

# Biochemical Changes and Pathogenicity Variations Related to Esterase Polymorphism for Morphological Traits of *Rhizocotonia solani*

AMANY H. ABO ELLIL

Botany Department, Faculty of Science, Cairo University, Cairo, Egypt

Corresponding author's email: amanyaboellil@yahoo.com

## ABSTRACT

Four morphological traits R<sub>1</sub>, R<sub>2</sub>, R<sub>3</sub> and R<sub>4</sub> of *Rhizocotonia solani* obtained by growing on different nitrogen sources were determined for some biochemical characteristics. A further comparative study was the impact of melanin on survival and pathogenicity of *R. solani*, these parameters were used to estimate the total fitness for each morphological traits. It seems that these variations were due to the development of biotypes. In the present study two isolates of *R. solani* were obtained: one of which is melanin rich R<sub>1</sub> (Dark) showed the most sclerotial density and the most virulent to *Phaseolus vulgaris*, while the other is melanin poor R<sub>4</sub> (Albino) showed the least sclerotial density and the least virulent. Esterase isoenzyme was used as a molecular marker to discriminate the two morphological traits, which showed highly significant biochemical variation. Two phenotypes evaluated through polyacrylamide gel electrophoresis (PAGE) for isoenzyme pattern for esterase in relation to color of fungus, pathogenicity, and its sclerotial density. Specific bands on PAGE could be associated with each of these phenotypes. In addition to presence of particular banding pattern, intensity of certain bands also helped in characterization of phenotype. Esterase is considered as an effective discriminating system in characterizing phenotypes and to detect the genetic changes involved.

**Key Words:** *Rhizocotonia solani*; Nitrogen; Melanin; Sugar; Disease index; Esterase; Phenotype; Biotype; Pathotype

## INTRODUCTION

*Rhizocotonia solani* is a highly damaging fungus with a wide host range. Sclerotia are produced among the hyphae on the surface of infected plant. These sclerotia appear as minute black dots and measure on an average up to 100 µm in diameter (Pandy, 1982).

The role of melanin as a virulence factor in pathogenicity is confirmed by HeungTae *et al.* (2001), Kurahashi (2001), HeungTae *et al.* (2002) and Solomon *et al.* (2004) in plant pathogens *Magnaporthe grisea*, *Rhizocotonia solani* and *Stagonospora nodorum*. However, some fungi including *Rhizocotonia solani*, *Sclerotium rolfsii*, *Magnaporthe grisea* etc. grow on melanin (Richard, 1993; Deacon, 1997; Ellil, 1999; Kurahashi, 2001) and therefore their growth could be controlled by chemicals that specifically block the pathway of melanin biosynthesis. Recent studies have increased our understanding of the chemical and physiological properties of melanin (Henson, 2001). Compounds that interfere with its synthesis or promote its degradation may help in preventing plant disease (Leroux, 2003).

Costa *et al.* (2004) studied the effect of different nitrogen sources on glucose uptake, production of melanin precursors and fungal mass of *Foinsecaea pedrosoi* cultured in tricyclazole. Changing nitrogen sources, added as supplements to a basic liquid mineral medium, affected the color and characteristics of the pigment; the dark pigment was melanin, which was produced by *Ophiostoma piceae*. The mycelia became colored early during the active growth phase and attaining maximum color intensity during

stationary growth phase (Eagen *et al.*, 1997). As the main nitrogen source in *Malassezia furfur*, tryptophan induced the formation of pigments, which made the yeast less sensitive to UV light (Mayser *et al.*, 2002).

Expression of pKS4 (polyketide synthase gene) is responsible for the first step of the red polyketide pigment bikaverin and repressed by high amounts of ammonium basic pH. Unexpectedly, pKS4 was overexpressed in mutants of the regulatory gene A, which is responsible for the activation of nitrogen assimilation genes (Linnemannstons *et al.*, 2002). An extracellular esterase from *Candida albicans* A714 was induced in a medium containing 0.7% yeast nitrogen base and tween 80 (Tsuboi *et al.*, 1996).

Complex nitrogen sources such as tryptone or yeast extract increased growth and esterase production while mineral sources (ammonium chloride or sulfate), glycine or glutamate showed no effect. An increase of tryptone plus yeast extract and glucose concentrations stimulated growth and esterase production of *Bacillus circulans* which reached 160 µL<sup>-1</sup> (Kademi *et al.*, 1999). Media containing yeast extract as nitrogen source was found to be optimal for extracellular esterase (221 µdm<sup>-3</sup>) from the fungus *Ophiostoma piliferum*. Further increase on these enzyme activities was achieved by decreasing medium pH from 6.5 to 5.5 (George *et al.*, 1999).

A few studies were aimed at interconversion between melanin and esterase and pathogenicity (Jwa & Chung, 1993; Kuc, 1997 and Kuc *et al.*, 1999) and its possible role in disease resistance. Four morphological traits of *R. solani*

from previous work (grown on basal media supplemented with different nitrogen sources) were named as R<sub>1</sub>, R<sub>2</sub>, R<sub>3</sub> and R<sub>4</sub> according to their color (Table I) and were tested against nitrogen, phosphorus, sugar uptake, phenols and melanin content. The impact of melanin on pathogenicity was further studied. Esterase isoenzyme analyses were used as molecular markers to discriminate morphological traits and related to the esterase patterns.

## MATERIALS AND METHODS

Culture of *Rhizocotonia solani* was kindly provided by Central Laboratory, Plant Pathology Institute, Center of Agricultural Research, Cairo, Egypt. *R. solani* was grown on modified basal medium (Ko & Hara, 1971) containing the following ingredients: 10g sucrose; 0.05g CaCl<sub>2</sub>; 0.025g NaCl; 0.25g (NH<sub>4</sub>) HPO<sub>4</sub>; 0.5g KH<sub>2</sub>PO<sub>4</sub>; 0.15g MgSO<sub>4</sub> · 7 H<sub>2</sub>O; 1.0 ml FeCl<sub>3</sub> (1% w/v) 25µg thiamine HCL; 2.5 g malt extract and distilled water up to 1000ml. The experimental medium was the basal liquid medium with changed nitrogen sources in the medium using NaNO<sub>3</sub> (4g L<sup>-1</sup>), peptone (5g L<sup>-1</sup>) and malt extract (2.5g L<sup>-1</sup>).

**Determination of phenols in filtrate.** Phenolic compounds were determined according to Mahto *et al.* (1987). The free phenols were determined as mg catechol mL<sup>-1</sup> filtrate from the standard curve of catechol. Total phenols were determined after boiling the sample with concentrated HCl in a water-bath for 10 min. conjugated phenols were determined by subtracting the amount of free phenols from that of total phenols.

**Determination of total soluble nitrogen, total soluble phosphorus and total soluble sugars.** Samples were assayed using the method of Fawcett and Scott (1960) for total soluble nitrogen and Burton and Riley (1954) method for total soluble phosphorus. Soluble sugars were estimated by the Dubois *et al.*, (1956) method.

**Determination of ion concentration.** To determine ion concentrations the samples were oven-dried, ground in a Wiley mill and ashed at 485°C for 10-12 h. Ash was equilibrated with 2 M HCl at room temperature and analyzed by inductively coupled plasma atomic emission spectroscopy (ICPAES), model ARL 3560 (Dahlquist & Knoll, 1987 and Karla & Maynerd, 1991). Elemental concentration was represented as micrograms per gram dried mycelia.

**Isolation and characterization of melanin.** The isolation of melanin from the fungus was carried out according to the method of Ellis and Griffith (1975). A 20g of air dried ground material were used for melanin extraction. Because of rapid oxidation of the pigment in air, extraction and analysis were carried out under nitrogen. Weights were determined for samples dried at 105°C. The extraction of melanin was ensured based on their (1) solubility in water, (2) color, (3) solubility in MKOH 100°C; 2 hrs, (4) precipitation in HCl, (5) solubility in organic solvents, (6) reaction with oxidant (NaOCl and H<sub>2</sub>O<sub>2</sub>), (7) gradient of log absorbance in visible light 400-600 nm and (8) IR

spectrum. Melanization was monitored at 400 nm as the incremental optical density (O.D.). Cultures with melanization beyond an O.D. of 1 were diluted 10 folds and measured against a similarly dilute reference culture (Nicolaus *et al.*, 1964).

**Test for the degree of pathogenicity.** This method was adopted from Kloepper (1991). Seeds of *Phaseolus vulgaris* were surface sterilized with sodium hypochlorite (2%) for 2 min. The surface sterilized seeds were soaked for 2 h in autoclaved distilled water. Seeds were then placed on sterile wet cotton, 5 seeds per plate (9 cm in diameter) and incubated at 30°C for 48 hrs until emergence of radical. The germinating seeds 5 per plate were placed over 7 days old culture of the tested isolates of *R. solani* grown on different nitrogen sources. A 40g of sieved fine particles of natural soil (autoclaved for 2 hrs at 121°C and 1.5 Bar) were spread over each plate. Each plate was then moistened with 15 mL sterile distilled water. Plates were covered and incubated at 30°C until the seedling had grown up, after which the plate cover was removed (after 72 h approximately). Results were recorded as disease scale according to the following index as reported by Kloepper (1991). 0: white root without brown patches; 1: yellow root; 2: yellow root with brown patches; 3: yellow root with brown patches and surface lesions; 4: deep lesions and preliminary symptoms of root rot; 5: brown discoloration, deep lesions, and absence of fungal mycelium; 6: brown discoloration, deep lesions and obvious fungal mycelium on root surface; 7: root is completely rotted (dead).

**Determination of esterase.** The mycelium was harvested from each culture by filtration and washed in sterile distilled water followed by homogenization in an extraction medium of 0.2 M-tris citrate buffer (pH 8.3). Extracts were absorbed into pieces of Whatmann No 3 filter paper (0.5x0.1mm), which were inserted in a slot made 3 cm away from cathode of the gels. The electrophoresis was performed in a discontinuous system as described by Scandalias (1996) for acrylamide gel. The gel staining and fixation for esterases were carried out according to Steiner and Toslyn (1979).

All estimations were carried out in triplicates. The arithmetic means were tabulated and the least significant difference (LSD) at 0.1% level confidence limits was calculated.

## RESULTS

Four isolates of *R. solani* grown on different nitrogen sources (Table I) were: R<sub>1</sub> grown on sodium nitrate, the 1<sup>st</sup> isolate very dark in color, medium growth and high sclerotia production; R<sub>2</sub> grown on peptone, the 2<sup>nd</sup> isolate brown in color, high in growth and medium in number of sclerotia; R<sub>3</sub> with light brown on malt and ammonium phosphate as nitrogen source gave less growth and less number of sclerotia and R<sub>4</sub> grown on malt only is white in color (albino) represented the least growth and the least sclerotial production. Therefore, the highest density of sclerotia was recorded in R<sub>1</sub> (dark) while the significant drop was

recorded in R<sub>2</sub> (albino) isolates of *R. solani*.

The highest production of total phenol and conjugated phenol was recorded in filtrate of R<sub>4</sub> (albino) (Table II). Contrarily, the amounts of total, free and conjugated phenol (dark) were the lowest in R<sub>1</sub>. The amount of the 3 forms of phenols was higher in filtrate of R<sub>2</sub> than in that of R<sub>3</sub>. Table (III) showed that sugars (total and reducing), total nitrogen and total phosphorus were accumulated in the filtrate of R<sub>4</sub> (albino) with highest significance than that in filtrate of others. R<sub>1</sub> filtrate recorded the least significant accumulation of these parameters.

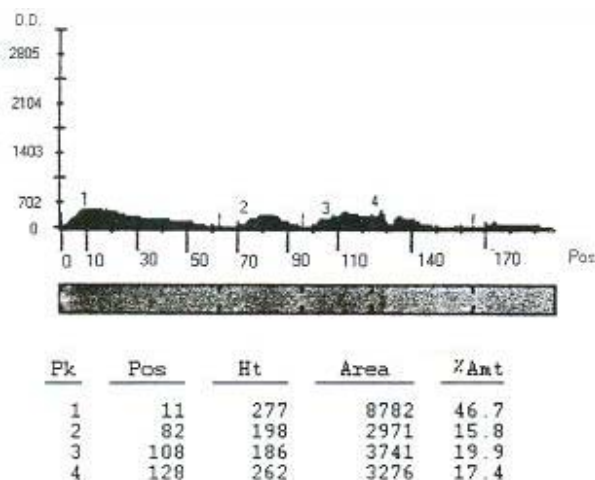
Table (IV) shows the degree of pathogenicity of 4 isolates of *R. solani* infecting *Phaseolus vulgaris* seedlings according to disease index mentioned by Kloepper (1991). It was obvious that R<sub>1</sub> (dark isolate) was the most virulent; R<sub>2</sub> (brown) more virulent than the light brown R<sub>3</sub>, while R<sub>4</sub> (albino isolate) the least virulent. Melanin pigment showed the same trend, while R<sub>1</sub> represented the highest amount of melanin accumulation R<sub>4</sub> recorded the lowest amount of it. Further comparison on the biomass and other characteristics of 2 isolates the most virulent dark isolate (R<sub>1</sub>) and the least virulent albino isolate (R<sub>4</sub>) is given in Table (V). The higher amounts of phenol and phosphate ions were recorded in R<sub>1</sub> biomass than that in R<sub>4</sub> biomass while the total dissolved salts were higher in R<sub>4</sub> than R<sub>1</sub>. The SO<sub>4</sub><sup>2-</sup> and Cl<sup>-</sup> concentrations were similar in both. Concentrations of K<sup>+</sup> and Na<sup>+</sup> were higher in R<sub>1</sub>, while those of divalent cations of Cu<sup>2+</sup>, Mg<sup>2+</sup> and Fe<sup>2+</sup> were in higher quantities in R<sub>4</sub> than that in R<sub>1</sub> biomass.

Both R<sub>1</sub> and R<sub>4</sub> were analyzed for esterase isoenzymes by SDS-PAGE electrophoresis of esterase gave 4 bands to each (Fig. 1 & Fig. 2). Electrophoresis revealed variation in position width and intensity of bands. Analysis of esterase pattern of R<sub>1</sub> has shown 4 distinct bands (Fig. 1) but for R<sub>4</sub> 3 distinct bands were presented and a weak fourth band appeared, which was the only band similar to that of R<sub>1</sub> (Fig. 2).

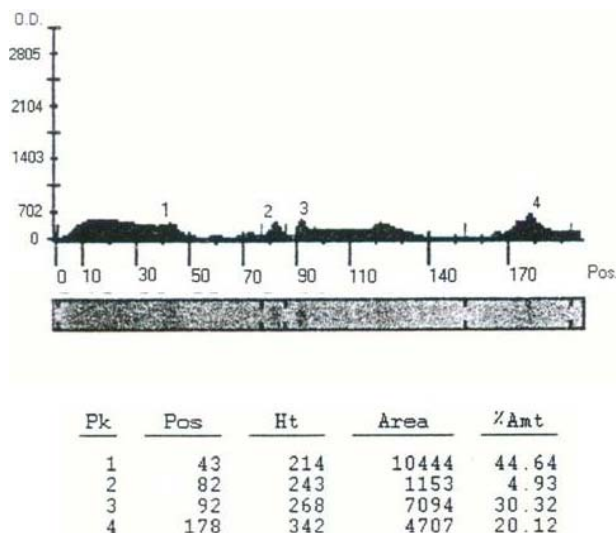
## DISCUSSION

Several authors have suggested that N-sources can influence sclerotium production (Willems, 1978; Robert, 1989). Mayser *et al.* (2002) and Eagen *et al.* (1997) confirmed that different nitrogen sources added as supplements to a basic liquid mineral medium affected the color and characteristics of pigment or induced the formation of pigment. Among the different nitrogen, sodium nitrate caused a highly significant increase in growth and number and density of sclerotia (Table I). The darkest pigmentation of sclerotia and hyphae was observed on sodium nitrate indicating an enhanced melanin formation. Accordingly, this isolate was nominated as dark R<sub>1</sub>. Contrarily, using malt as a sole nitrogen source in the basal medium stopped pigmentation of the hyphae and reduced the growth and the sclerotia formation. Accordingly, this isolate was nominated as albino R<sub>4</sub>. Other nitrogen sources had the same trend but with different degree. Kusz *et al.*

**Fig. 1. Esterase pattern obtained by electrophoresis on polyacrylamide gel of extract of dark isolate R<sub>1</sub> of *R. solani* after 14 days growth on liquid media containing NaNO<sub>3</sub> as a sole nitrogen source**



**Fig. 2. Esterase pattern obtained by electrophoresis on polyacrylamide gel of extract of albino isolate R<sub>4</sub> of *R. solani* after 14 days growth on liquid media containing malt as a sole nitrogen source**



(2001) noted the direct or indirect free radical character of nitrate. This most probably is due to accumulation of high amount of organic acids through the course of assimilation of this complex natural nitrogen e.g. malt, peptone (Griffin, 1994). Williams *et al.* (2000) suggested that phenol oxidase activity might be constrained by low pH. Also, in a fresh water prawn, Cheng *et al.* (2000) found that its phenol oxidase activity was highest at pH 7.5-7.7.

In filtrate of albino isolate (R<sub>4</sub>), the highest significant total phenol and conjugated phenol were recorded, the opposite was in dark isolate (R<sub>1</sub>) (Table II). Use of organic nitrogen (tryptone, beef extract and yeast extract) yielded more phenol than the inorganic nitrogen sources by

**Table I. Color, dry weight and number of sclerotia of *R. solani* isolates grown on different nitrogen sources**

Isolate	Nitrogen source*	Final color of isolate	Final air dry weight (mg)	Final number of scleration
R <sub>1</sub>	Sodium nitrate (4g L <sup>-1</sup> )	Very dark	250.5**	154.2**
R <sub>2</sub>	Peptone (5g L <sup>-1</sup> )	Brown	378.3**	102.4**
R <sub>3</sub>	Malt ammonium phosphate (2.5g L <sup>-1</sup> + 0.25f L <sup>-1</sup> )	Light brown	173.1	68.3
R <sub>4</sub>	Malt (2.5g L <sup>-1</sup> )	White	161.2*	58.0*

LSD at 1% \* Significant, \*\* Highly significant (Concentrations of nitrogen source were calculated on equimolecular nitrogen concentration base).

*Botryodipodia theobromae* in culture (Mahapatra *et al.*, 2001).

Accumulation of total sugar, nitrogen and phosphorus was more in filtrate of R<sub>4</sub> culture than the others, while R<sub>4</sub> filtrate recorded the least accumulation of the previous compounds (Table III). A negative correlation between biomass production for *Fonsecaea pedrosori* and glucose uptake was observed in the presence of phenol or melanin accumulation, according to different nitrogen source supplementation (Costa *et al.*, 2004).

The impact of melanin on pathogenicity in the work was also studied and was proved to be a virulence factor. This could be realized by the use of all the 4 isolates aiming to study their virulence experimentally. R<sub>4</sub> failed to produce a high disease index as did the melanized isolates on pathogenicity are noteworthy (Table IV). Such behavior assures the role played by melanin as a pathogenicity factor that may lead to the development of “pathotype” within *R. solani*. The role of melanin as a virulence factor in pathogenicity is well documented (Kurahashi, 2001; Solomon *et al.*, 2004).

The oddest morphological variants were obtained. Avirulent and melanin rich designated R<sub>1</sub> was obtained by growing the fungus *R. solani* on NaNO<sub>3</sub> as a sole source of nitrogen source. This variant, when compared with that less virulent and melanin poor (R<sub>4</sub>) grown on malt also showed biochemical changes. These changes displayed an increase in growth and sclerotia number, phenol and phosphate accumulations in R<sub>1</sub> than R<sub>4</sub> biomass. Also sodium and potassium were more in R<sub>1</sub> than R<sub>4</sub>. The total dissolved salts, copper, magnesium and ferrous were higher in R<sub>4</sub> isolate as compared with R<sub>1</sub> (Table V). It seems likely, therefore, that this behavior is due to the development of “biotypes” within *R. solani* grown on different nitrogen sources. Scott (1981) stated that the differences in the behavior of phenotypes of *Sclerotinia trifoliorum* in pathogenicity are due to the development of “biotypes” within it, which are particularly virulent on white clover. The dendrogram of esterase isoenzyme of the two morphological traits showed less than 25% similarity. Similar results were obtained by Lloyd *et al.* (1972) in *Aspergillus niger* where modifications in esterase patterns were detected when growing a strain in different media, some of which prevented conidia formation.

Esterases are often used to measure genetic variation, yet they may be influenced by external factors (Perrotey *et al.*, 2002). Isoelectric focusing was used to determine the effect of different nitrogen sources on the esterase variation in dark and albino phenotypes (isolates) of *R. solani* (Mohammadi *et al.*, 2003). Results from esterase isoenzyme

analysis for *R. solani* suggested that the morphological traits (subgrouping concept) is genetically based (Fig. 1 & 2). This morphological variant may be due probably to one or more genetic blocks used. It was shown that the morphological trait was related to the absence or presence of one enzyme.

**Table II. Amount of phenols in filtrate of 4 isolates of *R. solani* grown on different nitrogen sources**

Isolate	Total phenol (mg L <sup>-1</sup> )	Free phenol (mg L <sup>-1</sup> )	Conjugated phenol (mg L <sup>-1</sup> )
R <sub>1</sub>	28.8 **	24.5 *	4.3 *
R <sub>2</sub>	64.5 *	29.4 **	35.1 *
R <sub>3</sub>	30.9 *	26.4	4.5
R <sub>4</sub>	133.5 **	28.2 **	95.3 **

LSD at 1% \* Significant, \*\* Highly significant

**Table III. Estimation of sugars, total nitrogen and total phosphorus in filtrate of 4 isolates of *R. solani* grown on different nitrogen sources**

Isolate	Sugars (mg L <sup>-1</sup> )			Total nitrogen (mg L <sup>-1</sup> )	Total phosphorus (mg L <sup>-1</sup> )
	Total	Reducing	Non reducing		
R <sub>1</sub>	15.8**	5.3 *	10.5 *	6.7 *	5.9 *
R <sub>2</sub>	27.5 *	7.2	15.3 *	19.1 **	5.7 *
R <sub>3</sub>	27.9 *	7.0	20.9 **	15.6 **	4.3 **
R <sub>4</sub>	67.9**	65.5 **	2.9 **	22.4 **	6.3 **

L.S.D. at 1% \* Significant, \*\* Highly significant

**Table IV. Degree of pathogenicity and the melanin content (mg/g) of 4 isolates of *R. solani* grown on different nitrogen sources**

Isolate	Color	Disease index	Melanin content (mg g <sup>-1</sup> )
R <sub>1</sub>	Very dark	7 **	140 **
R <sub>2</sub>	Brown	6 *	110 *
R <sub>3</sub>	Light brown	5 **	70 *
R <sub>4</sub>	White (albino)	2 **	10 **

L.S.D. at 1% \* Significant, \*\* Highly significant

**Table V. Biomass analysis of 2 isolates R<sub>1</sub> (dark) and R<sub>4</sub> (albino) for phenols, some ions and total dissolved salts (TDS)**

Chemical characteristics	R <sub>1</sub> (melanin rich)		R <sub>4</sub> (melanin poor)
Phenol (mg/g)	14.6	>	3.2
Sulphate (mg/g)	126	=	120
Orthophosphate (mg/g)	134.4	>	92.8
Chloride (mg/g)	35.5	=	35.5
Copper (mg/g)	6.407	<	15.23
Potassium (mg/g)	332.8	>	170.6
Sodium (mg/g)	125.4	>	67.64
Magnesium (mg/g)	7.769	<	18.64
Iron (mg/g)	0.821	<	3.019
Total dissolved salts (TDS) (mg/g)	582.8	<	1766.7

## CONCLUSION

The chemical analysis variations and the differences in pathogenicity occurred. This behavior may be due to the development of “biotypes” within *R. solani*. The extremely

distorted segregation ratios for esterase enzyme system were clearly evident. Esterase linked phenotypes of *R. solani* isolates may be indicative for the genetic changes involved. It is worth to mention that use of nitrate fertilizers is not recommended for the soil cultivated by plants susceptible to *Rhizocotonia* infections, so the variant may become more virulent and more survival by increasing its melanin content.

## REFERENCES

- Burton, J.O. and J.P. Riley, 1954. In: Peach, K. and M.V. Tracey, (eds.), "Modern Methods of Plant Analysis", p. 370. Springer Verlag, Berlin.
- Cheng, W.T., J.C. Chen, W.T. Cheng and J.C. Chen, 2000. Effect of pH, temperature and salinity on immune parameters of fresh water prawn *Macrobrachium rosenbergii*. *Shell Fish Immunol.*, 10: 387–91
- Costa, J.M., V.A. Corbellini and M.L. Scroferneker, 2004. Study of different nitrogen sources on glucose uptake and production of melanin precursors and fungal mass of *Fonsecaea pedrosoi* cultured in tricyclazole. *Proc. Biochem.*, 39: 633–6
- Dahlquist, R.L. and J.W. Knoll, 1987. Inductively coupled plasma–atomic emission spectrometry, Analysis of biological materials and soils for trace and ultra–trace elements. *Appl. Spectrosc.*, 32: 1–30
- Deacon, J.W., 1997. *Modern Mycol.*, p. 303. Blackwell Science Ltd. USA.
- Dubois, M., K.A. Gillies, J.A. Hamilton, P.A. Rebers and F. Smith, 1956. Colorimetric method for determination of sugars and related substances. *Anal. Chem.*, 28: 250–6
- Eagen, R., A. Brisson and C. Breuil, 1997. The sap–staining fungus *Ophiostoma piceae* synthesizes different types of melanin in different growth media. *Canadian J. Microbiol.*, 43: 592–5
- Ellil, A.H.A.A., 1999. Oxidative stress in relation to lipid peroxidation, sclerotial development and melanin production in *Sclerotium rolfsii*. *J. Phytopathol.*, 147: 561–6
- Ellis, D.H. and D.A. Griffith, 1975. Melanin deposition in the hyphae of a species of *Phomopsis*. *Canadian J. Microbiol.*, 21: 422–52
- Fawcett, J.K. and J.E. Scott, 1960. A rapid and precise method for determination of urea. *J. Clin. Path.*, 13: 156–9
- George, E., C. Tamerler, A. Martinez, M.J. Martinez. and T. Keshavarz, 1999. Influence of growth medium composition on the lipolytic enzyme activity of *Ophiostoma piliferum* (CartapipTM). *J. Chem. Technol. Biotechnol.*, 74: 137–40
- Griffin, D.H., 1994. *Fungal Physiology*. John Wiley and Sons Inc. Publication. New York.
- Henson, J.M., 2001. *Biopolymers*. pp. 371–425. Lignin, humic substances and coal, Wiley–VCH, Weinheim, New York, Toronto
- HeungTae, K., Y.R. Chung, Y.K. Cho, H.T. Kim, Y.R. Chung and K.Y. Cho, 2001. Mycelial melanization of *Rhizocotonia solani* AG1 affecting pathogenicity in rice. *Plant Pathol. J.*, 17: 210–5
- HeungTae, K., J. Min, G.J. Choi *et al.*, 2002. Synthesis of a new 1, 2, 3–oxadiazole 5–one derivative, KC10017, and its controlling activity against rice blast disease caused by *Magnaporthe grisea*. *J. Pestic. Sci.*, 27: 229–34
- Jwa, N.S. and H.S. Chung, 1993. Complementation of melanin deficient mutant by protoplast fusion in *Pyricularia grisea*. Role of Fungi as frontiers of biosciences: *Proceed. of the Asian Mycological Symposium in Commemoration of the 20<sup>th</sup> Anniversary of Korean Society of Mycology*, pp. 160–72. October 1–4 1992, Seoul, Korea 1993
- Kademi, A., L. Fakhreddine, N.A. Abdkader and J.C. Baratti, 1999. Effect of culture conditions on growth and esterase production by the moderate thermophile *Bacillus circulans* MAS2.T. *Microbiol. Biotechnol.*, 23: 188–93
- Karla, Y.P. and D.G. Maynerd, 1991. Methods manual for forest soil and plant analysis. *Can. Serve Northwest Res.*, p. 319. Cent. Inf. Rep. Nor–X
- Kloepper, J.W., 1991. Development of *in vivo* assays for prescreening antagonists of *Rhizocotonia solani* on cotton. *Phytopathol.*, 81: 1006
- Ko, W. and F.K. Hara, 1971. A selective medium for the quantitative determination of *Rhizocotonia solani* in soil. *Phytopathol.*, 61: 707
- Kuc, J., H. Lyr, P.E. Russell, H.W. Dehne and H.D. Sisler, 1999. Specificity and lack of specificity as they relate to plant defence compounds and disease control. *Modern fungicides and antifungal compounds II*. 12<sup>th</sup> International Reinhardtsbrunn Symposium, Friedrichroda, Thuringia, Germany, pp. 31–7. 24<sup>th</sup>–29<sup>th</sup> May 1998–99
- Kuc, J., 1997. Molecular aspects of plant responses to pathogens. *Acta Physiol. Plant.*, 4: 551–9
- Kurahashi, Y., 2001. Melanin biosynthesis inhibitors (MBIs) for control of rice blast. *Pesticide Outlook.*, 12: 32–5
- Kusz, E., M. Wardas, M. Stec., W. Radwanska and A. Manowski, 2001. Estimation of antioxidative enzymes of the rats' liver remaining on diet enriched with nitrates (V) and vitamin E. *Poland J. Food Nutr. Sci.*, 10: 65–8
- Leroux, P., 2003. Fungicides resistance in plant pathogens: a phenomenon difficult to manage? *Phytoma*, 566: 36–40
- Linnemannstons, P., J. Schulte, P.M. Mar, R.H. Proctor, J. Avalos, B. Tudzynski and M. Prado, 2002. The polyketidesynthase gene pks4 from *Gibberella fujikuroi* encodes a key enzyme in the biosynthesis of the red pigment bikaverin. *Fung. Genet. Biol.*, 37: 143–8
- Lloyd, G.I., J.G. Anderson, J.E. Smith and E.O. Morris, 1972. Conidiation and esterase synthesis in *Aspergillus niger*. *Transact. British Mycol. Soc.*, 59: 63–70
- Mahapatra, N.P., S.P. Pati and R.C. Ray, 2001. Spoilage of sweet potato tuber in tropics. V. Java black rot by *Batrachydia theobromae* Pat: phenol accumulation in culture and in tubers during spoilage. *J. Mycopathol Res.*, 39: 21–7
- Mahto, B.N., R.N. Singh, C.P. Awasthi and A.B. Abidi, 1987. Sugars and phenolic compounds in rice leaves in relation to varietal resistance to bacterial blight pathogen. *Internat. Rice Res. Newslet.*, 12: 12–3
- Mayser, P., U. Schfer, H.J. Kramer, B. Irlinger and W. Steglich, 2002. Pityriacitrin–an ultraviolet–absorbing indole alkaloid from the yeast *Malassezia furfur*. *Arch. Dermatol. Res.*, 294: 131–4
- Mohammadi, M., M. Banihashmi, G.A. Hedjaroude and H. Rahimian, 2003. Genetic diversity among Iranian isolates of *Rhizocotonia solani* Kuhn anastomosis group 1 subgroups based on isozyme analysis and total soluble protein pattern. *J. Phytopathol.*, 151: 162
- Nicolaus, R.A., M. Piattelli and E. Fattorusso, 1964. The structure of natural black pigments. *Rend Acad. Sci. Fis. Mat. Naples Ser.*, 4: 83–97
- Pandy, B.P., 1982. *A Text Book of Plant Pathology. Pathogen and Plant Disease*. p. 452. Chand & Company Ltd., Ram Nagar, New Delhi
- Perrotey, S., B. Klein, L.G. Madulo, M. Gallego and B. Pesson, 2002. Variation of esterase electrophoresis associated with blood meal digestion in phlebotomine sand flies (Diptera: Psychodidae). *J. Med. Entomol.*, 39: 720–2
- Richard, N.S., 1993. *Plant Disease Control, Towards Environmentally Acceptable Methods*. p. 354. Chapman & Hall, London–Glasgow, New York, Tokyo, Melbourne and Madras
- Robert, K.A., 1989. Formation and structure of sclerotia and sclerotium–specific proteins in *Hygrospora aurantiaca*. *Mycologia*, 81: 905
- Scandalias, J.G., 1996. Genetic control of multiple molecular forms enzymes in plants. *Rev. Biochem. Genet.*, 3: 37–79
- Scott, S.W., 1981. Separation of *Sclerotina* isolates collected from three herbage legume hosts. *Trans. British Mycol. Soc.*, 76: 321–3
- Solomon, P.S., K. Tan, P. Sanchez, R.M. Cooper, R.P. Oliver and K.C. Tan, 2004. The disruption of a Galpha subunit sheds new light on the pathogenicity of *Stagonospora nodorum* on wheat. *Molec. Plant Microb. Interact.*, 17: 456–66
- Steiner, W.W.M. and D.J. Toslyn, 1979. Electrophoretic techniques for the genetic study of mosquitoes. *Mosq. New.*, 39: 35–54
- Tsuboi, R., H. Komatsuzak and H. Ogawa, 1996. Induction of extracellular esterase from *Candida albicans* and some of its properties. *Infect. Imm.*, 64: 2936–40
- Willets, H.J., 1978. Sclerotium formation. In: Smith, J.E. and D.R. Berry (eds.), *Filamentous Fungi III*, pp. 197–213. Edward Arnold, London.
- Williams, C.J., E.A. Shingara and J.B. Yavitt, 2000. Phenol oxidase activity in peatlands in New York State: response to summer drought and peat type. *Wetlands*, 20: 416–21

(Received 13 February 2005; Accepted 20 May 2005)