chromosome regions on chromosomes 2, 3, 4, 6 and 10 had pleiotropic effects on two or more traits, indicating large genetic background effect on expression of QTLs for yield-related traits. Total of 19 (54%) QTLs including newly identified QTLs, qGY3, qFGN10.2 and qSN10 with IR75862 alleles increased yield-related traits. Therefore, it's possible to develop new super rice varieties through introgressing or pyramiding the IR75862 alleles at the pleiotropic QTLs (qPL2.1 and qPL6.1) and novel QTLs (qGY3, qFGN10.2 and qSN10) into Ce258 and Zhongguangxiang1 by marker-assisted selection. © 2018 Friends Science Publishers

Keywords: Rice; Introgression line; Yield-related trait; Quantitative trait locus; Genetic background

Introduction

Rice (*Oryza sativa* L.) is a very important crop in the world, providing staple food for 60% of the world's people (Kumar *et al.*, 2013). In the past sixty years, rice yield has been sharply increased due to use of semi-dwarfism and heterosis (Zhang, 2007). However, world agriculture faces a big challenge to produce more 70% food for another 2.3 billion people by 2050 (FAO, 2009). The efficient way to solve this problem is to continuously increase rice yield per unit.

Grain yield per plant (GY) consisted of three components including panicle number per plant (PN), filled grain number per panicle (FGN), and 1000 grain weight (TGW), and affected by some other traits, such as plant height (PH), panicle length (PL), spikelet number per panicle (SN), and seed setting rate (SSR). All of them are typical quantitative traits and controlled by multiple genes which distributed on all 12 chromosomes (http://archive.gramene.org/qtl/). Classical genetic research indicated that both genetic (additive effect, dominant effect and epistasis effect) and environment are very important for them (Yu et al., 1997; Xing et al., 2002). With the development of rice functional genomics and molecular marker technology, researchers have mapped many QTLs for them using different populations (Ye et al., 2005; Dong et al., 2009; Fu et al., 2010; Zhang et al., 2010; Wei et al., 2012; Tan et al., 2013; Zhao et al., 2013; Shen et al., 2014). Some of QTLs with large effect were successfully cloned using map-based cloning strategy, including MOC1 (Li et al., 2003), D53 (Jiang et al., 2013; Zhou et al. 2013), Gn1a (Ashikari et al., 2005), Ghd7 (Xue et al., 2008), Ghd7.1 (Yan et al., 2013), Ghd8/DTH8 (Wei et al., 2010; Yan et al., 2011), IPA1/ WFP (Jiao et al., 2010; Miura et al., 2010), GN2 (Chen et al., 2017), GNP1(Wu et al., 2016), GS3 (Fan et al., 2006; Mao et al., 2010), GW2 (Song et al., 2007), qSW5/ GW5 (Shomura et al., 2008; Weng et al., 2008),

GS5 (Li et al., 2011), GW8 (Wang et al., 2012a, b), TGW6 (Ishimaru et al., 2013), GL7/ GW7 (Wang et al., 2015a, b), OsGRF4/ GL2/ GS2 (Che et al., 2015; Duan et al., 2015; Hu et al., 2015; Sun et al., 2016), and most of them are pleiotropic genes, which had large potential values on molecular breeding. Although more and more genes for yield-related traits have been characterized, only a few genes have been successfully used in molecular breeding for increasing rice yield (Wang et al., 2012b). Therefore, identification of more favorable genes/QTLs especially pleiotropic genes affecting yield-related traits from rice germplasms is very important and essential to develop high yield varieties in molecular breeding.

Although many genes were detected to have large effects on grain yield, the application of them on molecular breeding aren't always successful, because of insistence of genetic background between mapping population and breeding population. Many researchers indicated that genetic background has large effect on expression of QTL for panicle and grain size, drought and salt tolerance (Liao et al., 2001; Mei et al., 2006; Xie et al., 2008; Zheng et al., 2011; Cheng et al., 2012; Wang et al., 2013, 2014; Qiu et al., 2015). Although a great number of QTLs for rice yieldrelated traits have been reported, our understanding of the effect of genetic background on detection of QTL for vieldrelated traits is very limited, and very few QTL independent from genetic background have been identified. QTL identified in different populations could be compared and common QTLs could be mined in some special mapping populations derived from common parents, such as reciprocal recombinant inbred line (Zhao et al., 2016), reciprocal introgression line (IL) (Wang et al., 2014), two or more introgression lines with common parents (Peng et al., 2014; Qiu et al., 2015).

In our previous study, two sets of IL populations were developed derived from a common donor parent, IR75862-206-2-8-3-B-B-B (IR75862), and two recipient parents, Ce258 and Zhongguangxiang1 (ZGX1) (Qiu *et al.*, 2015). We used these two sets of introgression line populations (ILs) to map QTLs for yield-related traits. The objectives of this study were to (1) illustrate the genetic basis of yield-related traits, (2) dissect genetic background effect on QTL detection and detect QTLs independent from genetic background; (3) identify pleiotropic QTLs for two or more yield-related traits. The results will give us useful information to enhance rice potential in molecular breeding.

Materials and Methods

Plant Materials and Field Experiment

Two sets of IL populations were used in this study. They were derived from IR75862 as common donor parent, and Ce258 and ZGX1 as recipient parents, respectively. IR75862 is a tropical *japonica* glutinous variety with high

zinc and iron concentration introduced from IRRI (Impa *et al.*, 2013). Ce258 is an elite *indica* three-line restorer line with high general combining ability. ZGX1 is an *indica* elite variety with high quality. IR75862 was crossed with Ce258 and ZGX1, respectively and then the F_1 individuals were backcrossed with the recurrent parents for one time. The BC₁F₁ progenies were self-crossed without selections for nine generations to BC₁F₁₀. Ultimately, two sets of introgression lines (ILs) were developed for QTL mapping, each comprising 200 lines in Ce258 background (Ce258-ILs) and ZGX1 background (ZGX1-ILs), respectively.

A total of 403 lines, including 400 ILs and three parents, were planted in the summer season of 2012 on the farm of College of Agriculture, Yangtze University, Jingzhou, China (30.2° N, 112.7°E). Jinghzou is located in the middle of Yangtze River, and is a main production area of rice in China. Seeds were sown on May 10 and transplanted on June 5 with a randomized complete block design. Each IL was planted in three rows and ten individuals in each row at spacing of 20 cm × 20 cm with two replications. All field management followed local farmers' practices.

Trait Measurements

Upon harvest, yield-related traits were measured followed the method described by Wang *et al.* (2012a). GY, PH and PN were measured for five plants in the second row of each IL. PL, FGN, SN and SSR were recorded for three randomly selected panicles from each plant. TGW was measured based on 1000 filled grains.

DNA Extraction and Marker Analysis

Young leaves of about 30 individuals per IL were bulked and used to extract DNA using CTAB method with minor modifications (Murray and Thompson, 1980). Total of 128 and 133 SSR markers evenly distributed on rice genome (http://archive.gramene.org/markers/) were selected for genotyping ILs for Ce258-ILs and ZGX1-ILs, respectively. Among them, both two populations have 81 markers (Fig. 2). Marker locations on linkage maps were according to published rice linkage map (Temnykh *et al.*, 2001; McCouch *et al.*, 2002).

Data Analysis

Statistical description and correlations among different traits were analyzed by Statistica 5.5 (Morales, 2001). Inclusive composite interval mapping method (ICIM) in QTL IciMapping 3.2 was used for QTL mapping (Li *et al.*, 2007). The permutation method based on 1000 runs of randomly shuffling the trait values was used to obtain empirical thresholds (Churchill and Doerge, 1994).

Results

Performance of Yield-related Traits in the Two IL Populations

The statistical description of yield-related traits was demonstrated in Table 1. Significant differences were observed between the Ce258/ZGX1 and IR75862 for most of the traits. IR75862 had significantly higher SN and TGW but significantly lower FGN and SSR than those of the two recurrent parents Ce258 and ZGX1. For PH, IR75862 was similar to Ce258 but significantly higher than that of ZGX1. GY of IR75862 was similar to that of ZGX1, and they were higher than Ce258. All above traits exhibited continuous distributions with transgressive segregations in both two IL populations (Table 1; Fig. 1), indicating that all measured traits are typical quantitative traits controlled by multi-genes.

Correlations of the Traits in the Two Backgrounds

Correlation coefficients between different traits are given in Table 2. GY were significantly correlated with PN, PL, FGN, SN and SSR but had no correlation with TGW in both backgrounds, suggesting yield more dependent on panicle number and grain number than grain size in the two populations. As expected, TGW had significant negative correlations with PN and FGN in both backgrounds, suggesting there is trade-off among the three component traits. However, FGN was not significantly correlated with PN, suggesting yield could be to some extent improved through simultaneous increase of FGN and PN in the two populations.

There were some inconsistent correlations among the traits in the two populations, such as significant correlations existed between SN and PN, and FGN and PH only in the Ce258-ILs population, while between TGW and PH, TGW and PL and TGW and SSR only in the ZGX1-ILs population.

Identification of QTLs for Yield-related Traits

QTL identification of PH: Three QTLs were identified in the ZGX1-ILs (Table 3; Fig. 1). They were located on chromosome 3, 4 and 11. The phenotypic variations caused by each QTL were around 13.77–23.35%, respectively. Among these QTLs, *qPH11* had the biggest phenotypic variance rate. IR75862 alleles at all three loci increased PH. No QTL for PH was detected in Ce258-ILs, maybe because PH of Ce258 and IR75862 were similar.

QTL identification of PN: Three QTLs were detected in the Ce258-ILs (Table 3; Fig. 1). They were located on chromosome 3, 4 and 8. The QTLs explained 22.86–26.04% of phenotypic variance, respectively. Among these QTLs, qPN3 had the largest effect, and IR75862 allele at this locus increased PN. No QTL for PN was detected in ZGX1-ILs, probably because of no difference between that of ZGX1 and IR75862.

QTL identification of PL: Six QTLs influencing PL were identified in the two populations, located on chromosome 2, 3, 4 and 6 (Table 3; Fig. 1). Two QTLs were detected in Ce258-ILs and four in ZGX1-ILs, respectively. Each QTL explained 6.36-25.12% of phenotypic variance, respectively. Among them, *qPL2.1* detected in ZGX1-ILs had the largest effect. All positive alleles of QTL came from IR75862 except for *qPL3* and *qPL6.2*.

QTL identification of FGN: A total of five QTLs controlling FGN were detected in the two populations, located on chromosome 2, 3, 4 and 10. One QTL was identified in Ce258-ILs and four in ZGX1-ILs, respectively Each QTL explained 16.00–22.55% of phenotypic variance, respectively. Among these QTLs, qFGN4 detected in ZGX1-ILs contributed most. IR75862 alleles decreased FGN at all loci except for qFGN10.2

QTL identification of SN: Three QTLs influencing SN were scanned in the two populations, located on chromosome 2, 8 and 10 (Table 3; Fig. 1). One QTL was identified in Ce258-ILs and two in ZGX1-ILs, respectively, respectively. QTLs explained 15.15-23.07% of phenotypic variance, respectively. Among these QTLs, *qSN2* detected in ZGX1-ILs contributed most. IR75862 alleles increased SN at all loci except for *qSN8*.

QTL identification of SSR: Seven QTLs affecting SSR were founed for SSR in the two populations, located on chromosome 2, 3, 4, 7 and 10 (Table 3; Fig. 1). Three QTLs were found in Ce258-ILs and four in ZGX1-ILs, respectively. The QTLs explained up to 36.37% of phenotypic variance, respectively. Among these QTLs, qSSR10 detected in C258-ILs contributed most. IR75862 alleles decreased SSR at all loci.

QTL identification of TGW: Seven QTLs for TGW were mined in the two populations, located on chromosome 1, 3, 6, 9 and 12 (Table 3; Fig. 1). Three QTLs were detected in Ce258-ILs and four in ZGX1ILs, respectively. The QTLs explained up to 25.64% of phenotypic variance, respectively. *qTGW6* identified in ZGX1-ILs contributed most. IR75862 alleles at all loci increased TGW.

QTL identification of GY: One QTL for GY was identified in Ce258-ILs, located between RM282 and RM156 on chromosome 3 (Table 3; Fig. 1). This QTL explained 30.26% of phenotypic variance. IR75862 alleles increased GY. No QTL for GY was detected in ZGX1-ILs.

QTLs Common Detected in Both Backgrounds

Among total of 35 QTLs identified for yield-related traits in the two ILs, none QTL was consistently detected in both genetic backgrounds, clearly indicating large genetic background effect on expression of QTLs for yield-related traits.

	Ce258	ZGX1	IR75862	P_1-P_3	P ₂ -P ₃		Ce258-ILs			ZGX1-ILs	
	(P1)	(\mathbf{P}_2)	(P ₃)			$Mean \pm SD$	Range	CV (%)	Mean \pm SD	Range	CV (%)
PH	115.33	101.17	119.40	-4.07	-18.23**	113.78±5.83	93.67-129.00	5.13	104.46±6.22	89.10-128.42	5.95
PN	15.33	18.33	17.17	-1.84	1.16	13.87±2.67	5.00-24.83	19.95	18.17±3.13	8.67-29.35	17.24
PL	28.97	26.29	26.44	2.53	-0.15	28.24±1.67	19.83-33.70	5.91	26.18 ± 1.28	22.13-33.17	4.88
FGN	170.17	192.00	148.67	21.5*	43.33**	163.70±25.45	63.33-242.50	15.55	185.12±23.63	90.50-247.83	12.77
SN	201.17	223.92	243.08	-41.91**	-19.16*	212.03±30.18	79.67-305.58	14.24	228.50±24.35	145.00-327.50	10.66
SSR	84.59	85.65	63.86	20.73**	21.79**	77.37±7.36	43.26-87.98	9.51	81.02±6.26	48.79-89.68	7.72
TGW	23.43	20.62	26.36	-2.93**	-5.74***	24.69±1.75	18.20-31.98	7.10	18.22 ± 2.34	15.25-28.85	12.83
GY	61.14	72.57	71.70	-10.56**	0.87	53.19±14.22	16.08-129.76	26.25	60.74 ± 12.90	23.51-108.32	21.23

Table 1: Statistical descriptions of yield-related traits in two sets of ILs populations derived from a common donor, IR75862, and two recipient parents, Ce258 and ZGX1

* and **, represent significant levels at $P \le 0.05$ and 0.01, respectively; ZGX1, Zhongguangxiang 1; SD, Standard deviation; CV, Coefficient of Variation; Ce258-ILs, introgression lines at Ce258 background; ZGX1-ILs, introgression lines at Zhongguangxiang 1 background; PH: plant height, PN: panicle number per plant, PL: panicle length, FGN: filled grain number per panicle, SN: spikelet number per panicle, SSR: seed setting rate, TGW: thousand grain weight, GY: grain yield per plant

 Table 2: Correlation coefficients of yield-related traits in two sets of ILs populations derived from the crosses between IR75862 and Ce258 or ZGX1

Characteristics	PH	PN	PL	FGN	SN	SSR	TGW	GY
PH		-0.13	0.51**	-0.1	0.11	-0.31**	0.51**	0.08
PN	0.17*		-0.12	0.11	0.07	0.07	-0.25**	0.75**
PL	0.31**	0.13		0.22**	0.40**	-0.22**	0.54**	0.29**
FGN	0.22**	0.14	0.53**		0.79**	0.60**	-0.39**	0.53**
SN	0.33**	0.15*	0.55**	0.78**		-0.02	-0.06	0.52**
SSR	-0.11	-0.01	0.02	0.45**	-0.2**		-0.58**	0.16*
TGW	0.09	-0.16*	0.01	-0.14*	-0.21**	0.07		0.05
GY	0.26**	0.81**	0.39**	0.62**	0.49**	0.27**	0.02	

Note: Data under and above the diagonal are correlation coefficients in Ce258-ILs and ZGX1-ILs, respectively. * and **, represent significant levels at $P \le 0.05$ and 0.01, respectively; ZGX1, Zhongguangxiang 1; Ce258-ILs, introgression lines at Ce258 background; ZGX1-ILs, introgression lines at Zhongguangxiang 1 background; PH: plant height, PN: panicle number per plant, PL: panicle length, FGN: filled grain number per panicle, SN: spikelet number per panicle, SSR: seed setting rate, TGW: thousand grain weight, GY: grain yield per plant

Discussion

Genetic Background Effect on QTL Detection of Yieldrelated Traits

A lot of evidences indicated that effect of genetic background on OTL mapping is very large. The OTL effects in different backgrounds were diverse when different recipient parents were used (Yousef and Juvik, 2002). It was reported that below twenty percent of the QTLs for panicle size, resistance to drought and salt were detected in reciprocal ILs (Mei et al., 2006; Xie et al., 2008; Cheng et al., 2012; Wang et al., 2014; Oiu et al., 2015). Till now, very little researches reported genetic background effect on QTL expression for yield-related traits. It's uncertain whether it will be successful when a QTL is applied in molecular breeding for increasing yield potential, because no one knows if the QTL still works in a different genetic background. In this study, none QTL out of 35 QTLs for yield-related traits was identified in the both ILs. It was fully demonstrated that genetic background had very large effect on QTL detection for yield-related traits, and expressions of QTL for yield-related traits were diverse when genetic background changed. Therefore, if QTL mapping information is wanted to apply to rice molecular breeding for yield-related traits, much attention should be paid. It is essential for an

integration of QTL mapping with molecuar breeding in the same genetic background.

Comparison of QTLs among Different Populations and Novel QTLs Detected in this Study

In this study, 35 QTLs were detected for yield-related traits using two sets of IL populations with the same donor parent. Some QTLs were in the same or near regions with those previously reported. qPH3 flanked by markers of RM293 and RM571, qPH4 flanked by markers of RM349 and RM127 and *qPH11* between RM332 and RM167 for PH were mapped in the same regions or near OPh3b, OPh3c, OPh11, respectively (Zhang et al., 2013). qPN3 flanked by markers of RM51 and RM5488 and *qPN8* between RM284 and RM80 for PN were mapped in the adjacent regions harboring QPn3a and QPn8, respectively (Ishimaru et al., 2001; Zhang et al., 2013). qPL2.1, qPL3 and qPL4 located in the region of RM109-RM110, RM411-RM565 and RM349-RM127 for PL were mapped together with QTL in the region of RZ123-RZ446 (Xiao et al., 1996), qPLT3-3 (Hittalmani et al., 2003) and QTL in the region of XBpb331-XNpb235 (Kobayashi et al., 2003) for PL, respectively. qFGN3 and qFGN4 located in the region of RM85-RM227 RM119-RM273 for FGN were mapped near qSPP3 (Zhang et al., 2009) and the cloned gene NAL1 (Fujita et al., 2013), respectively. qSN8 located in the region

Background	Trait ¹	QTL	Chr.	Marker interval	LOD	A^2	$R^2(\%)^3$	
Ce258	PN	qPN3	3	RM251-RM5488	6.34	6.26	26.04	
		qPN4	4	RM119-RM273	4.86	-5.15	24.95	
		$\hat{q}PN8$	8	RM284-RM80	4.81	-4.23	22.86	
	PL	qPL2.2	2	RM112-RM250	2.61	1.36	6.36	
		qPL3	3	RM411-RM565	2.54	-3.54	16.45	
	FGN	qFGN10.1	10	RM467-RM3773	3.45	-26.30	18.56	
	SN	qSN10	10	RM467-RM3773	3.08	28.72	17.62	
	SSR	qSSR3.1	3	RM411-RM565	2.67	-6.89	17.71	
		qSSR4.1	4	RM335-RM1155	3.19	-5.71	35.62	
		qSSR10	10	RM467-RM3773	4.31	-10.97	36.37	
	TGW	qTGW1.1	1	RM3234-RM243	3.74	0.84	11.70	
		qTGW3.1	3	RM231-RM1324	3.42	0.84	12.52	
		qTGW12	12	RM4-RM247	7.27	1.59	20.65	
	GY	qGY3	3	RM282-RM156	4.38	3.44	30.26	
ZGX1	PH	qPH3	3	RM293-RM571	2.66	5.70	13.77	
		qPH4	4	RM349-RM127	4.02	6.62	15.93	
		qPH11	11	RM332-RM167	6.37	4.82	23.35	
	PL	qPL2.1	2	RM109-RM110	12.47	2.01	25.12	
		qPL4	4	RM349-RM127	3.58	1.00	11.97	
		qPL6.1	6	RM136-RM527	4.44	1.16	14.29	
		qPL6.2	6	RM3-RM162	7.48	-1.73	25.08	
	FGN	qFGN2	2	RM573-RM318	3.30	-27.17	16.00	
		qFGN3	3	RM85-RM227	3.91	-25.20	22.23	
		qFGN4	4	RM119-RM273	3.40	-28.49	22.55	
		qFGN10.2	10	RM258-RM228	2.84	38.25	19.75	
	SN	qSN2	2	RM110-RM211	3.74	48.69	23.07	
		qSN8	8	RM72-RM331	2.93	-39.67	15.15	
	SSR	qSSR2	2	RM573-RM318	4.59	-12.37	22.48	
		qSSR3.2	3	RM571-RM148	5.35	-11.69	22.02	
		qSSR4.2	4	RM349-RM127	6.00	-13.35	22.42	
		qSSR7	7	RM125-RM542	4.75	-11.97	21.96	
	TGW	qTGW1.2	1	RM488-RM473	9.72	1.97	17.90	
		qTGW3.2	3	RM571-RM148	8.84	1.97	16.73	
		qTGW6	6	RM276-RM136	13.70	1.83	25.64	
		qTGW9	9	RM316-RM105	4.56	1.41	14.28	

Table 3: QTLs affecting yield-related traits in two sets of ILs populations derived from crosses between IR75862 and Ce258 or ZGX1

¹ PH: plant height, PN: panicle number per plant, PL: panicle length, FGN: filled grain number per panicle, SN: spikelet number per panicle, SSR: seed setting rate, TGW: thousand grain weight, GY: grain yield per plant; QTL, quantitative trait locus; Chr., chromosome; ZGX1, Zhongguangxiang 1; ² A, addictive effect; the additive effect is the effect associated with substitution of Ce258 and ZGX1 alleles by the corresponding IR75862 alleles; ³ R^2 , Phenotypic variation explained by the QTL

of RM72-RM331 on chromosome 8 controlling SN was mapped near the IPA1/WFP (Jiao et al., 2010; Miura et al., 2010). qSSR2 flanked by RM573 and RM318 on chromosome 2, qSSR3.1 flanked by markers RM411 and RM565, which controlled SSR, were mapped in the adjacent regions with qFLn2-1 and qFLl3-1 (Wei et al., 2012). qSSR4.2 in the region of RM349-RM127 on chromosome 4 was mapped together with qSSR4 (Zhao et al., 2013). qTGW1.1 located in the region of RM3234-RM243, qTGW1.2 in the region of RM488-RM473 on chromosome 1, qTGW3.1 in the region of RM231-RM1324 and *qTGW3.2* in the region of RM571-RM148 on chromosome 3 for TGW were mapped in the same regions or near kgwl.1, kgwl.2, kgw3.1 and kgw3.2, respectively (Fu et al., 2010). qTGW6, qTGW9 and qTGW12 in the region of RM276-RM136, RM316-RM105 and RM4-RM247 were mapped together with gw6 (Dong et al., 2009), qGW9-1 (Li et al., 2009) and qKW12 (Ye et al., 2005), respectively. Whether the above QTLs identified in this study and the reported grain shape QTLs/genes are the same genes will need to be further clarified after fine mapping and cloning of the QTLs.

Based on above comparative mapping, eleven QTLs for yield components were found to be newly identified in the present study. Phenotypic variations explained by most QTLs were above 15%, and they had large addictive effects. For instance, the Ce258 alleles at qPN4, qFGN10.1, qSSR4.1 and qSSR10 can increase 5.15 panicles, 26.30 filled grains, 5.71 and 10.97% SSR, respectively. The ZGX1 allele at qFGN2, qSSR3.2 and qSSR7 can increase 27.17 filled grains, 11.69 and 11.97% SSR, respectively. The IR75862 alleles at qSN2, qSN10, qFGN10.2 and qGY3 can increase 48.69 and 28.72 spilekets, 38.25 filled grains and 3.44 g grain yield, respectively. Fine mapping and cloning of above new QTLs will facilitate understanding genetic basis of rice yield.





Fig. 1: Frequency distributions of eight yield-related traits in the two sets of IL populations using a common donor IR75862 introgressed separately into two recipients used as female parents, Ce258 and ZGX1. White and black bars represent IL populations with Ce258 and ZGX1 backgrounds, respectively ZGX1, Zhongguangxiang 1

Pleiotropic QTLs and QTL Clusters in this Study

One QTL or QTL clusters controlling two or more traits are very common in plants. For example, *Ghd7* (Xue *et al.*, 2008), *Ghd7.1* (Yan *et al.*, 2013) and *Ghd8/DTH8* (Wei *et al.*, 2010; Yan *et al.*, 2011) simultaneously control heading date, PH and SN. *GW2* (Song *et al.*, 2007), *qSW5/GW5* (Shomura *et al.*, 2008; Weng *et al.*, 2008), *GS5* (Li *et al.*, 2011), *GW8* (Wang *et al.*, 2012b) control grain width and weight, and GS3 and qSS7 had pleiotropic effects on grain length and width (Fan *et al.*, 2006; Qiu *et al.*, 2012). Wang *et al.* (2012a) detected nine QTL clusters affecting two or more plant type and yield traits.

In this study, seven regions on chromosomes 2, 3, 4, 6 and 10 had pleiotropic effects on two or more traits. For instance, both qFGN2 and qSSR2 were in the region of RM573-RM318, probably being a same QTL having pleiotropic effects on FGN and SSR with same direction of gene effect. Similarly, qPL3 and qSSR3.1 in RM411-RM565 region have pleiotropic effects on PL and SSR with same direction of gene effect. The QTLs qPH4, qPL4and qSSR4.2 in the region of RM349-RM127 on chromosome 4 had pleiotropic effects on PH, PL and SSR. The direction of gene effect of qSSR4.2 is opposite to that of qPH4 and qPL4. The QTLs qFGN10.1, qSN10and qSSR10 in the region of RM467-RM3773 on



ZGX1, Zhongguangxiang1; Ce258-ILs, introgression lines at Ce258 background; ZGX1-ILs, introgression lines at Zhongguangxiang 1 background; PH: plant height, PN: panicle number per plant, PL: panicle length, FGN: filled grain number per panicle, SN: spikelet number per panicle, SSR: seed setting rate, TGW: thousand grain weight, GY: grain yield per plant

chromosome 10 had pleiotropic effects on PH, PL and SSR, in which direction of gene effect of qSN10 is opposite to that of qFGN10.1 and qSSR10.

Besides, some QTLs were identified in the adjacent regions. *qPH3*, *qFGN3*, *qSSR3.2* and *qTGW3.2* were in the adjacent regions on chromosome 3, simultaneously controlling PH, FGN, SSR and TGW. Thus, we suppose that some tightly linked genes in a chromosome block control a series of yield-related traits. Similarly, two tightly linked genes in a block on chromosome 2 control PL and SN due to adjacent QTLs, *qPL2.1* and *qSN2*. Two tightly linked genes in a block on chromosome 6 control PL and TGW due to adjacent QTLs, qPL6.1 and qKGW6. These QTL clusters may be responsible for high correlations among the related traits.

The pleiotropic QTLs and QTL clusters were very valuable gene resources in molecular breeding to increase rice yield (Wang *et al.*, 2012a). Breeders can introgressing only one locus to enhance more yield-related traits instead

of pyramiding two or more locus, which needs more labour, time and money, and has higher risk of failure. However, some pleiotropic QTLs and QTL clusters had negative effects, which was unpopular. For instance, The QTLs qPH4, qPL4 and qSSR4.2 in the region of RM349-RM127 on chromosome 4 had pleiotropic effects on PH, PL, and SSR, in which the direction of gene effect of qSSR4.2 is opposite to that of qPH4 and qPL4. The region of RM467-RM3773 on chromosome 10 controls FGN, SSR and SN. The direction of gene effect on SN is opposite to that on FGN and SSR. Researcher can't simultaneously improve these three traits. Breeder should avoid using these gene recourses in high-yield breeding.

Implications for Molecular Breeding

Improving rice yield potential is undoubtedly the first goal of most rice breeding programs. However, yield-related traits are typical quantitative traits controlled by multigenes, how to efficiently select for high yield in breeding populations remains a large challenge. Fortunately, rice has a large number of germplasms, which are gene carriers with all kinds of alleles (Xing and Zhang, 2012). Identification and use of OTL especially pleiotropic OTLs and OTL clusters in rice molecular breeding can greatly speed up breeding proceeding. In the present study, 19 (54%) QTLs from the donor parent, IR75862 can increase vield-related traits. Among favorable alleles, IR75862 at qPL2.1 and qPL6.1 could improve SN and TGW due to pleiotropic effect or tight linkage, thus resulting in increase of grain yield. So, it's possible to develop new super rice varieties through introgressing and/ or pyramiding the IR75862 alleles at the pleiotropic QTLs (qPL2.1 and qPL6.1) or new QTLs (qGY3, qFGN10.2 and qSN10) into Ce258 and ZGX1 by molecular-assistant selection.

Acknowledgments

This work was supported by the National Natural Science Foundation of China (31261140369, 31371248), the Key Special Project on Molecular Design Breeding for Rice (2016YFD0101801) from the Chinese Ministry of Science & Technology, scientific research plan project of education department of Hubei (Q20151303), open founds of Hubei Collaborative Innovation Center for Grain Industry (2015MS008, LXT-16-01, LXT-17-08), National Key Laboratory of Crop Genetic Improvement (ZK201603), the Yangtze Youth Fund (2015cqn56), the Yangtze Fund for Youth Teams of Science and Technology Innovation (2015cqt02), Engineering Research Center of Ecology and Agricultural Use of Wetland, Ministry of Education, China (KF201508), and Hubei Key Laboratory of Food Crop Germplasm and Genetic Improvement (2014lzjj04).

Reference

- Ashikari, M., H. Sakakibara, S.Y. Lin, T. Yamamoto, T. Takashi, A. Nishimura and M. Matsuoka, 2005. Cytokinin Oxidase regulates rice grain production. *Science*, 309: 741–745
- Che, R.H., H.N. Tong, B.H. Shi, Y.Q. Liu, S.R. Fang, D.P. Liu, Y.H. Xiao, B. Hu, L.C. Liu, H.R. Wang, M.F. Zhao and C.C. Chu, 2015. Control of grain size and rice yield by *GL2*-mediated brassinosteroid responses. *Nat. Plants*, 2: 15195
- Chen, H., Y.Y. Tang, J.F. Liu, L.B. Tan, J.H. Jiang, M.M. Wang, Z.F. Zhu, X.Y. Sun and C.Q. Sun, 2017. Emergence of a novel chimeric gene underlying grain number in rice. *Genetics*, 205: 993–1002
- Cheng, L.R., Y. Wang, L.J. Meng, X. Hu, Y.R. Sun, L.H. Zhu, J. Ali, J.L. Xu and Z.K. Li, 2012. Identification of salt-tolerant QTLs with strong genetic background effect using two sets of reciprocal lines in rice. *Genome*, 55: 1–11
- Churchill, G.A. and R.W. Doerge, 1994. Empirical threshold values for quantitative trait mapping. *Genetics*, 138: 963–971
- Duan, P.G., S. Ni, J.M. Wang, B.L. Zhang, R. Xu, Y.X. Wang, H.Q. Chen, X.D. Zhu and Y.H. Li, 2015. Regulation of OsGRF4 by OsmiR396 controls grain size and yield in rice. Nat Plants, 2: 15203
- Dong, H.L., C.X. Zhang, B. Zeng and S.B. Yu, 2009. Identification of Agronomic traits QTL in common wild rice advanced backcrossed population. J. Huazhong Agric. Univ., 28: 645–650

- Fan, C.C., X.Z. Xing, H.L. Mao, T.T. Lu, B. Han, C.G. Xu, X.H. Li and Q.F. Zhang, 2006. GS3, a major QTL for grain length and weight and minor QTL for grain width and thickness in rice, encodes a putative transmembrance protein. *Theor. Apple. Genet.*, 112: 1164–1171
- FAO, 2009. *High Level Expert Forum- How to Feed the World in 2050*, 1st edition. Economic and social development department. Food and Agricultural Organization of the United State, Rome
- Fu, Q., P.J. Zhang, L.B. Tan, Z.F. Zhu, D. Ma, Y.C. Fu, X.C. Zhan, H.W. Cai and C.Q. Sun, 2010. Analysis of QTLs for yield-related traits in Yuanjiang common wild rice (*Oryza rufipogon* Griff). *J. Genet. Genom.*, 37: 147–157
- Fujita, D., K.R. Trijatmiko, A.G. Tagle, M.V. Sapasap, Y. Koide, K. Sasaki, N. Tsakirpaloglou, R.B. Gannaban, T. Nishimura, S. Yanagihara and N. Kobayshi, 2013. *NAL1* allele from a rice landrace greatly increases yield in modern indica cultivars. *Proc. Nat. Acad. Sci.*, 110: 20431–20436
- Hittalmani, S., N. Huang, B. Courtois, R. Venuprasad, H.E. Shashidhar, J.Y. Zhuang, K.L. Zheng, G.F. Liu, G.C. Wang, J.S. Sidhu, S. Srivantaneeyahul, V.P. Singh, P.G. Bagali, H.C. Prasanna, G. McLaren and G.S. Khush, 2003. Identification of QTLs for growthand grain yield-related traits in rice across nine locations in Asia. *Theor. Appl. Genet.*, 107: 679–690
- Hu J., Y.X. Wang, Y.X. Fang, L.J. Zeng, J. Xu, H.P. Yu, Z.Y. Shi, J.J. Pan, D. Zhang, S.J. Kang, L. Zhu, G.J. Dong, L.B. Guo, D.L. Zeng, G.H. Zhang, L.H. Xie, G.S. Xiong, J.Y. Li and Q. Qian, 2015. A rare allele of *GS2* enhances grain size and grain yield in rice. *Mol. Plant*, 8: 1455–1465
- Impa, S.M., M.J. Morete, A.M. Ismail, R. Schulin and S.E. JohnsonBeebout, 2013. Zn uptake, translocation and grain Zn loading in rice (Oryza sativa L.) genotypes selected for Zn deficiency tolerance and high grain Zn. J. Exp. Bot., 64: 2739–2751
- Ishimaru, K., N. Hirotsu, Y. Madoka, N. Murakami, N. Hara, H. Onodera, T. Kashiwagi, K. Ujiie, B. Shimizu, A. Onishi, H. Miyagawa and E. Katoh, 2013. Loss of function of IAA-glucose hydrolase gene *TGW6* enhances rice grain weight and increases yield. *Nat. Genet.*, 45: 707– 711
- Ishimaru, K., M. Yano, N. Aoki, K. Ono, T. Hirose, S.Y. Lin, L. Monna, T. Sasaki and R. Ohsugi, 2001. Toward the mapping of physiological and agronomic characters on a rice function map: QTL analysis and comparison between QTLs and expressed sequence tags. *Theor. Appl. Genet.*, 102: 793–800
- Jiang, L., X. Liu, G.S. Xiong, H.H. Liu, F.L. Chen, L. Wang, X.B. Meng, G.F. Liu, H. Yu, Y.D. Yuan, W. Yi, L.H. Zhao, H.L. Ma, Y.Z. He, Z.S. Wu, K, Meicher, Q. Qian, H.E. Xu, Y.H. Wang and J.Y. Li, 2013. DWARF 53 acts as a repressor of strigolactone signalling in rice. *Nature*, 504: 401–405
- Jiao, Y.Q., Y.H. Wang, D.W. Xue, J. Wang, M.X. Yan, G.F. Liu, G.J. Dong, D.L. Zeng, Z.F. Lu, X.D. Zhu, Q. Qian and J.Y. Li, 2010. Regulation of *OsSPL14* by OsmiR156 defines ideal plant architecture in rice. *Nat. Genet.*, 42: 541–545
- Kobayashi, S., Y. Fukuta, T. Sato, M. Osaki and G.S. Khush, 2003. Molecular marker dissection of rice (*Oryza sativa* L.) plant architecture under temperate and tropical climates. *Theor. Appl. Genet.*, 107: 1350–1356
- Kumar, K., M. Kumar, S.R. Kim, H. Ryu and Y.G. Cho, 2013. Insight into genomics of salt stress response in rice. *Rice*, 6: 27
- Li, H.H., G.Y. Ye and J.K. Wang, 2007. A modified algorithm for the improvement of composite interval mapping. *Genetics*, 175: 361– 374
- Li, X.Y., Q. Qian, Z.M. Fu, Y.H. Wang, G.G. Xiong, D.L. Zeng, X.Q. Wang, X.F. Liu, S. Teng, F. Hiroshi, M. Yuan, D. Luo, B. Han and J.Y. Li, 2003. Control of tillering in rice. *Nature*, 422: 618–621
- Li, Y.B., C.C. Fan, Y.Z. Xing, Y.H. Jiang, L.J. Luo, L. Sun, D. Shao, C.J. Xu, X.H. Li, J.H. Xiao, Y.Q. He and Q.F. Zhang, 2011. Natural variation in GS5 plays an important role in regulating grain size and yield in rice. *Nat. Genet.*, 43: 1266–1269
- Liao, C.Y., P. Wu, B. Hu and K.K. Yi, 2001. Effects of genetic background and environment on QTLs and epistasis for rice (*Oryza sativa* L.) panicle number. *Theor. Appl. Genet.*, 103: 104–111

- Mao, H.L., S.Y. Sun, J.L. Yao, C.R. Wang, S.B. Yu, C.G. Xu, X.H. Li and Q.F. Zhang, 2010. Linking differential domain functions of GS3 protein to natural variation of grain size in rice. *Proc. Nat. Acad. Sci.*, 107: 19579–19584
- McCouch, S.R., L. Teytelman, Y.B. Xu, K.B. Lobos, K. Clare, M. Walton, B.Y. Fu, R. Maghirang, Z.K. Li, Y.Z. Xing, Q.F. Zhang, I. Kono, M. Yano, R. Fjellstrom, G. DeClerck, D. Schneider, S. Cartinhour, D. Ware and L. Stein, 2002. Development and mapping of 2240 new SSR markers for rice (*Oryza sativa* L.). *DNA Res.*, 9: 199–207
- Mei, H.W., J.L. Xu, Z.K. Li, X.Q. Yu, L.B. Guo, Y.P. Wang C.S. Ying and L.J. Luo, 2006. QTLs influencing panicle size detected in two reciprocal introgressive line (IL) populations in rice (*Oryza sativa* L.). *Theor. Appl. Genet.*, 112: 648–656
- Miura, K., M. Ikeda, A. Matsubara, S.J. Song, M. Ito, K. Asano, M. Matsuoka, H. Kitano and M. Ashikari, 2010. OsSPL14 promotes panicle branching and higher grain productivity in rice. Nat. Genet., 42: 545–549
- Morales, M., 2001. Statistica, Version 5.5. Bulletin of the Ecological Society of America, 82: 126–127
- Murray, M.G. and W.F. Thompson, 1980. Rapid isolation of high molecular-weight plant DNA. *Nucl. Acid. Res.*, 8: 4321–4325
- Peng, B., L.Q. Wang, C.C. Fan, G.H. Jiang, Y.B. Li and Y.Q. He, 2014. Comparative mapping of chalkiness components in rice using five populations across two environments. *BMC Genet.*, 15: 49
- Qiu, X.J., R. Gong, Y.B. Tan and S.B. Yu, 2012. Mapping and characterization of the major quantitative trait locus *qSS7* associated with increased length and decreased width of rice seeds. *Theor. Appl. Genet.*, 125: 1717–1726
- Qiu, X.J., Z.H. Yuan, H. Liu, X.J. Xiang, L.W. Yang, W.J. He, B. Du, G.Y. Ye, J.L. Xu and D.Y. Xing, 2015. Identification of salt tolerance-improving quantitative trait loci alleles from a saltsusceptible rice breeding line by introgression breeding. *Plant Breed.*, 134: 653–660
- Shen, G.J., W. Zhan, H.X. Chen and Y.Z. Xing, 2014. Dominance and epitasis are the main contributors to heterosis for plant height in rice. *Plant Sci.*, 215–216: 11–18
- Shomura, A., T. Izawa, K. Ebana, T. Ebitani, H. Kanegae, S. Konishi and M. Yano, 2008. Deletion in a gene associated with grain size increased yields during domestication. *Nat. Genet.*, 40: 1023–1028
- Song, X.J., W. Huang, M. Shi, M.Z. Zhu and H.X. Lin, 2007. A QTL for rice grain width and weight encodes a previously unknown RINGtype E3 ubiquitin ligase. *Nat. Genet.*, 39: 623–630
- Sun, P.Y., W.H. Wang, Y.H. Wang, Q. He, F. Shu, H. Liu, J. Wang, J.M. Wan, L.P. Yuan and H.F. Deng, 2016. OsGRF4 controls grain shape, panicle length and seed shattering in rice. J. Integr. Plant Biol., 2016, 58: 836–847
- Tan, C., Z.M. Han, H.H. Yu, W. Zhan, W.B. Xie, X. Chen, H. Zhao, F.S. Zhou and Y.Z. Xing, 2013. QTL scanning for rice yield using a whole genome SNP array. J. Genet. Genom., 40: 629–638
- Temnykh, S., G. DeClerck, A. Lukashova, L. Lipovich, S. Cartinhour and S. McMouch, 2001. Computational and experimental analysis of microsatellites in rice (*Oryza sativa* L.): frequency, length variation, transposon associations, and genetic marker potential. *Genom. Res.*, 11: 1441–1452
- Wang, P., Y.Z. Xing, Z.K. Li and S.B. Yu, 2012a. Improving rice yield and quality by QTL pyramiding. *Mol. Breed.*, 29: 903–913
- Wang, S.K., L. Shan, Q. Liu, K. Wu, J.Q. Zhang, S.S. Wang, Y. Wang, X.B. Chen, Y. Zhang, C.X. Gao, F. Wang, H.X. Huang and X.D. Fu, 2015a. The OsSPL16-GW7 regulatory module determines grain shape and simultaneously improves rice yield and grain quality. Nat. Genet., 47: 949–954
- Wang, S.K., K. Wu, Q.B. Yuan, X.Y. Liu, Z.B. Liu, X.Y. Lin, R.Z. Zeng, H.T. Zhu, G.J. Dong, Q. Qian, G.Q. Zhang and X.D. Fu, 2012b. Control of grain size, shape and quality by *OsSPL16* in rice. *Nat. Genet.*, 44: 950–954
- Wang, Y., J.P. Zang, Y. Sun, J. Ali, J.L. Xu and Z.K. Li, 2013. Background-independent quantitative trait loci for drought tolerance identified using advanced backcross introgression lines in rice. *Crop Sci.*, 53: 430–441

- Wang, Y.X., G.S. Xiong, J. Hu, L. Jiang, H. Yu, J. Xu, Y.X. Fang, L.J. Zeng, E.B. Xu, J. Xu, W.J. Ye, X.B. Meng, R.F. Liu, H.Q. Chen, Y.H. Jing, Y.H. Wang, X.D. Zhu, J.Y. Li and Q. Qian, 2015b. Copy number variation at the GL7 locus contributes to grain size diversity in rice. *Nat. Genet.*, 47: 944–948
- Wang, Y., Q. Zhang, T.Q. Zheng, Y.R. Cui, W.Z. Zhang, J.L. Xu and Z.K. Li, 2014. Drought-tolerance QTLs commonly detected in two sets of reciprocal introgression lines in rice. *Crop Pasture Sci.*, 65: 171– 184
- Wei, D., K.H. Cui, J.F. Pan, Q. Wang, K. Wang, X.M. Zhang, J. Xiang, L.X. Nie and J.L. Huang, 2012. Identification of quantitative trait loci for grain yield and its components in response to low nitrogen application in rice. *Aust. J. Crop Sci.*, 6: 986–994
- Wei, X.J., J.F. Xu, H.N. Guo, L. Jiang, S.H. Chen, C.Y. Yu, Z.L. Zhou, P S. Hu, H.Q. Zhai and J.M. Wan, 2010. *DTH8* suppresses flowering in rice, influencing plant height and yield potential simultaneously. *Plant Physiol.*, 153: 1747–1758
- Weng, J.F., S.H. Gu, X.J. Wan, H. Gao, T. Guo, N. Su, C.L. Lin, X. Zhang, Z.J. Cheng, X.P. Guo, J.L. Wang, L. Jiang, H.Q. Zhai and J.M. Wan, 2008. Isolation and initial characterization of *GW5*, a major QTL associated with rice grain width and weight. *Cell Res.*, 18: 1199– 1209
- Wu, Y., Y. Wang, X.F. Mi, J.X. Shan, X.M. Li, J.L. Xu and H.X. Lin, 2016. The QTL GNP1 encodes GA20ox1, which increases grain number and yield by increasing Cytokinin activity in rice panicle meristems. PLos Genet., 12: e1006386
- Xiao, J., J. Li, L. Yuan and S.D. Tansley, 1996. Identification of QTLs affecting traits of agronomic importance in a recombinant inbred population derived from a subspecific rice crosses. *Theor. Appl. Genet.*, 92: 230–244
- Xie, X.W., M.R. Xu, J.P. Zang, Y. Sun, L.H. Zhu, J.L. Xu, Y. L. Zhou and Z.K. Li, 2008. Genetic background and environmental effects on expression of QTL for sheath blight resistance in reciprocal introgression lines of rice. *Acta Agron. Sin.*, 34: 1885–1893
- Xing, Y.Z. and Q.F. Zhang, 2012. Genetic and molecular bases of rice yield. Annu. Rev. Plant Biol., 61: 11.1-11.22
- Xing, Y.Z., Y.F. Tan, J.P. Hua, X.L. Sun, C.G. Xu and Q.F. Zhang, 2002. Characterization of the main effects, epistasis effects and their environmental interactions of QTLs on the genetic basis of yield traits in rice. *Theor. Appl. Genet.*, 105: 248–258
- Xue, W.Y., Y.Z. Xing, X.Y. Weng, Y. Zhao, W.J. Tang, L. Wang, H.J. Zhou, S.B. Yu, C.G. Xu, X.H. Li and Q.F. Zhang, 2008. Natural variation in *Ghd7* is an important regulator of heading date and yield potential in rice. *Nat. Genet.*, 40: 761–767
- Yan, W.H., H.Y. Liu, X.C. Zhou, Q.P. Li, J. Zhang, L. Lu, T.M. Liu, H.J. Liu, C.J. Zhang, Z.Y. Zhang, G.J. Shen, W. Yao, H.X. Chen, S.B. Yu, W.B. Xie and Y.Z. Xing, 2013. Natural variation in *Ghd7.1* plays an important role in grain yield and adaptation in rice. *Cell Res.*, 23: 969–971
- Yan, W.H., P. Wang, H.X. Chen, H.J. Zhou, Q.P. Li, C.R. Wang, Z.H. Ding, Y.S. Zhang, S.B. Yu, Y.Z. Xing and Q.F. Zhang, 2011. A major QTL, *Ghd8*, plays pleiotropic roles in regulating grain productivity, plant height, and heading date in rice. *Mol. Plant*, 4: 319–330
- Ye, S.P., Q.J. Zhang, J.Q. Li, B. Zhao and P. Li, 2005. QTL mapping for yield component traits using (Pei' ai 64s/ Nipponbare) F₂ population. *Acta Agron. Sin.*, 31: 1620–1627
- Yousef, G.G. and J.A. Juvik, 2002. Enhancement of seedling emergence in sweet corn by marker-assisted backcrossing of beneficial QTL. Crop Sci., 42: 96–104
- Yu, S.B., J.X. Li, C.G. Xu, Y.F. Tan, Y.J. Gao, X.H. Li, Q.F. Zhang and M.A.S. Maroof, 1997. Importance of epistasis as the genetic basis of heterosis in an elite rice hybrid. *Proc. Nat. Acad. Sci.*, 94: 9226–9231
- Zhang, C.X., X.J. Qiu, H.L. Dong and S.B. Yu, 2010. Development and characterization of chromosome segment substitution lines using *O. rufipogon* as donor. *Molecular Plant Breed.*, 8: 1113–1119
- Zhang, F., Y.Z. Jiang, S.B. Yu, J. Ali, A.H. Paterson, G.S. Khush, J.L. Xu, Y.M. Gao, B.Y. Fu, R. Lafitte and Z.K. Li, 2013. Three genetic systems controlling growth, development and productivity of rice (*Oryza sativa L.*): a reevaluation of the "Green Revolution". *Theor. Appl. Genet.*, 126: 1101–1024

- Zhang, Q.F., 2007. Strategy for developing green super rice. Proc. Natl. Acad. Sci., 104: 16402–16409
- Zhang, Y.S., L.J. Luo, T.M. Liu, C.G. Xu and Y.Z. Xing, 2009. Four rice QTLs controlling number of spikelets per panicle expressed the characteristics of single Mendelian gene in near isogenic backgrounds. *Theor. Appl. Genet.*, 118: 1035–1044
- Zhao, J.G., K.F. Jiang, Y. Li, Q.H. Yang, X.Q. Wan, Y.J. Cao, S.M. You, J. Luo, T. Zhang and J.K. Zheng, 2013. QTL mapping for yield components in a RIL population of rice. *Chin. J. Rice Sci.*, 27: 344– 352
- Zhao, X.Q., V.D. Daygon, K.L. McNally, R.S. Hamilton, F.M. Xie, R.F. Reinke and M.A. Fitzgerald, 2016. Identification of stable QTLs causing chalk in rice grains in nine environments. *Theor. Appl. Genet.*, 129: 141–153
- Zheng, T.Q., Y. Wang, A.J. Ali, L.H. Zhu, Y. Sun, H.Q. Zhai, H.W. Mei, Z.J. Xu, J.L. Xu and Z.K. Li, 2011. Genetic effects of backgroundindependent loci for grain weight and shape identified using advanced reciprocal introgression lines from Lemont × Teqing in rice. Crop Sci., 51: 2525–2534
- Zhou, F., Q.B. Lin, L.H. Zhu, R.L. Ren, K.N. Zhou, N. Shabek, F.Q. Wu, H.B. Mao, W. Dong, L. Gan, W.W. Ma, H. Gao, J. Chen, C. Yang, D. Wang, J.J. Tan, X. Zhang, X.P. Guo, J.L. Wang, L. Jiang, X. Liu, W.Q. Chen, J.F. Chu, C.Y. Yan, K. Ueno, S. Ito, T. Asami, Z.J. Cheng, J. Wang, C.L. Lei, H.Q. Zhai, C.Y. Wu, H.Y. Wang, N. Zheng and J.M. Wan, 2013. D14-SCF^{D3}-dependent degradation of D53 regulates strigolactone signalling. *Nature*, 504: 406–410

(Received 19 September 2016; Accepted 04 January 2017)