**Isolation and Optimization Growth Parameters for Enhanced bioactivity profiles of Streptomyces strain isolated from Saudi Arabia**

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**Abstract**

The genus Streptomyces is a unique subgroup of actinomycetes bacteria that well-known as prolific products of many bioactive secondary metabolites as well as antibiotics. *Streptomyces globosus* (*S.* *globosus*) is a bacterium species from the genus of [*Streptomyces*](https://en.wikipedia.org/wiki/Streptomyces)*.* Objective: This study aimed to optimize the growth conditions required for the maximum antibacterial activity of *S.* *globosus* isolated from soil samples in Saudi Arabia. The effect of dark/light, temperature and pH on the growth and antibacterial effect against *E. coli ATCC35218, E. coli ATCC25922, Salmonella sp., P. aeruginosa ATCC27853, S. aureus ATCCBAA977,* and *St. pneumonia ATCC49619* were optimized. The results of this study showed that*.* the highest antibacterial activity was at 30 ° C after 7 days of incubation. Furthermore, the best to produce antibacterial activity was under alkaline conditions (pH 8.5). Besides, the isolate didn't show antibacterial activity at the shaking incubator. This study showed that all organic solvents (ethyl-alcohol, methanol, chloroform, ethyl-acetate, and acetone) succeeded in extracting the antibacterial compound. Still, the compounds extracted by chloroform had the highest antibacterial activity. The MIC of the chloroform extract was against *S. aureus ATCCBAA977.* The study described the optimized condition for best *S. globosus* activity. **Conclusions:** The *S. globosus* showed a broad spectrum of antimicrobial activities against the test organisms and this opened further research investigations on purification and structural characterization of the active compounds from the crude extract.

**Keyword:** *Streptomyces globosus*, condition, dark/light, temperature Antibacterial activity, and pH

**Introduction**

Antimicrobial is used to spare people from contamination. It is a substance or a synthetic item with an inhibitory or destructive impact on the microorganisms ( Piddock, 2012). The advancement of safe strains is a characteristic wonder that happens through choice weight on the antimicrobial microorganism populace ( Chellat et al., 2016 2). Nonetheless, some of these antimicrobials have been rendered dormant by resistant systems (Nikolaidis et al., 2014). Several of the natural antimicrobials were extracted from soil microbes (Jain and Pundir, 2011 ).

Actinomycetes are a group of prokaryotic microorganisms which are gram-positive bacteria with high guanine+cytosine in their DNA (Adegboye and Babalola, 2012 ). They are considered as biotechnologically important organisms since they are responsible for producing about half of the bioactive secondary metabolites including antibiotics. They are filamentous bacteria which produce two types of branching mycelium, namely aerial and substrate mycelium [Yuan et al., 2010 6].

Streptomyces (family Streptomycetaceae) are the most significant Actinobacteria (Rong et al., 2009 ). It is disseminated in soil, water, and other indigenous habitats (Singh et al., 2006). In all years Streptomyces strains as wellspring of various bioactive mixes, for example, anthelminthic chemicals, herbicides, hostile to malignancy drugs, development components and insusceptible modulators (Kumar et al., 2020).

Factors influencing the number and types of actinomycetes present in a particular soil are a geographical location, such as soil type, temperature, organic matter content, moisture content, cultivation and aeration. Actinomycetes act as a major component of the microbial population in most of the soil. About 90% of the total actinomycetes population consists of *Streptomyces* species (Harir et al., 2018). Almost 80% of the world’s antibiotics are known to come from actinomycetes, mostly from the genera *Streptomyces* and *Micromonospora* (Pandey et al., 2004).

A major global healthcare problem has occurred in the 21st century because of increasing resistance of microorganisms against commonly used antibiotics. A renewed search for new antimicrobial agents from *Streptomyces* has occurred because of this rapid emergence of antimicrobial resistance among pathogenic microorganisms (Alanis, 2005).

Microscopic organisms are profoundly versatile pathogens that create protection from antimicrobials through a few mechanisms. The creation of β-lactamases that hydrolyze the β-lactam ring is the most widely recognized resistant system for these microscopic organisms against b-lactam antimicrobials (Pfeifer et al., 2010). The creation of board range β-lactamases (ESBLs) is a noteworthy opposition instrument that obstructs the antimicrobial treatment of contaminations brought about by Enterobacteriaceae and is a genuine risk to the presently accessible antimicrobial arsenal (Shaikh et al., 2015). Bacterial efflux pumps effectively transport numerous antimicrobials out of the cell. They are significant contributors to the characteristic resistant of gram-negative microscopic organisms to many medications that can be utilized to treat gram-positive bacterial contaminations (Blair et al., 2015).

This study aimed to optimize the growth conditions required for the maximum antibacterial efficacy of Streptomyces globosus (S. globosus) separated from soil samples in Saudi Arabia.

**Materials and Methods**

**Effect of different pH values**

Calibration of 100 ml of sterile starch-nitrate broth medium for different pH levels; 6, 6.5, 7, 7.5, 8, 8.5, 9, and 9.5 using Na OH for alkaline and HCl for acidic before sterilizing medium was performed. The cultures were incubated at 30Co. The culture filtrates were centrifuged (4000 rpm in 30 m). The antibacterial efficacy of CFF was investigated using the agar well diffusion technique. Twenty-four hours post incubation (35 ± 2°C), the antibacterial efficacy was calculated by observing the zone of inhibition radius in millimeters.

**Effects of the light factor**

Nine Petri plates by the selected *streptomyces* isolate were incubated (three in light, dark, or 12 h light/ 12 h dark) at 28°C for 7 days. Two agar discs (6 diameters) of *Streptomyces* isolate were placed over the surface of the tested bacteria's cultures with equal distance and incubated at 35 ± 2 °Cfor one day. After incubation, the antibacterial activity was estimated by measuring the inhibition zones' diameter in (mm).

**Effect of different incubation periods at different incubation temperatures**

The chosen *Streptomyces sp.* was cultured on sterile starch-nitrate broth (100 ml) medium at different temperatures (25°, 30°, 35° and 40° C) for various periods (1, 2, 3, 4, 5, 6,7, 8 and 9). At each incubation period and different temperatures, the culture filtrates were centrifuged (4000 rpm in 30 m). The antibacterial efficacy of cell-free filtrate (CFF) was investigated using the agar well diffusion technique. Twenty-four hours post incubation (35 ± 2 ° C), the antibacterial efficacy was calculated by observing the zone of inhibition radius in millimeters.

**Effect of shaking incubator**

The chosen *Streptomyces sp.* was cultured on sterile starch-nitrate broth (100 ml) medium in optimum conditions. Half of the cultures were incubated on a shaking incubator, and others were incubated in the constant incubator. The culture filtrates were centrifuged (4000 rpm in 30 m). The antibacterial efficacy of CFF was investigated using the agar well diffusion technique. Twenty-four hours post incubation (35 ± 2 ° C), the antibacterial efficacy was calculated by observing the zone of inhibition radius in millimeters.

**Extraction of an antibacterial compound with organic solvents**

Four different solvents (ethyl-alcohol, methanol, chloroform, ethyl-acetate, and acetone) were used to extract the antibacterial activity agent that was produced by the selected *Streptomyces* sp. The chosen *Streptomyces sp.* was cultured on sterile starch-nitrate broth (100 ml) medium in optimum conditions. The culture filtrates were centrifuged (4000 rpm in 30 m). The CFF was transferred aseptically into the conical flasks, and an equal volume of solvents was separately added to the CFF and shaken for 20 min. After gathering the organic phase, the solvent was evaporated under vacuum. 50 µl of the extract was used to investigate antibacterial efficacy using the agar well diffusion technique. Twenty-four hours post incubation (37 ° C), the antibacterial efficacy was calculated by observing the zone of inhibition radius in millimeters.

**MIC values against nosocomial infection bacteria**

The process includes the preparation of a double dilution of the antimicrobial agents in a liquid medium. Upon dilution of the standardized bacterial culture, each tube with a bacterial suspension was packed in the same medium. The inoculated tubes were incubated (mostly without agitation) under suitable conditions depending upon the test microorganism.

**Statistics**

The data presented as mean of 3 replicates ± SD. The difference between the means was statistically compared using Microsoft Excel software.

**Results**

**Effect of the light factor on antibacterial activity of *S. globosus***

Results in table (1) revealed no significant difference in the antibacterial activity when *S. globosus* was grown in different lighting. It had antibacterial activity against all the test bacteria except *P.earuginosa* ATCC27853, but no differences in the antibacterial activity against the other test bacterial isolates when *S. globosus* was grown in light, dark or 12h light / 12h dark, it showed antibacterial activity against *E.coli* ATCC35218 only when it was grown in dark

**Table 1.** Effect of the light factor on antibacterial activity of *S. globosus*

|  |  |
| --- | --- |
| Light factors | Inhibition zone diameter (mm) |
| *E. coli ATCC35218* | *E. coli ATCC25922* | *Salmonella sp.* | *P. earuginosa ATCC27853* | *S. aureus ATCCBAA977* | *St. pneumonia ATCC49619* |
| Dark | 11.3 ± 0.47 | 15.6 ± 0.47 | 18.3 ± 0.47 | - | 24.3 ± 0.47 | 14.6 ± 0.47 |
| Light | - | 15.3 ± 0.47 | 17.16 ± 0.23 | - | 23.16 ± 0.23 | 13.3 ± 0.23 |
| 12h light / 12h dark | - | 15.3 ± 0.47 | 17.83 ± 0.23 | - | 24 ± 0.00 | 14.3 ± 0.47 |

**Effect of different incubation periods and different incubation temperature on antibacterial activity of *S. globosus***

Incubation of *S. globosus* at 25oC showed antibacterial activity against *S. aureus* ATCCBAA977 only, after five days of incubation and increased by increasing the incubation period**.** The most excellent antibacterial activity was after seven days of incubation, and the inhibition zone was 14.3 mm. Still, there was no antibacterial activity against other Gram-positive and Gram-negative bacteria (Figure 1).

Results in Figures 1 revealed that antibacterial activity against most of the test bacteria was noticed and increased with increasing the incubation period at 30oC. The optimum antibacterial activity was after seven days of incubation. The diameters of inhibition zones were (13.3 mm, 18.0 mm, 12.0 mm, 19.3 mm, and 16.3 mm) against *E. coli* ATCC35218, *E. coli* ATCC25922, *Salmonella* sp., *S.**aureus* ATCCBAA977 and *St. pneumonia* ATCC49619, respectively.

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**Figure 1.** Effect of different incubation periods and different incubation temperature on antibacterial activity of *S. globosus*

From the results in Figure 1 when *S. globosus* was incubated at 35oC, the antibacterial efficacy was versus *E. coli* ATCC25922, *Salmonella* sp., and *S. aureus* ATCCBAA977. The antibacterial activity against *Salmonella* sp. and *S. aureus* ATCCBAA977 began from the third day but after five days produced antibacterial activity against *E. coli* ATCC25922. The optimum antibacterial activity was at the fifth day, and the diameters of inhibition zones were (12.6 mm, 15.0 mm, and 16.3 mm) versus *E. coli* ATCC25922, *Salmonella* sp, and *S.**aureus* ATCCBAA977. *S. globosus* at 40oC did not show antibacterial activity against all test bacteria

**Impact of various** **pH condition on antibacterial efficacy of *S. globosus***

*S. globosus* produce antibacterial activity against *S. aureus* ATCCBAA977 at pH 6, against *E. coli* ATCC25922, and *S. aureus* ATCCBAA977 at pH 6.5, and against all test bacterial isolate except *P. earuginosa* ATCC27853 at pH 7.Figure 2 shows that the antibacterial activity of S. globosus against all test bacteria increased by increasing the fermentation medium's pH values. The highest antibacterial activity was at pH 8.5, and it was decreased above. It was noticed that *S. globosus* produced antibacterial activity against all test isolates and the highest was against *S. aureus* ATCCBAA977. The diameter of the inhibition zone was (20.3 mm) and decreased slowly above.



**Figure 2.** Impact of various pH efficacy of *S. globosus.*condition on antibacteria

**Effect of shaking incubator on antibacterial activity of *S. globosus***

Results in Figure 3 show the difference between the antibacterial activity produced by *S. globosus* when the cultures of this isolate were incubated in a constant and shaking incubator. *S. globosus* can't have antibacterial activity agents against all the test bacteria when incubated in a shaking incubator. Different antibacterial activities were produced when cultures of *S. globosus* were incubated in a constant incubator.

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**Figure 3.** Effect of shaking incubator on antibacterial activity of *S. globosus.*

The diameters of inhibition zones were (15.3 mm, 14.0 mm, 13.3 mm, 14.3 mm, 24.0 mm, and 15.3 mm) against *E. coli* ATCC35218, *E. coli* ATCC25922, *Salmonella* sp., *P. earuginosa* ATCC27853, *S. aureus* ATCCBAA977 and *St. pneumonia* ATCC49619, respectively.

**Effect of different organic solvents used in the extraction of the antibacterial compound from *S. globosus* on its antibacterial activity.**

The results in Figures 10 revealed that all organic solvents succeeded in extracting the antibacterial compound. Better extraction was obtained using chloroform. The diameters of inhibition zones were (17.6 mm, 16.3 mm, 17.3 mm, 15.6 mm, 27.6 mm,and 17.3 mm) against *E. coli* ATCC35218, *E. coli* ATCC25922, *Salmonella* sp., *P. earuginosa* ATCC27853, *S. aureus* ATCCBAA977 and *St. pneumonia* ATCC49619, respectively.

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**Figure 4.** Effect of different organic solvents used to extract the antibacterial compound from *S. globosus* on its antibacterial activity**.**

**MIC values of the chloroform extract against nosocomial infection bacteria**

The MIC values against all the tested bacterial showed different results. *S. aureus* ATCCBAA977 has been inhibited at (16 µg/m), which was the lowest concentration for inhibition*. E. coli* ATCC35218, *Salmonella* sp. and *St.* pneumoniaATCC49619 bacteria have been inhibited at (32 µg/ml). *E. coli*ATCC25922 inhibited at (64 µg/ml). The highest concentration for inhibition was at (125 µg/ml) against *P. earuginosa* ATCC27853 (table2).

**Table2:** MIC values of the chloroform extract against nosocomial infection bacteria.

|  |  |
| --- | --- |
| Nosocomial infection bacteria | Minimum inhibitory concentration (MIC) |
| *E. coli ATCC35218* | *E. coli ATCC25922* | *Salmonella sp.* | *P. earuginosa ATCC27853* | *S. aureus ATCCBAA977* | *St. pneumonia ATCC49619* |
| Chloroform | 32 | 64 | 32 | 125 | 16 | 32 |

**Discussion**

In this work, *Streptomyces sp.* was dominantly separated from the soil specimen on starch-nitrate medium. This finding is congruent with earlier studies, which reported that the percentage of recovery of *Streptomyces sp.* with antimicrobial activity was higher than other actinobacterial genera (Kitouni et al., 2005). Rare actinobacteria are also important sources in discovering novel antibiotics (Tiwari and Gupta 2012).

This study aimed to optimize the culture condition, which will yield the most optimum antimicrobial efficacy of *Streptomyces sp*. The incubation temperature can influence the bacterial growth and the development of biosynthetic pathway of the metabolites (Stanbury, 2016). Also Pandey *et al.* (2005); Vijayakumar et al.(2012) [19-20] showed that for the optimum production of antibiotics certain carbon and nitrogen sources are required study done by Vasavada *et al.* (2006) showed that the use of different media, pH, salinity, and carbon and nitrogen affect the growth and antibiotic production by Actinomycetes.

Results of the impact of incubation time and temperature on the optimum efficacy of *S. globosus* showed that the highest antibacterial activity was at 30 ° C after 7 days of incubation. This result similar to that found by Sujatha *et al.* [2005 22] with the highest antimicrobial activity at 30 ° C. On the other hand, Ripa *et al.* (2009) reported that isolated *Streptomyces sp.* produced high antibiotic levels when incubated at 39 °C.

This study showed that the best to produce antibacterial activity was under alkaline conditions (pH 8.5)*.* A similar results had been reported earlier by Basilio *et al.* [2003 24]. Ripa *et al.* (2009) showed that the pH 8.0 was optimum for antibiotic produced by *Streptomyces sp*. This may be attributed to the presence of active enzymes for antimicrobial metabolites synthesis at pH 7-9. Each *Streptomyces* strain has an optimum, minimum, and maximum pH at which it gave an optimum enzyme activity (Vijayakumar et al., 2012).

The results of this study showed that the isolate didn't show antibacterial activity at the shaking incubator. Gottschalk *et al.* (2003) in their research using *S. viridosporus* suggested that the rise in the rate of agitation up to 500 rpm contributed to a persistent decline in the growth and development of cells. A rise in the stirring rate may have contributed to high shearing outcomes that were harmful to the solidity of the fungi.

This study showed that all organic solvents (ethyl-alcohol, methanol, chloroform, ethyl-acetate, and acetone) succeeded in extracting the antibacterial compound. Still, the compounds extracted by chloroform had the highest antibacterial activity. Our results were similar to those found by Zakia et al. (2001). Disagree with our results, Vijayakumar *et al.* (2012) confirmed that extraction of *Streptomyces* *sp*. with ethyl acetate produced more active by generating the maximum inhibition versus *P. vulgaris* and minimum inhibition versus *S. aureus*. Besides, Balachandran *et al.* )2012) showed that extraction of *Methylobacterium sp.* With ethyl acetate produced the maximum inhibition versus *K. pneumonia* and minimum inhibition versus *B. subtilis*.

**Conclusion**

These findings indicated that the soil samples of the Al-Baha region have actinomycetes with metabolites that control bacterial pathogens. The extracted metabolites obtained from isolateof our study showed very good antimicrobial activity against four different clinical pathogens. The further structural characterization of these molecules promotes the utilization of this active principle as a lead in case of drug discovery from soil based formulations to eradicate the emerging infectious drug-resistant pathogenic microorganisms.

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