



### Full Length Article

## ***Rhizobium* and Phosphate Solubilizing Bacteria Improve the Yield and Phosphorus Uptake in Wheat (*Triticum aestivum*)**

AFTAB AFZAL AND ASGHARI BANO<sup>1</sup>

Department of Plant Sciences, Faculty of Biological Sciences, Quaid-i-Azam University Islamabad, Pakistan

<sup>1</sup>Corresponding author's e-mail: [asgharibano@yahoo.com](mailto:asgharibano@yahoo.com)

### ABSTRACT

*Rhizobium* and phosphorus (P) solubilizing bacteria are important to plant nutrition. These microbes also play a significant role as plant growth-promoting rhizobacteria (PGPR) in the biofertilization of crops. A study was conducted in order to investigate the effects of a rhizobial strain (Thal 8) and a P solubilizer strain (54RB) in single and dual combination with and without P<sub>2</sub>O<sub>5</sub> on wheat in a P deficient natural non-sterilized sandy loam soil. The results of this pot experiment revealed that single and dual inoculation with fertilizer (P<sub>2</sub>O<sub>5</sub>) significantly increased root and shoot weight, plant height, spike length, grain yield, seed P content, leaf protein and leaf sugar content of the test crop. It is concluded that single and dual inoculation along with P fertilizer is 30-40% better than only P fertilizer for improving grain yield of wheat and dual inoculation without fertilizer (P) improved grain yield up to 20% as compared to P application.

**Key Words:** Phosphate solubilizing bacteria; P uptake; *Rhizobium*; Wheat yield

### INTRODUCTION

Soil-plant-microbe interaction has got much importance in recent decades. Many types of microorganisms are known to inhabit soil, especially rhizosphere and play important role in plant growth and development. Phosphate solubilizing bacteria (PSB) are used as biofertilizer since 1950's (Kudashev, 1956; Krasilnikov, 1957). These microorganisms secrete different types of organic acids e.g., carboxylic acid (Deubel & Merbach, 2005) thus lowering the pH in the rhizosphere (He & Zhu, 1988) and consequently dissociate the bound forms of phosphate like Ca<sub>3</sub>(PO<sub>4</sub>)<sub>2</sub> in calcareous soils.

Use of these microorganisms as environment friendly biofertilizer helps to reduce the much expensive phosphatic fertilizers. Phosphorus biofertilizers could help increase the availability of accumulated phosphate (by solubilization), efficiency of biological nitrogen fixation and increase the availability of Fe, Zn etc., through production of plant growth promoting substances (Kucey *et al.*, 1989). Trials with PSB indicated yield increases in rice (Tiware *et al.*, 1989), maize (Pal, 1999) and other cereals (Afzal *et al.*, 2005; Ozturk *et al.*, 2003).

*Rhizobium* previously well known as a symbiotic N fixer is reported as asymbiotic (associative & endophytic) microorganisms in recent years (Biswas *et al.*, 2000a, b). The most inoculation studies have focused on free living diazotrophs, although a few reports indicate *rhizobia* can act as plant growth promoting rhizobacteria (PGPR) (Hoflich *et al.*, 1995; Noel *et al.*, 1996; Yanni *et al.*, 1997). The PGPR influence the crop growth and development by releasing plant growth regulators (Glick & Bashan, 1997; Volpin & Philips, 1998) and improving morphological characteristics

of inoculated roots (Biswas, 1998), which favored nutrient uptake (Okon & Kapulnik, 1986). The growth promoting effects of rhizobacteria may include phytohormone production (Chabot *et al.*, 1996), N<sub>2</sub> fixation (Urquiaga *et al.*, 1992) and more efficient use of nutrients (Chabot *et al.*, 1996). Yanni *et al.* (1997) and Biswas (1998) reported increased N uptake in rice plants inoculated with rhizobia.

Combined inoculation with N<sub>2</sub> fixing and phosphate solubilizing bacteria was more effective than single microorganism for providing a more balanced nutrition for plants (Belimov *et al.*, 1995). Dual inoculation increased yields in sorghum (Algawadi & Gaur, 1992), barley (Belimov *et al.*, 1995), black gram (Tanwar *et al.*, 2002), soybean (Abdalla & Omar, 2001) and wheat (Galal, 2003).

Reports regarding co-inoculation of *Rhizobium* and P solubilizing bacteria on wheat are rare, especially in Pakistan very little work has been done on these lines. The objective of present investigation was to explore the effect of single and dual inoculations with N<sub>2</sub>-fixing and P-solubilizing bacterial species on wheat yield and P uptake and to explore the hidden potential of *Rhizobium* as a PGPR for this important non-legume.

### MATERIALS AND METHODS

A pot experiment was carried out in experimental area of Quaid-i-Azam University Islamabad, Pakistan, to investigate the effects of single and dual inoculations with N-fixing bacteria (*Rhizobium leguminismarum* Thal-8/SK8) and phosphorus solubilizing bacteria (*Pseudomonas* sp. strain 54RB) on yield and yield components of wheat (*Triticum aestivum*) variety Margalla 99. The bacterial strains, Thal-8 originally isolated from Thal area and 54RB isolated from rice, were obtained from Soil Biology and

Biochemistry Lab. National Agricultural Research Centre (NARC) Islamabad, Pakistan. Wheat seeds were obtained from Institute of Field Crops (wheat section), NARC Islamabad. Clay pots (10 kg soil capacity) were used for trial. Sandy loam soil having organic matter 0.65%, pH 7.9, P content 8 ppm and NO<sub>3</sub>-N 10 ppm collected from farmer field was used. Urea and single super phosphate (SSP) were applied as source of N and P respectively.

Un-sterilized soil was air dried and mixed with sand in 1:1 ratio and filled in pots. Composite soil samples were collected and analyzed (analysis given above). These pots were placed under natural conditions. Urea was mixed in upper soil (0-10 cm) @ 1.8 g pot<sup>-1</sup> before sowing. Similarly SSP was applied as treatment in respective pots @ 2 g per pot. Eight seeds were sown in each pot. With average minimum temperature of 9°C and average maximum temperature of 25°C, a total rainfall of 430 mm, relative humidity of 80% and day length ranging from 10-13 h, plant growth in the region is restricted to the period between November and May. The experimental design for the study was completely randomized block design with six replicates. Treatments were: T0 control (without inoculation & with N) T1 (P<sub>2</sub>O<sub>5</sub>), T2 (*Rhizobium* @10 mL broth culture per plant), T3 (PSB @10 mL broth culture per plant), T4 (PSB+*Rhizobium*), T5 (P<sub>2</sub>O<sub>5</sub>+*Rhizobium*), T6 (P<sub>2</sub>O<sub>5</sub>+PSB) and T7 (P<sub>2</sub>O<sub>5</sub>+PSB+*Rhizobium*).

Inoculum for *Rhizobium* was prepared in yeast mannitol (YM) broth (Somasegaran & Hoben, 1985) by inoculating autoclaved YM with *Rhizobia* (1 mL) and placed in shaker at 28°C. Inoculum for PSB was prepared in Pikovskaya medium by the same procedure. Inoculation @10 mL per seedling was done at log phase i.e., about 10<sup>8</sup> cells mL<sup>-1</sup> at the base of plant. Five plants per pot were kept after thinning and grown until maturity to get final yield.

During the growing period, leaf samples were collected (at 108 days after sowing) for sugar and protein analysis before heading. The analyses were done according to Johnson *et al.* (1966) and Lowry *et al.* (1951) respectively. At maturity, data regarding plants height, spike length, root weight, shoots weight and grain yield were recorded using standard procedures. Soil samples from each treatment were collected and analyzed for phosphorus content according to Soltanpur and Workman (1979). Seed P content was measured according to Cottenie (1980). In the present study, P solubilizing ability of 54RB and Thal-8 was demonstrated based on the qualitative methods (Metha & Nautiyal, 2001) under the laboratory condition. Indole acetic acid (IAA) and gibberellic acid (GA) production for both the strains was also determined using HPLC (Table I). Phosphate solubilization was determined using following formula:

$$SI = \frac{\text{colony diameter} + \text{halozone diameter}}{\text{Colony diameter}}$$

**Statistical analysis.** The data obtained in the study were subjected to analysis of variance using MSTATC, computer

software (Bricker, 1991) and means were compared by Duncan's Multiple Range Test (Duncan, 1955). Correlations between growth parameter (s) were also calculated (Bricker, 1991).

## RESULTS AND DISCUSSION

In this study generally all the parameters were significantly affected by inoculation treatments over control (Table II). Root weight and seed P content (%), however, were non-significantly affected by dual inoculation without fertilizer P (Table II). The highest shoot weight was obtained in pots, which received *Rhizobium* inoculation along with fertilizer that was statistically similar to pots, which received PSB with fertilizer and dual inoculation (*Rhizobium* + PSB) with fertilizer (Table II). Single inoculation of *Rhizobium* and PSB behaved similar to that of control for root and shoot weight (Table II). Dual inoculation with fertilizer yielded maximum root weight followed by single inoculation of *Rhizobium* with P and single inoculation of PSB along with P. Control and other treatments had similar root weight (Table II). Increased root, shoot weight with dual inoculation have been reported by Höflich *et al.* (1994) in barley, Yanni *et al.* (1997) in rice, Chabot *et al.* (1993) in maize. Our results are also in line with similar previous findings. We also found a highly positive correlation ( $r = 0.79$ ) between root and shoot weight (Table III). Better root development may be due to synergistic relationship of the inoculated bacteria for improving root length and weight by producing growth regulators (Okon, 1985) like IAA and GA (Table I) that along with P fertilizer favored better root development, which enhanced water and nutrient uptake and ultimately helped better shoot development. Single inoculation of either PSB or *Rhizobium* showed no statistical difference over control (Höflich, 2000). However, dual inoculation was the most effective in increasing root/shoot weight. According to Bashan (1998) soil microbial cultures with statistically similar or different functions might express beneficial actions in a soil or rhizosphere.

All inoculations with fertilizer P and dual inoculation without P had significantly greater plant height than other treatments. Single inoculation of *Rhizobium*/PSB, without fertilizer P did not increase plant height over control. Spike length was increased by fertilizer P, dual inoculation and combined application of inoculants + P. Fertilizer P alone and in combinations with inoculations and dual inoculation all significantly increased the grain yield over control as well as single inoculations. These results confirmed the previous findings (Khalid *et al.*, 1997; Biswas *et al.*, 2000a, b; Hilali *et al.*, 2000, 2001), who reported increased plant height and spikelets per spike of various crop plants by microbial inoculation. Plant height and spike length were significantly correlated ( $r = 0.80$  &  $r = 0.94$ ) with shoot weight and with each other ( $r = 0.80$ ) (Table III).

Highest grain yield (3.19 g pot<sup>-1</sup>) was obtained with PSB+P<sub>2</sub>O<sub>5</sub> but it was similar to P<sub>2</sub>O<sub>5</sub> and combination of

**Table I. Some important biochemical features of microbial strains tested during this investigation. Bacterial cultures were grown in respective media incubated for one week. Production of IAA and GA was determined in cell free medium**

Bacterial strains	IAA production $\mu\text{g/ml}$	GA production $\mu\text{g/ml}$	P solubilization index	Gram reaction
<i>Rhizobium leguminosarum</i> (Thal 8)	65.90	833.7	2.5	Negative
<i>Pseudomonas</i> sp. (54RB)	8.034	1766	4.1	Negative

**Table II. Effect of *Rhizobium* and phosphate solubilizing bacteria (PSB) on agronomic traits, yield components, seed and soil P content, leaf sugar and protein content in wheat**

Treatments	Shoot wt. (g pot <sup>-1</sup> )	Root wt. (g pot <sup>-1</sup> )	Plant height (cm)	Spike length (cm)	Grain yield (g pot <sup>-1</sup> )	Seed content (%)	P Soil (ppm)	P content	Leaf sugar ( $\mu\text{g g}^{-1}$ )	Leaf protein ( $\mu\text{g g}^{-1}$ )
Control (T0)	3.10 c	0.10 c	55.30 b	7.30 c	1.22 b	0.30d	4.10		21.40 c	3767 c
P <sub>2</sub> O <sub>5</sub> (T1)	6.22 b	0.30 c	64.00 ab	8.80 bc	2.28 a	0.36c	4.33		70.20 b	3930 bc
<i>Rhizobium</i> (T2)	4.10 c	0.20 c	56.70 b	7.50 c	1.15 b	0.31d	4.67		65.30 b	4474 a
PSB (T3)	4.05 c	0.12 c	55.50 b	7.20 c	1.25 b	0.38c	4.57		25.80 c	3966 bc
<i>Rhizobium</i> + PSB (T4)	6.41 b	0.33 c	68.30 a	9.70 ab	2.85 a	0.29d	4.53		64.50 b	4419 a
<i>Rhizobium</i> + P <sub>2</sub> O <sub>5</sub> (T5)	10.3 a	0.68 b	69.30 a	11.00 a	2.31 a	0.45b	4.10		73.70 b	3826 bc
PSB + P <sub>2</sub> O <sub>5</sub> (T6)	8.93 a	0.65 b	71.70 a	10.50 ab	3.19 a	0.45b	4.87		66.70 b	4188 ab
<i>Rhizobium</i> + PSB + P <sub>2</sub> O <sub>5</sub> (T7)	9.20 a	0.93 a	67.30 a	10.00 ab	2.93 a	0.49a	5.00		94.50 a	4448 a
LSD values	2.075	0.235	9.044	1.551	0.881	0.25	NS		19.31	376.4

Values followed by different letters in a column were significantly different ( $P < 0.05$ ), using Duncan's multiple range test. NS, not significant. PSB, Phosphate Solubilizing Bacteria (*Pseudomonas* sp.) rhizobium (*Rhizobium leguminosarum*), P<sub>2</sub>O<sub>5</sub> @ 0.38 g pot<sup>-1</sup>

**Table III. Correlation effects of *Rhizobium* and phosphate solubilizing bacteria on wheat yield and yield traits (per plant basis)**

Variable	Shoot wt	Root wt	Plant height	Spike length	Grain yield	Flag leaf Sugar	Soil P
Root wt.	0.796243**						
Plant height	0.803548**	0.65216**					
Spike length	0.944879**	0.707167**	0.815159**				
Grain yield	0.786934**	0.655576**	0.768131**	0.78742**			
Flag leaf Sugar	0.671061**	0.608289**	0.627023**	0.623612**	0.569902**		
Soil P	0.718864**	0.690239**	0.614421**	0.620678**	0.630866**	0.625122**	
Seed P	0.619352**	0.741055**	0.398551 <sup>NS</sup>	0.471929*	0.34184 <sup>NS</sup>	0.521501**	0.692944**

\*\* = Highly significant at 1% level of probability, \* = Significant at 5% level of probability, NS = Non-significant

P<sub>2</sub>O<sub>5</sub> and inoculations. *Rhizobium* and PSB alone did not increase the grain yield. Dual inoculation with P<sub>2</sub>O<sub>5</sub> had highest seed P content of 0.49%. The *Rhizobium* alone did not increase seed P content. However, PSB alone increase P content as compared to control. Many researchers reported increased seed P content by phosphate solubilizing microorganisms (Kucey, 1987; Mehana & Wahid, 2002; Zaidi *et al.*, 2004), our study revealed the same effect. But dual inoculation without fertilizer could not improve seed P concentration. This may be due to competition of the microbes for P (Azcon-Aguillar *et al.*, 1986). Soil P were not affected by any of the treatment significantly.

Leaf sugar content was significantly increased by *Rhizobium* inoculation and fertilizer treatments. Highest value of leaf sugar content (49.5  $\mu\text{g g}^{-1}$ ) was recorded in dual inoculation with fertilizer treated plants. Leaf protein was significantly affected by rhizobial treatments and best treatment in this case was *Rhizobium* along with P<sub>2</sub>O<sub>5</sub>. This may be due to increased N uptake by a larger root surface area associated with additional root hairs and lateral root development and/or to BNF, either directly by the inoculants strains or indirectly by stimulating BNF activity of the associated rhizosphere community (Ladha *et al.*, 1998). Maximum sugar was accumulated in dual inoculated and fertilized plants. Sahin *et al.* (2004) documented increased sugar content in sugar beet by co-inoculation of N-fixing and P solubilizing bacteria.

The present study indicated that single inoculations with *Rhizobium* could not increase grain yield as compared to control. Inoculation with PSB alone increased yield by 2.5% over control (Table II). Dual inoculation of *Rhizobium* and PSB gave yield increases of 29% and 25% with and without fertilizer respectively. Fertilizer P application, however gave yield increases up to 80%. It is concluded that mixed culturing is better than single inoculation. Reduced fertilization (up to 50%) is required with inoculations. Rhizobial and/or PSB inoculation without fertilization is not effective. *Rhizobium* with non-legumes could act as phosphate solubilizer, hormone producer and to some extent as N-fixer. Addition of *Azospirillum* or other diazotrophs (for N fixation) with present combination would give promising results, which will reduce the N fertilizer use for the profit of farmer and prove environment-friendly.

## REFERENCES

- Abdalla, M.H. and S.A. Omer, 2001. Survival of Rhizobia/Bradirrhizobia and a rock phosphate solubilizing fungus *Aspergillus niger* on various carriers from some agroindustrial wastes and their effect on nodulation and growth of fababean and soybean. *J. Plant Nut.*, 24: 261–72
- Afzal, A., M. Ashraf, S.A. Asad and M. Farooq, 2005. Effect of phosphate solubilizing microorganisms on phosphorus uptake, yield and yield traits of wheat (*Triticum aestivum* L.) in rainfed area. *Int. J. Agric. Biol.*, 7: 207–9
- Alagawadi, A.R. and A.C. Gaur, 1992. Inoculation of *Azospirillum brasilense* and phosphate-solubilizing bacteria on yield of sorghum (*Sorghum bicolor* L. Moench) in dry land. *Trop. Agric.*, 69: 347–50

- Azcon-Aguillar, C., V. Gianinazzi-Pearson and J.C. Fardeau, 1986. Effect of vesicular-arbuscular mycorrhizal fungi and phosphate-solubilizing bacteria on growth and nutrition of soybean in a neutral calcareous soil amended with 32P-45Ca tricalcium phosphate. *Plant Soil*, 96: 3–15
- Bashan, Y., 1998. Inoculations of plant growth-promoting bacteria for use in agriculture. *Biotechnol. Adv.*, 16: 729–70
- Belimov, A.A., P.A. Kojemiakov and C.V. Chubarlyyeva, 1995. Interaction between barley and mixed cultures of nitrogen fixing and phosphate-solubilizing bacteria. *Plant Soil*, 17: 29–37
- Biswas, J.C., J.K. Ladha and F.B. Dazzo, 2000a. *Rhizobia* inoculation improves nutrient uptake and growth of lowland rice. *Soil Sci. Soc. America J.*, 164: 1644–50
- Biswas, J.C., J.K. Ladha, F.B. Dazzo, Y.G. Yanni and B.G. Rolfe, 2000b. Rhizobial inoculation influences seedling vigor and yield of rice. *Agron. J.*, 92: 880–6
- Biswas, J.C., 1998. Effect of Nitrogen Fixing Bacteria on Growth Promotion of Lowland Rice (*Oryza sativa* L.). *Ph. D Thesis*, Department of Soil Science University of Philippines, Los Banos
- Bricker, B., 1991. *A Micro Computer Programme for the Design, Manage and Analysis of Agronomic Research Expt.* Crop and Science Department, AUS, Lansin USA
- Çakmakçi, R., F. Kantar and F. Sahin, 2001. Effect of N<sub>2</sub>-fixing bacterial inoculations on yield of sugar beet and barley. *J. Plant Nutr. Soil Sci.*, 164: 527–31
- Chabot, R., H. Antoun and M.P. Cescas, 1993. Stimulation de la croissance du maïs et de la laitue romaine par des microorganismes dissolvant le phosphore inorganique. *Canadian J. Microbiol.*, 39: 941–7
- Chabot, R., H. Antoun and M.P. Cescas, 1996. Growth promotion of maize and lettuce by phosphate-solubilizing *Rhizobium leguminosarum* biovar phaseoli. *Plant Soil*, 184: 311–21
- Cottenie, A., 1980. Soil and Plant testing as a basis of fertilizer recommendations. *FAO. Soil's Bulletin*, 38: 64–5
- Deubel, A. and W. Merbach, 2005. Influence of microorganisms on phosphorus bioavailability in soils. In: Buscot, F. and A. Varma (eds.), *Microorganisms in Soils: Roles in Genesis and Functions*, pp: 177–91. Springer-Verlag, Berlin Heidelberg, Germany
- Duncan, D.B., 1955. Multiple range and multiple F-test. *Biometrics*, 11: 1–42
- Galal, Y.G.M., 2003. Assessment of nitrogen availability to wheat (*Triticum aestivum* L.) from inorganic and organic N sources as affected by *Azospirillum brasilense* and *Rhizobium leguminosarum* inoculation. *Egyptian J. Microbiol.*, 38: 57–73
- Glick, B.R. and Y. Bashan, 1997. Genetic manipulation of plant growth promoting bacteria to enhance biocontrol of phytopathogens. *Biotechnol. Adv.*, 15: 353–78
- He, Z. and J. Zhu, 1988. Microbial utilization and transformation of phosphate adsorbed by variable charged minerals. *Soil Biol. Biochem.*, 30: 917–23
- Hilali, A., D. Przvost, W.J. Broughton and H. Antoun, 2000. Potential use of *Rhizobium leguminosarum* bv. trifolii as plant growth promoting rhizobacteria with wheat Abstract. *17<sup>th</sup> North American Conference on Symbiotic Nitrogen Fixation*, Laval University, Quebec, Canada 23-28 July 2000
- Hilali, A., D. Przvost, W.J. Broughton and H. Antoun, 2001. Effets de l'inoculation avec des souches de *Rhizobium leguminosarum* bv. trifolii sur la croissance du blé dans deux sols du Maroc. *Canadian J. Microbiol.*, 47: 590–3
- Höflich, G., 2000. Colonization and growth promotion of non-legumes by *Rhizobium* bacteria. Microbial Biosystems: New Frontiers. In: Bell, C.R., M. Brylinsky and P. Johnson-Green (eds.), *Proceedings of the 8<sup>th</sup> International Symposium on Microbial Ecology*, pp: 827–30. Atlantic Canada Society for Microbial Ecology, Halifax, Canada
- Höflich, G., W. Wiehe and G. Köhn, 1994. Plant growth stimulation by inoculation with Symbiotic and associative rhizosphere microorganisms. *Experientia*, 50: 897–905
- Johnson, R.R., T.L. Balwani, L.J. Johnson, K.E. Meclure and B.A. Denority, 1966. Corn plant maturity II. Effect on *in-vitro* cellular digestibility and soluble carbohydrate content. *J. Anim. Sci.*, 25: 617
- Khalid, A., M. Arshad, Z.A. Zahir and A. Khaliq, 1997. Potential of plant growth promoting rhizobacteria for enhancing wheat yield. *J. Anim. Plant Sci.*, 7: 53–6
- Krasilnikov, N.A., 1957. On the role of soil micro-organism in plant nutrition. *Microbiologiya*, 26: 659–72
- Kudashev, I.S., 1956. The effect of phosphobacterin on the yield and protein content in grains of Autumn wheat, maize and soybean. *Dokl. Akad. Skh. Nauk*, 8: 20–3
- Kucey, R.M.N., H.H. Janzen and M.E. Leggett, 1989. Microbially mediated increases in plant-available phosphorus. *Ad. Agron.*, 42: 199–228
- Kucey, R.M.N., 1987. Increased phosphorus uptake by wheat and field beans inoculated with a phosphorus-solubilizing *Penicillium bilaji* strain and with vesicular-arbuscular mycorrhizal fungi. *Appl. Environ. Microbiol.*, 53: 2699–703
- Ladha, J.K., G.J.D. Kirk, J. Bennett, C.K. Reddy, P.M. Reddy and U. Singh, 1998. Opportunities for increased nitrogen use efficiency from improved lowland rice germplasm. *Field Crops Res.*, 56: 36–69
- Lowry, O.H., N.J. Rosbrough, A.L. Farr and R.J. Randall, 1951. *J. Biol. Chem.*, 193: 265
- Mehana, T.A. and O.A. Wahid, 2002. Associative effect of phosphate dissolving fungi, *Rhizobium* and phosphate fertilizer on some soil properties, yield components and the phosphorus and nitrogen concentration and uptake by *Vicia faba* L. under field conditions. *Pakistan J. Biol. Sci.*, 5: 1226–31
- Methaa, S. and C.S. Nautiyal, 2001. An efficient method for qualitative screening of phosphate-solubilizing bacteria. *Curr. Microbiol.*, 43: 51–6
- Noel, T.C., C. Sheng, C.K. Yost, R.P. Pharis and M.F. Hynes, 1996. *Rhizobium leguminosarum* as a plant growth-promoting rhizobacterium: direct growth promotion of canola and lettuce. *Canadian J. Microbiol.*, 42: 279–83
- Okon, Y., 1985. *Azospirillum* as a potential inoculant for agriculture. *Trends Biotechnol.*, 3: 223–8
- Okon, Y. and Y. Kapulnik, 1986. Development and function of *Azospirillum* inoculated roots. *Plant Soil*, 90: 3–16
- Öztürk, A., O. Cağlar and F. Sahin, 2003. Yield response of wheat and barley to inoculation of plant growth promoting rhizobacteria at various levels of nitrogen fertilization. *J. Plant Nutr. Soil Sci.*, 166: 1–5
- Pal, S.S., 1999. Interaction of an acid tolerant strain of phosphate solubilizing bacteria with a few acid tolerant crops. *Plant Soil*, 213: 221–30
- Sahin, F., R. Cakmakci and F. Kantar, 2004. Sugar beet and barley yields in relation to inoculation with N<sub>2</sub>-fixing and phosphate solubilizing bacteria. *Plant Soil*, 265: 123–9
- Soltanpur, P.N. and S. Workman, 1979. Modification of the NH<sub>4</sub>CO<sub>3</sub>-DTPA soil test to omit carbon black. *Commun. Soil Sci. Plant Annl.*, 10: 1411–20
- Somasegaran, P. and H.J. Hoben, 1985. *Methods in Legume-rhizobium Technology*. NifTAL project and MIRCEN. Department of Agronomy, 2<sup>nd</sup> Soil Science Hawaii Institute Tropical Agriculture Human research, Univ Hawaii Manoa
- Tanwar, S.P.S., G.L. Sharma and M.S. Chahar, 2002. Effect of phosphorus and biofertilizers on the growth and productivity of black gram. *Annl. Agric. Res.*, 23: 491–3
- Tiwari, V.N., L.K. Lehri and A.N. Pathak, 1989. Effect of inoculating crops with phospho-microbes. *Exp. Agric.*, 25: 47–50
- Urquiaga, S., K.H.S. Cruz and R.M. Boddey, 1992. Contribution of nitrogen fixation to sugar cane: nitrogen-15 and nitrogen-balance estimates. *Soil Sci. Soc. America Proc.*, 56: 105–14
- Volpin, H. and D.A. Philips, 1998. Respiratory elicitors from *Rhizobium meliloti* effect intact alfalfa roots. *Plant Physiol.*, 116: 777–83
- Yanni, Y.G., R.Y. Rizk, V. Corich, A. Squartini, K. Ninke, S. Philip-Hollingsworth, G. Orgambide, F. De Bruijn, J. Stoltzfus, D. Buckley, T.M. Schmidt, P.F. Mateos, J.K. Ladha and F.B. Dazzo, 1997. Natural endophytic association between *Rhizobium leguminosarum* bv. trifolii and rice roots and assessment of its potential to promote rice growth. *Plant Soil*, 194: 99–114
- Zaidi, A., M.S. Khan and M. Aamil, 2004. Bioassociative effect of rhizospheric microorganisms on growth, yield, and nutrient uptake of greengram. *J. Plant Nut.*, 27: 601–12

(Received 29 May 2007; Accepted 05 October 2007)